

Package ‘destiny’

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Type Package

Title Creates diffusion maps

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Description Create and plot diffusion maps.

License GPL-3

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<https://www.helmholtz-muenchen.de/icb/destiny>,
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LinkingTo Rcpp, RcppEigen, grDevices

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Collate 'RcppExports.R' 'aaa.r' 'accessor-generics.r' 'censoring.r' 'colorlegend.r' 'cube_helix.r' 'dataset-helpers.r' 'destiny-package.r' 's4-unions.r' 'dist-matrix-coerce.r' 'sigmas.r' 'diffusionmap.r' 'diffusionmap-methods-accession.r' 'diffusionmap-methods.r' 'plotheelpers.r' 'diffusionmap-plotting.r' 'dpt-branching.r' 'dpt-helpers.r' 'dpt.r' 'dpt-methods-matrix.r' 'dpt-methods.r' 'utils.r' 'dpt-plotting.r' 'eig_decomp.r' 'expressionset-helpers.r' 'find_dm_k.r' 'gene-relevance.r' 'gene-relevance-methods.r' 'gene-relevance-plotting-differential-map.r' 'gene-relevance-plotting-gr-map.r' 'gene-relevance-plotting-rank.r' 'gene-relevance-plotting.r' 'guo-data.r' 'knn.r' 'l_which.r' 'methods-coercion.r' 'methods-extraction.r' 'methods-update.r' 'predict.r' 'projection-dist.r' 'rankcor.r' 'sigmas-plotting.r'

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Coercion methods	<i>Coercion methods</i>
------------------	-------------------------

Description

Convert a [DiffusionMap](#) or [DPT](#) object to other classes

Usage

```
## S4 method for signature 'DiffusionMap'
as.data.frame(x, row.names = NULL, optional = FALSE, ...)
```

```
fortify.DiffusionMap(model, data, ...)
```

```
## S4 method for signature 'DPT'
as.data.frame(x, row.names = NULL, optional = FALSE, ...)
```

```
fortify.DPT(model, data, ...)
```

```
## S4 method for signature 'DPT'
as.matrix(x, ...)
```

Arguments

<code>x, model</code>	A DiffusionMap or DPT object
<code>row.names</code>	NULL or a character vector giving the row names for the data frame. Missing values are not allowed.
<code>optional</code>	logical. If TRUE, setting row names and converting column names (to syntactic names: see <code>make.names</code>) is optional.
<code>...</code>	Passed to as.data.frame
<code>data</code>	ignored

Details

[fortify](#) is a ggplot2 generic allowing a diffusion map to be used as data parameter in [ggplot](#) or [qplot](#).

Value

An object of the desired class

See Also

[DiffusionMap accession methods](#), [Extraction methods](#), [DiffusionMap methods](#) for more

Examples

```
library(Biobase)
data(guo)
dm <- DiffusionMap(guo)
classes <- vapply(as.data.frame(dm), class, character(1L))
stopifnot(all(classes[paste0('DC', 1:20)] == 'numeric'))
stopifnot(all(classes[featureNames(guo)] == 'numeric'))
stopifnot(all(classes[varLabels(guo)] == c('factor', 'integer')))
```

colorlegend

Color legend

Description

Creates a color legend for a vector used to color a plot. It will use the current [palette\(\)](#) or the specified `pal` as reference.

Usage

```

colorlegend(
  col,
  pal = palette(),
  log = FALSE,
  posx = c(0.9, 0.93),
  posy = c(0.05, 0.9),
  main = NULL,
  cex_main = par("cex.sub"),
  cex_axis = par("cex.axis"),
  col_main = par("col.sub"),
  col_lab = par("col.lab"),
  steps = 5,
  steps_color = 100,
  digit = 2,
  left = FALSE,
  ...,
  cex.main = NULL,
  cex.axis = NULL,
  col.main = NULL,
  col.lab = NULL
)

