## Package 'mapscape'

June 12, 2025

Title mapscape

Version 1.33.0

**Description** MapScape integrates clonal prevalence, clonal hierarchy, anatomic and mutational information to provide interactive visualization of spatial clonal evolution. There are four inputs to MapScape: (i) the clonal phylogeny, (ii) clonal prevalences, (iii) an image reference, which may be a medical image or drawing and (iv) pixel locations for each sample on the referenced image. Optionally, MapScape can accept a data table of mutations for each clone and their variant allele frequencies in each sample. The output of MapScape consists of a cropped anatomical image surrounded by two representations of each tumour sample. The first, a cellular aggregate, visually displays the prevalence of each clone. The second shows a skeleton of the clonal phylogeny while highlighting only those clones present in the sample. Together, these representations enable the analyst to visualize the distribution of clones throughout anatomic space.

**Depends** R (>= 3.3)

**Imports** htmlwidgets (>= 0.5), jsonlite (>= 0.9.19), base64enc (>= 0.1-3), stringr (>= 1.0.0)

License GPL-3

LazyData true

RoxygenNote 6.0.1

biocViews Visualization

Suggests knitr, rmarkdown

VignetteBuilder knitr

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Author Maia Smith [aut, cre]

Maintainer Maia Smith <maiaannesmith@gmail.com>

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MapScape

#### Description

MapScape is a visualization tool for spatial clonal evolution. MapScape displays a cropped anatomical image surrounded by two representations of each tumour sample representing the distribution of clones throughout anatomic space. The first, a cellular aggregate or donut view, displays the prevalence of each clone. The second shows a skeleton of the patient's clonal phylogeny while highlighting only those clones present in the sample. Note: the cellular aggregate does not accurately represent the positions of clones within a sample. We therefore provide the alternative donut chart view as a less artistic representation of the tumour sample. See the Interactivity section below for instructions to switch between views.

#### Usage

```
mapscape(clonal_prev, tree_edges, sample_locations, img_ref,
    clone_colours = "NA", mutations = "NA", sample_ids = c("NA"),
    n_cells = 100, show_low_prev_gtypes = FALSE,
    phylogeny_title = "Clonal Phylogeny", anatomy_title = "Anatomy",
    classification_title = "Phylogenetic Classification",
    show_warnings = TRUE, width = 960, height = 960)
```

#### Arguments

clonal_prev	data.frame Clonal prevalence. Required columns are:
	<pre>sample_id: character() id for the tumour sample.</pre>
	<pre>clone_id: character() clone id.</pre>
	<pre>clonal_prev: numeric() clonal prevalence.</pre>
tree_edges	data.frame Tree edges of a rooted tree. Required columns are:
	<pre>source: character() source node id.</pre>
	target: character() target node id.
sample_location	15
	data.frame Anatomic locations for each tumour sample. Required columns are:
	<pre>sample_id: character() id for the tumour sample.</pre>
	<b>location_id:</b> character() name of anatomic location for this tumour sample.
	<b>x:</b> numeric() x-coordinate (in pixels) for anatomic location on anatomic image.
	y: numeric() y-coordinate (in pixels) for anatomic location on anatomic image.

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img_ref	character() A reference for the custom anatomical image to use, *** in PNG format ***, either a URL to an image hosted online or a path to the image in local file system.
clone_colours	data.frame (Optional) Clone ids and their corresponding colours (in hex for- mat). Required columns are:
	<pre>clone_id: character() clone id.</pre>
	colour: character() the corresponding Hex colour for each clone id.
mutations	data.frame (Optional) Mutations occurring at each clone. Any additional field will be shown in the mutation table. Required columns are:
	chrom: character() chromosome number.
	coord: numeric() coordinate of mutation on chromosome.
	<pre>clone_id: character() clone id.</pre>
	<pre>sample_id: character() id for the tumour sample.</pre>
	<b>VAF:</b> numeric() ariant allele frequency of the mutation in the corresponding sample.
sample_ids	vector (Optional) Ids of the samples in the order your wish to display them (clockwise from positive x-axis).
n_cells	numeric() (Optional) The number of cells to plot (for voronoi tessellation).
show_low_prev_	gtypes
	logical() (Optional) Whether or not to show low-prevalence (< 0.01) clones in the view. Default is FALSE.
phylogeny_titl	
	character() (Optional) Legend title for the phylogeny. Default is "Clonal Phylogeny".
anatomy_title	character() (Optional) Legend title for the anatomy. Default is "Anatomy".
classification	
	character() (Optional) Legend title for the phylogenetic classification. De- fault is "Phylogenetic Classification".
show_warnings	logical() (Optional) Whether or not to show any warnings. Default is TRUE.
width	numeric() (Optional) Width of the plot. Minimum width is 930.
height	numeric() (Optional) Height of the plot. Minimum height is 700.
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#### Details

Interactive components in the top toolbar:

- 1. Click the download buttons to download the current view as PNG or SVG.
- 2. Click the reset button to exit a clone or mutation selection.
- 3. Click the view switch button to switch between cellular

Interactive components in main view:

- 1. Reorder samples by grabbing the sample name or cellular aggregate / donut and dragging it radially.
- 2. Hover over anatomic location of interest to view the anatomic location name and the patient data associated with that location.
- 3. Hover over a tree node of a particular sample to view cellular prevalence of that clone in that particular sample.

