

# Package ‘gscreend’

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**Type** Package

**Title** Analysis of pooled genetic screens

**Version** 1.22.0

**Description** Package for the analysis of pooled genetic screens (e.g. CRISPR-KO). The analysis of such screens is based on the comparison of gRNA abundances before and after a cell proliferation phase. The gscreend packages takes gRNA counts as input and allows detection of genes whose knockout decreases or increases cell proliferation.

**License** GPL-3

**Encoding** UTF-8

**LazyData** false

**Depends** R (>= 3.6)

**Imports** SummarizedExperiment, nloptr, fGarch, methods, BiocParallel, graphics

**Suggests** knitr, testthat, rmarkdown, BiocStyle

**VignetteBuilder** knitr

**RoxygenNote** 7.2.3

**biocViews** Software, StatisticalMethod, PooledScreens, CRISPR

**URL** <https://github.com/imkeller/gscreend>

**BugReports** <https://github.com/imkeller/gscreend/issues>

**git\_url** <https://git.bioconductor.org/packages/gscreend>

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|                |                            |
|----------------|----------------------------|
| assignGeneData | <i>Calculate gene rank</i> |
|----------------|----------------------------|

---

Description

Calculate gene rank

Usage

assignGeneData(object, alpha\_cutoff)

Arguments

- object            PoolScreenExp object
- alpha\_cutoff    alpha cutoff for alpha-RRA (default: 0.05)

Value

object

---

calculateIntervalFits    *Calculate fit parameters for every subset of data*

---

**Description**

Calculate fit parameters for every subset of data

**Usage**

```
calculateIntervalFits(object, quant1, quant2)
```

**Arguments**

|        |                                                                        |
|--------|------------------------------------------------------------------------|
| object | PoolScreenExp object                                                   |
| quant1 | lower quantile for least quantile of squares regression (default: 0.1) |
| quant2 | upper quantile for least quantile of squares regression (default: 0.9) |

**Value**

object

---

calculateLFC    *Calculate log fold changes*

---

**Description**

Calculate log fold changes

**Usage**

```
calculateLFC(object)
```

**Arguments**

|        |                      |
|--------|----------------------|
| object | PoolScreenExp object |
|--------|----------------------|

**Value**

object

---

|                  |                           |
|------------------|---------------------------|
| calculatePValues | <i>Calculate p-values</i> |
|------------------|---------------------------|

---

**Description**

Calculate p-values

**Usage**

```
calculatePValues(object)
```

**Arguments**

|        |                      |
|--------|----------------------|
| object | PoolScreenExp object |
|--------|----------------------|

**Value**

object

---

|                     |                                        |
|---------------------|----------------------------------------|
| createPoolScreenExp | <i>Create PoolScreenExp Experiment</i> |
|---------------------|----------------------------------------|

---

**Description**

Create PoolScreenExp Experiment

**Usage**

```
createPoolScreenExp(data)
```

**Arguments**

|      |                                                                     |
|------|---------------------------------------------------------------------|
| data | Input data object containing gRNA level data (SummarizedExperiment) |
|------|---------------------------------------------------------------------|

**Value**

object PoolScreenExp object

**Examples**

```
raw_counts <- read.table(  
  system.file('extdata', 'simulated_counts.txt',  
    package = 'gscreeend'),  
  header=TRUE)  
  
counts_matrix <- cbind(raw_counts$library0, raw_counts$R0_0, raw_counts$R1_0)  
  
rowData <- data.frame(sgRNA_id = raw_counts$sgrna_id,  
  gene = raw_counts$Gene)  
  
colData <- data.frame(samplename = c('library', 'R1', 'R2'),  
  timepoint = c('T0', 'T1', 'T1'))  
  
library(SummarizedExperiment)  
se <- SummarizedExperiment(assays=list(counts=counts_matrix),  
  rowData=rowData, colData=colData)  
  
# create a PoolScreenExp experiment  
pse <- createPoolScreenExp(se)
```