```

Arguments

<code>col</code>	Vector of factor, integer, or double used to determine the ticks.
<code>pal</code>	If <code>col</code> is double, <code>pal</code> is used as a continuous palette, else as categorical one
<code>log</code>	Use logarithmic scale?
<code>posx</code>	Left and right borders of the color bar relative to plot area (Vector of length 2; 0-1)
<code>posy</code>	Bottom and top borders of color bar relative to plot area (Vector of length 2; 0-1)
<code>main</code>	Legend title
<code>cex_main</code>	Size of legend title font (default: subtitle font size <code>par('cex.sub')</code>)
<code>cex_axis</code>	Size of ticks/category labels (default: axis font size <code>par('cex.axis')</code>)
<code>col_main</code>	Color of legend title (default: subtitle color <code>par('col.sub')</code>)
<code>col_lab</code>	Color of tick or category labels (default: axis color <code>par('col.lab')</code>)
<code>steps</code>	Number of labels in case of a continuous axis. If 0 or FALSE, draw no ticks
<code>steps_color</code>	Number of gradient samples in case of continuous axis
<code>digit</code>	Number of digits for continuous axis labels
<code>left</code>	logical. If TRUE, invert <code>posx</code>
<code>...</code>	Additional parameters for the <code>text</code> call used for labels
<code>cex.main, cex.axis, col.main, col.lab</code>	For compatibility with <code>par</code>

Details

When passed a factor or integer vector, it will create a discrete legend, whereas a double vector will result in a continuous bar.

Value

This function is called for the side effect of adding a colorbar to a plot and returns nothing/NULL.

Examples

```
color_data <- 1:6
par(mar = par('mar') + c(0, 0, 0, 3))
plot(sample(6), col = color_data)
colorlegend(color_data)
```

cube_helix

Sequential color palette using the cube helix system

Description

Creates a perceptually monotonously decreasing (or increasing) lightness color palette with different tones. This was necessary in pre-*viridis* times, by now you can probably just use [hcl.colors](#)

Usage

```
cube_helix(
  n = 6,
  start = 0,
  r = 0.4,
  hue = 0.8,
  gamma = 1,
  light = 0.85,
  dark = 0.15,
  reverse = FALSE
)

scale_colour_cube_helix(
  ...,
  start = 0,
  r = 0.4,
  hue = 0.8,
  gamma = 1,
  light = 0.85,
  dark = 0.15,
  reverse = FALSE,
  discrete = TRUE,
```

```

    guide = if (discrete) "legend" else "colourbar"
  )

scale_color_cube_helix(
  ...,
  start = 0,
  r = 0.4,
  hue = 0.8,
  gamma = 1,
  light = 0.85,
  dark = 0.15,
  reverse = FALSE,
  discrete = TRUE,
  guide = if (discrete) "legend" else "colourbar"
)

scale_fill_cube_helix(
  ...,
  start = 0,
  r = 0.4,
  hue = 0.8,
  gamma = 1,
  light = 0.85,
  dark = 0.15,
  reverse = FALSE,
  discrete = TRUE,
  guide = if (discrete) "legend" else "colourbar"
)

```

Arguments

<code>n</code>	Number of colors to return (default: 6)
<code>start</code>	Hue to start helix at ($\text{start} \in [0, 3]$, default: 0)
<code>r</code>	Number of rotations of the helix. Can be negative. (default: 0.4)
<code>hue</code>	Saturation. 0 means greyscale, 1 fully saturated colors (default: 0.8)
<code>gamma</code>	Emphasize darker ($\text{gamma} < 1$) or lighter ($\text{gamma} > 1$) colors (default: 1)
<code>light</code>	Lightest lightness (default: 0.85)
<code>dark</code>	Darkest lightness (default: 0.15)
<code>reverse</code>	logical. If TRUE, reverse lightness (default: FALSE)
<code>...</code>	parameters passed to discrete_scale or continuous_scale
<code>discrete</code>	If TRUE, return a discrete scale, if FALSE a continuous one (default: TRUE)
<code>guide</code>	Type of scale guide to use. See guides

Value

A character vector of hex colors with length `n`

Examples

```
palette(cube_helix())
image(matrix(1:6), col = 1:6, pch = 19, axes = FALSE)

cr <- scales::colour_ramp(cube_helix(12, r = 3))
r <- runif(100)
plot(1:100, r, col = cr(r), type = 'b', pch = 20)
```

destiny

Create and plot diffusion maps

Description

The main function is `DiffusionMap`, which returns an object you can `plot` (`plot.DiffusionMap` is then called).

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See Also

Useful links:

- <https://theislab.github.io/destiny/>
- <https://github.com/theislab/destiny/>
- <https://www.helmholtz-muenchen.de/icb/destiny>
- <https://bioconductor.org/packages/destiny>
- [doi:10.1093/bioinformatics/btv715](https://doi.org/10.1093/bioinformatics/btv715)
- Report bugs at <https://github.com/theislab/destiny/issues>

Examples

```
demo(destiny, ask = FALSE)
```

destiny generics	<i>destiny generics</i>
------------------	-------------------------

Description

destiny provides several generic methods and implements them for the [DiffusionMap](#) and [Sigmas](#) classes.