Interactive components in legend:

- 1. Hover over legend tree node to view the clone ID as well as the clone's prevalence at each tumour sample. Any anatomic locations expressing that clone will be highlighted.
- 2. Hover over legend tree branch to view tumour samples expressing all descendant clones.
- 3. Click on legend tree node(s) to view (a) updated mutations table showing novel mutations at that clone(s), and (b) tumour samples expressing the novel mutations at that clone(s).
- 4. Hover over a mixture class (e.g. "pure", "polyphyletic", "monophyletic") to view corresponding tumour samples, and the participating phylogeny in each tumour sample.

Interactive components in mutation table:

- 1. Search for any chromosome, coordinate, gene, etc.
- 2. Click on a row in the table, and the view will update to show the tumour samples with that mutation, and the variant allele frequency for that mutation in each tumour sample.
- 3. Sort the table by a column (all columns sortable except the Clone column).

Note: Click on the reset button to exit a selection. Click the download buttons to download a PNG or SVG of the view.

#### Examples

```
library("mapscape")
# EXAMPLE 1 - Patient A21, Gundem et al., 2015
# clonal prevalences
clonal_prev <- read.csv(system.file("extdata", "A21_clonal_prev.csv",</pre>
    package = "mapscape"))
# mutations
mutations <- read.csv(system.file("extdata", "A21_mutations.csv",</pre>
    package = "mapscape"))
# locations of each tumour sample on user-provided image
sample_locations <- read.csv(system.file("extdata",</pre>
    "A21_sample_locations.csv", package = "mapscape"))
# genotype tree edges
tree_edges <- read.csv(system.file("extdata", "A21_tree.csv", package</pre>
    = "mapscape"))
# image reference
img_ref <- system.file("extdata", "A21_anatomical_image.png", package</pre>
    = "mapscape")
# radial order of samples
sample_ids <- c("H", "F", "J", "D", "A", "I", "C", "E", "G")</pre>
# run mapscape
mapscape(clonal_prev = clonal_prev, tree_edges = tree_edges,
    sample_locations = sample_locations, mutations = mutations,
img_ref = img_ref, sample_ids = sample_ids)
```

#### mapscape

```
# EXAMPLE 2 - Patient 1, McPherson and Roth et al., 2016
# clonal prevalences
clonal_prev <- read.csv(system.file("extdata", "px1_clonal_prev.csv",</pre>
    package = "mapscape"))
# mutations
mutations <- read.csv(system.file("extdata", "px1_mutations.csv",</pre>
    package = "mapscape"))
# locations of each tumour sample on user-provided image
sample_locations <- read.csv(system.file("extdata",</pre>
    "px1_sample_locations.csv", package = "mapscape"))
# genotype tree edges
tree_edges <- read.csv(system.file("extdata", "px1_tree.csv",</pre>
    package = "mapscape"))
# image reference
img_ref <- system.file("extdata", "px1_anatomical_image.png",</pre>
    package = "mapscape")
# colours for each clone
clone_colours <- data.frame(</pre>
    clone_id = c("A", "B", "C", "D", "E", "F", "G", "H", "I"),
    colour = c("d0ced0", "2CD0AB", "7FE9D1", "FFD94B", "FD8EE5",
"F8766D", "4FD8FF", "B09AF5", "D4C7FC"))
# radial order of samples
sample_ids <- c("LFTB4", "LOvB2", "ApC1", "ROvA4", "ROv4", "ROv3",</pre>
    "ROv2", "ROv1", "RFTA16", "Om1", "SBwl", "SBwlE4")
# run mapscape
mapscape(clonal_prev = clonal_prev, tree_edges = tree_edges,
    sample_locations = sample_locations, mutations = mutations,
    img_ref = img_ref, clone_colours = clone_colours,
    sample_ids = sample_ids)
# EXAMPLE 3 - Patient 7, McPherson and Roth et al., 2016
# clonal prevalences
clonal_prev <- read.csv(system.file("extdata", "px7_clonal_prev.csv",</pre>
    package = "mapscape"))
# mutations
mutations <- read.csv(system.file("extdata", "px7_mutations.csv",</pre>
    package = "mapscape"))
# locations of each tumour sample on user-provided image
sample_locations <- read.csv(system.file("extdata",</pre>
    "px7_sample_locations.csv", package = "mapscape"))
# genotype tree edges
tree_edges <- read.csv(system.file("extdata", "px7_tree.csv",</pre>
    package = "mapscape"))
# image reference
```

mapscapeOutput Widget output function for use in Shiny

#### Description

Widget output function for use in Shiny Widget render function for use in Shiny

#### Usage

```
mapscapeOutput(outputId, width = "100%", height = "400px")
renderMapscape(expr, env = parent.frame(), quoted = FALSE)
```

#### Arguments

outputId	– id of output
width	- width of output
height	- height of output
expr	- expression for Shiny
env	- environment for Shiny
quoted	- default is FALSE

#### Examples

mapscapeOutput(1, '100%', '300px')
mapscapeOutput(1, '80%', '300px')
mapscapeOutput("hi")
mapscapeOutput("hello")

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