---

```
createPoolScreenExpFromSE
```

*Create PoolScreenExp Experiment from summarized experiment*

---

**Description**

Create PoolScreenExp Experiment from summarized experiment

**Usage**

```
createPoolScreenExpFromSE(data)
```

**Arguments**

|      |                                                        |
|------|--------------------------------------------------------|
| data | SummarizedExperiment object containing gRNA level data |
|------|--------------------------------------------------------|

**Value**

object

---

```
defineFittingIntervals
```

*Calculate interval limits for splitting data into subsets*

---

### Description

Calculate interval limits for splitting data into subsets

### Usage

```
defineFittingIntervals(object)
```

### Arguments

object                      PoolScreenExp object

### Value

object

---

```
fit_least_quantile
```

*Fit paramters for skew normal distribution*

---

### Description

Fit paramters for skew normal distribution

### Usage

```
fit_least_quantile(LFC, quant1, quant2)
```

### Arguments

LFC                      logarithmic fold changes of gRNA counts  
 quant1                      lower quantile for least quantile of squares regression (default: 0.1)  
 quant2                      upper quantile for least quantile of squares regression (default: 0.9)

### Value

fit\_skewnorm

---

**GeneData***GeneData: set and retrieve GeneData of PoolScreenExp*

---

**Description**

GeneData: set and retrieve GeneData of PoolScreenExp

**Usage**

GeneData(x)

**Arguments**

x                      PoolScreenExp object

**Value**

Gene slot of the object

**Examples**

```
# import a PoolScreenExp object that has been generated using
# RunGscreen()
pse_an <- readRDS(
  system.file('extdata', 'gscreen_analysed_experiment.RData',
    package = 'gscreen'))

GeneData(pse_an)
```

---

**GeneData, PoolScreenExp-method***Accessor function for the Gene slot of the PoolScreenExp class*

---

**Description**

Accessor function for the Gene slot of the PoolScreenExp class

**Usage**

```
## S4 method for signature 'PoolScreenExp'
GeneData(x)
```

**Arguments**

x                      PoolScreenExp object

**Value**

Gene slot of the object

**Examples**

```
# import a PoolScreenExp object that has been generated using
# RunGscreen()
pse_an <- readRDS(
  system.file('extdata', 'gscreen_analysed_experiment.RData',
    package = 'gscreen'))

GeneData(pse_an)
```

---

|                        |                            |
|------------------------|----------------------------|
| normalizePoolScreenExp | <i>Normalize raw count</i> |
|------------------------|----------------------------|

---

**Description**

Normalize raw count

**Usage**

```
normalizePoolScreenExp(object)
```

**Arguments**

object                      PoolScreenExp object

**Value**

object

---

|                     |                                               |
|---------------------|-----------------------------------------------|
| plotModelParameters | <i>Plot model parameters from the fitting</i> |
|---------------------|-----------------------------------------------|

---

**Description**

Plot model parameters from the fitting

**Usage**

```
plotModelParameters(object)
```

**Arguments**

object                      PoolScreenExp object

**Value**

plot

**Examples**

```
# import a PoolScreenExp object that has been generated using
# RunGscreen()
pse_an <- readRDS(
  system.file('extdata', 'gscreen_analysed_experiment.RData',
    package = 'gscreen'))
plotModelParameters(pse_an)
```

---

plotReplicateCorrelation

*Plot replicate correlation*

---

**Description**

Plot replicate correlation

**Usage**

```
plotReplicateCorrelation(object, rep1 = "R1", rep2 = "R2")
```

**Arguments**

object                      PoolScreenExp object  
rep1                        Name of replicate 1  
rep2                        Name of replicate 2

**Value**

replicate\_plot

**Examples**

```
# import a PoolScreenExp object that has been generated using RunGscreen()
pse_an <- readRDS(
  system.file('extdata', 'gscreen_analysed_experiment.RData',
    package = 'gscreen'))
plotReplicateCorrelation(pse_an, rep1 = 'R1', rep2 = 'R2')
```

---

|                     |                                                          |
|---------------------|----------------------------------------------------------|
| PoolScreenExp-class | <i>Class to store pooled CRISPR screening experiment</i> |
|---------------------|----------------------------------------------------------|

---

### Description

The poolScreenExp class is an S4 class used to store sgRNA and gene related data as well as parameters necessary for statistical model.