Usage

```
eigenvalues(object)

eigenvalues(object) <- value

eigenvectors(object)

eigenvectors(object) <- value

sigmas(object)

sigmas(object) <- value

dataset(object)

dataset(object) <- value

distance(object)

distance(object) <- value

optimal_sigma(object)
```

Arguments

object	Object from which to extract or to which to assign a value
value	Value to assign within an object

Value

`eigenvalues` retrieves the numeric eigenvalues
`eigenvectors` retrieves the eigenvectors matrix
`sigmas` retrieves the [Sigmas](#) from an object utilizing it as kernel width
`dataset` retrieves the data the object was created from
`distance` retrieves the distance metric used to create the object, e.g. euclidean
`optimal_sigma` retrieves the numeric value of the optimal sigma or local sigmas

See Also

[DiffusionMap methods](#) and [Sigmas](#) class for implementations

Examples

```
data(guo_norm)
dm <- DiffusionMap(guo_norm)
eigenvalues(dm)
eigenvectors(dm)
sigmas(dm)
optimal_sigma(dm)
dataset(dm)
distance(dm)
```

DiffusionMap accession methods

DiffusionMap accession methods

Description

Get and set eigenvalues, eigenvectors, and sigma(s) of a [DiffusionMap](#) object.

Usage

```
## S4 method for signature 'DiffusionMap'
eigenvalues(object)

## S4 replacement method for signature 'DiffusionMap'
eigenvalues(object) <- value

## S4 method for signature 'DiffusionMap'
eigenvectors(object)

## S4 replacement method for signature 'DiffusionMap'
eigenvectors(object) <- value

## S4 method for signature 'DiffusionMap'
sigmas(object)

## S4 replacement method for signature 'DiffusionMap'
sigmas(object) <- value

## S4 method for signature 'DiffusionMap'
dataset(object)

## S4 replacement method for signature 'DiffusionMap'
```

```
dataset(object) <- value

## S4 method for signature 'DiffusionMap'
distance(object)

## S4 replacement method for signature 'DiffusionMap'
distance(object) <- value

## S4 method for signature 'DiffusionMap'
optimal_sigma(object)
```

Arguments

object	A DiffusionMap
value	Vector of eigenvalues or matrix of eigenvectors to get/set

Value

The assigned or retrieved value

See Also

[Extraction methods](#), [DiffusionMap methods](#), [Coercion methods](#) for more

Examples

```
data(guo)
dm <- DiffusionMap(guo)
eigenvalues(dm)
eigenvectors(dm)
sigmas(dm)
dataset(dm)
optimal_sigma(dm)
```

DiffusionMap methods *DiffusionMap methods*

Description

Methods for external operations on diffusion maps

Usage

```
## S4 method for signature 'DiffusionMap'
print(x)

## S4 method for signature 'DiffusionMap'
show(object)
```

Arguments

x, object A [DiffusionMap](#)

Value

The DiffusionMap object (print), or NULL (show), invisibly

See Also

[DiffusionMap accession methods](#), [Extraction methods](#), [Coercion methods](#) for more

Examples

```
data(guo)
dm <- DiffusionMap(guo)
print(dm)
show(dm)
```

DiffusionMap-class *Create a diffusion map of cells*

Description

The provided data can be a double [matrix](#) of expression data or a [data.frame](#) with all non-integer (double) columns being treated as expression data features (and the others ignored), an [Expression-Set](#), or a [SingleCellExperiment](#).

Usage

```
DiffusionMap(
  data = stopifnot_distmatrix(distance),
  sigma = "local",
  k = find_dm_k(dataset_n_observations(data, distance) - 1L),
  n_eigs = min(20L, dataset_n_observations(data, distance) - 2L),
  density_norm = TRUE,
  ...,
  distance = c("euclidean", "cosine", "rankcor", "l2"),
  n_pcs = NULL,
  n_local = seq(to = min(k, 7L), length.out = min(k, 3L)),
  rotate = FALSE,
  censor_val = NULL,
  censor_range = NULL,
  missing_range = NULL,
  vars = NULL,
  knn_params = list(),
  verbose = !is.null(censor_range),
  suppress_dpt = FALSE
)
```