### Slots

sgRNAData A SummarizedExperiment containing the data related to sgRNAs.

FittingIntervals A vector defining the limits of the intervals used for fitting of null model.

LFCModelParameters A vector of parameters estimated when fitting the null model.

GeneData SummarizedExperiment containing the data related to genes.

FittingOptions A named list with options for fitting: IntervalFraction - fraction of sgRNAs used in every fitting interval (default 0.1), alphaCutoff - alpha cutoff for alpha RRA algorithm (default: 0.05).

---

|              |                                |
|--------------|--------------------------------|
| ResultsTable | <i>Extract a results table</i> |
|--------------|--------------------------------|

---

### Description

Extract a results table

### Usage

```
ResultsTable(object, direction = "negative")
```

### Arguments

|           |                                                                                                               |
|-----------|---------------------------------------------------------------------------------------------------------------|
| object    | PoolScreenExp object                                                                                          |
| direction | Whether the table should contain information on positive or negative fold changes<br>['negative'  'positive'] |

### Value

plot

**Examples**

```
# import a PoolScreenExp object that has been generated using
# RunGscreen()
pse_an <- readRDS(
  system.file('extdata', 'gscreen_analysed_experiment.RData',
    package = 'gscreen'))
ResultsTable(pse_an, direction = 'negative')
```

---

|            |                    |
|------------|--------------------|
| RunGscreen | <i>run gscreen</i> |
|------------|--------------------|

---

**Description**

run gscreen

**Usage**

```
RunGscreen(object, quant1 = 0.1, quant2 = 0.9, alphacutoff = 0.05)
```

**Arguments**

|             |                                                                        |
|-------------|------------------------------------------------------------------------|
| object      | PoolScreenExp object                                                   |
| quant1      | lower quantile for least quantile of squares regression (default: 0.1) |
| quant2      | upper quantile for least quantile of squares regression (default: 0.9) |
| alphacutoff | alpha cutoff for alpha-RRA (default: 0.05)                             |

**Value**

object

**Examples**

```
raw_counts <- read.table(
  system.file('extdata', 'simulated_counts.txt',
    package = 'gscreen'),
  header=TRUE)

# Create the PoolScreenExp to be analyzed
counts_matrix <- cbind(raw_counts$library0, raw_counts$R0_0, raw_counts$R1_0)

rowData <- data.frame(sgRNA_id = raw_counts$sgrna_id,
  gene = raw_counts$Gene)

colData <- data.frame(samplename = c('library', 'R1', 'R2'),
  timepoint = c('T0', 'T1', 'T1'))

library(SummarizedExperiment)
```

```
se <- SummarizedExperiment(assays=list(counts=counts_matrix),
  rowData=rowData, colData=colData)

pse <- createPoolScreenExp(se)

# Run Analysis
pse_an <- RunGscreen(pse)
```

---

**sgRNAData***sgRNAData: set and retrieve sgRNAData of PoolScreenExp*

---

## Description

sgRNAData: set and retrieve sgRNAData of PoolScreenExp

## Usage

```
sgRNAData(x)
```

## Arguments

x                      PoolScreenExp object

## Value

sgRNA slot of the object

## Examples

```
# import a PoolScreenExp object that has been generated using
# RunGscreen()
pse_an <- readRDS(
  system.file('extdata', 'gscreen_analysed_experiment.RData',
    package = 'gscreen'))

sgRNAData(pse_an)
```

---

`sgRNAData, PoolScreenExp-method`*Accessor function for the sgRNA slot of the PoolScreenExp class*

---

**Description**

Accessor function for the sgRNA slot of the PoolScreenExp class

**Usage**

```
## S4 method for signature 'PoolScreenExp'  
sgRNAData(x)
```

**Arguments**

x                      PoolScreenExp object

**Value**

sgRNA slot of the object

**Examples**

```
# import a PoolScreenExp object that has been generated using  
# RunGscreenend()  
pse_an <- readRDS(  
  system.file('extdata', 'gscreenend_analysed_experiment.RData',  
  package = 'gscreenend'))  
  
sgRNAData(pse_an)
```

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