Arguments

data	Expression data to be analyzed and covariates. Provide vars to select specific columns other than the default: all double value columns. If distance is a distance matrix, data has to be a data.frame with covariates only.
sigma	Diffusion scale parameter of the Gaussian kernel. One of 'local', 'global', a (numeric) global sigma or a Sigmas object. When choosing 'global', a global sigma will be calculated using find_sigmas . (Optional. default: 'local') A larger sigma might be necessary if the eigenvalues can not be found because of a singularity in the matrix
k	Number of nearest neighbors to consider (default: a guess between 100 and $n - 1$. See find_dm_k).
n_eigs	Number of eigenvectors/values to return (default: 20)
density_norm	logical. If TRUE, use density normalisation
...	Unused. All parameters to the right of the ... have to be specified by name (e.g. <code>DiffusionMap(data, distance = 'cosine')</code>)
distance	Distance measurement method applied to data or a distance matrix/ dist . For the allowed values, see find_knn . If this is a sparseMatrix , zeros are interpreted as "not a close neighbors", which allows the use of kNN-sparsified matrices (see the return value of find_knn).
n_pcs	Number of principal components to compute to base calculations on. Using e.g. 50 PCs results in more regular looking diffusion maps. The default NULL will not compute principal components, but use <code>reducedDims(data, 'pca')</code> if present. Set to NA to suppress using PCs.
n_local	If <code>sigma == 'local'</code> , the <code>n_local</code> th nearest neighbor(s) determine(s) the local sigma
rotate	logical. If TRUE, rotate the eigenvalues to get a slimmer diffusion map
censor_val	Value regarded as uncertain. Either a single value or one for every dimension (Optional, default: <code>censor_val</code>)
censor_range	Uncertainty range for censoring (Optional, default: none). A length-2-vector of certainty range start and end. TODO: also allow $2 \times G$ matrix
missing_range	Whole data range for missing value model. Has to be specified if NAs are in the data
vars	Variables (columns) of the data to use. Specifying NULL will select all columns (default: All floating point value columns)
knn_params	Parameters passed to find_knn
verbose	Show a progressbar and other progress information (default: do it if censoring is enabled)
suppress_dpt	Specify TRUE to skip calculation of necessary (but spacious) information for DPT in the returned object (default: FALSE)

Value

A DiffusionMap object:

Slots

eigenvalues Eigenvalues ranking the eigenvectors
 eigenvectors Eigenvectors mapping the datapoints to n_eigs dimensions
 sigmas [Sigmas](#) object with either information about the [find_sigmas](#) heuristic run or just local or [optimal_sigma](#).
 data_env Environment referencing the data used to create the diffusion map
 eigenvect0 First (constant) eigenvector not included as diffusion component.
 transitions Transition probabilities. Can be NULL
 d Density vector of transition probability matrix
 d_norm Density vector of normalized transition probability matrix
 k The k parameter for kNN
 n_pcs Number of principal components used in kNN computation (NA if raw data was used)
 n_local The n_localth nearest neighbor(s) is/are used to determine local kernel density
 density_norm Was density normalization used?
 rotate Were the eigenvectors rotated?
 distance Distance measurement method used
 censor_val Censoring value
 censor_range Censoring range
 missing_range Whole data range for missing value model
 vars Vars parameter used to extract the part of the data used for diffusion map creation
 knn_params Parameters passed to [find_knn](#)

See Also

[DiffusionMap methods](#) to get and set the slots. [find_sigmas](#) to pre-calculate a fitting global sigma parameter

Examples

```

data(guo)
DiffusionMap(guo)
DiffusionMap(guo, 13, censor_val = 15, censor_range = c(15, 40), verbose = TRUE)

covars <- data.frame(covar1 = letters[1:100])
dists <- dist(matrix(rnorm(100*10), 100))
DiffusionMap(covars, distance = dists)

```

dm_predict	<i>Predict new data points using an existing DiffusionMap. The resulting matrix can be used in the plot method for the DiffusionMap</i>
------------	---

Description

Predict new data points using an existing DiffusionMap. The resulting matrix can be used in [the plot method for the DiffusionMap](#)

Usage

```
dm_predict(dm, new_data, ..., verbose = FALSE)
```

Arguments

dm	A DiffusionMap object.
new_data	New data points to project into the diffusion map. Can be a matrix , data.frame , ExpressionSet , or SingleCellExperiment .
...	Passed to proxy::dist (new_data, data, dm@distance, ...).
verbose	Show progress messages?

Value

A $nrow(new_data) \times ncol(eigenvectors(dif))$ matrix of projected diffusion components for the new data.

Examples

```
data(guo)
g1 <- guo[, guo$num_cells != 32L]
g2 <- guo[, guo$num_cells == 32L]
dm <- DiffusionMap(g1)
dc2 <- dm_predict(dm, g2)
plot(dm, new_dcs = dc2)
```

DPT matrix methods	<i>DPT Matrix methods</i>
--------------------	---------------------------

Description

Treat DPT object as a matrix of cell-by-cell DPT distances.

Usage

```
## S4 method for signature 'DPT,index,index,logicalOrMissing'
x[i, j, ..., drop = TRUE]

## S4 method for signature 'DPT,index,missing,logicalOrMissing'
x[i, j, ..., drop = TRUE]

## S4 method for signature 'DPT,missing,index,logicalOrMissing'
x[i, j, ..., drop = TRUE]

## S4 method for signature 'DPT,missing,missing,logicalOrMissing'
x[i, j, ..., drop = TRUE]

## S4 method for signature 'DPT,index,index'
x[[i, j, ...]]

## S4 method for signature 'DPT'
nrow(x)

## S4 method for signature 'DPT'
ncol(x)

## S4 method for signature 'DPT'
dim(x)
```

Arguments

x	DPT object.
i, j	Numeric or logical index.
...	ignored
drop	If TRUE, coerce result to a vector if it would otherwise have 1 %in% dim(result).

Value

[returns a dense matrix or (if applicable and isTRUE(drop)) a vector.
 [[returns single distance value
 nrow and ncol return the number of cells
 dim returns c(n_cells, n_cells)

See Also

[as.matrix.DPT](#)

Examples

```
data(guo_norm)
dm <- DiffusionMap(guo_norm)
```

```
dpt <- DPT(dm)
set.seed(1)
plot(dpt[random_root(dpt), ], Biobase::exprs(guo_norm)['DppaI', ])
```

DPT methods

DPT methods

Description

Methods for the [DPT](#) class. `branch_divide` subdivides branches for plotting (see the examples).

Usage

```
branch_divide(dpt, divide = integer(0L))

tips(dpt)

## S4 method for signature 'DPT'
dataset(object)

## S4 replacement method for signature 'DPT'
dataset(object) <- value
```

Arguments

<code>dpt, object</code>	DPT object
<code>divide</code>	Vector of branch numbers to use for division
<code>value</code>	Value of slot to set

Value

`branch_divide` and `dataset<-` return the changed object, `dataset` the extracted data, and `tips` the tip indices.

See Also

[plot.DPT](#) uses `branch_divide` for its `divide` argument.

Examples

```
data(guo_norm)
dpt <- DPT(DiffusionMap(guo_norm))
dpt_9_branches <- branch_divide(dpt, 1:3)
plot(dpt_9_branches, col_by = 'branch')
```

DPT-class

*Diffusion Pseudo Time***Description**

Create pseudotime ordering and assigns cell to one of three branches

Usage

```
DPT(dm, tips = random_root(dm), ..., w_width = 0.1)
```

Arguments

dm	A DiffusionMap object. Its transition probabilities will be used to calculate the DPT
tips	The cell index/indices from which to calculate the DPT(s) (integer of length 1-3)
...	Unused. All parameters to the right of the ... have to be specified by name (e.g. <code>DPT(dm, w_width = 0.2)</code>)
w_width	Window width to use for deciding the branch cutoff

Details

Treat it as a matrix of pseudotime by subsetting (`[ dim nrow ncol as.matrix)`, and as a list of pseudotime, and expression vectors (`$ [[ names as.data.frame)`).

Value

A DPT object:

Slots

branch [matrix](#) (of [integer](#)) recursive branch labels for each cell (row); NA for undecided. Use [branch_divide](#) to modify this.

tips [matrix](#) (of [logical](#)) indicating if a cell (row) is a tip of the corresponding branch level (col)

dm [DiffusionMap](#) used to create this DPT object

Examples

```
data(guo_norm)
dm <- DiffusionMap(guo_norm)
dpt <- DPT(dm)
str(dpt)
```

eig_decomp

*Fast eigen decomposition using [eigs](#)***Description**

By default uses a random initialization vector that you can make deterministic using [set.seed](#) or override by specifying `opts = list(initvec = ...)`.

Usage

```
eig_decomp(m, n_eigs, sym, ..., opts = list())
```

Arguments

<code>m</code>	A matrix (e.g. from the Matrix package) or a function (see eigs).
<code>n_eigs</code>	Number of eigenvectors to return.
<code>sym</code>	defunct and ignored.
<code>...</code>	Passed to eigs .
<code>opts</code>	Passed to eigs .

Value

see [eigs](#).

Examples

```
eig_decomp(cbind(c(1,0,-1), c(0,1,0), c(-1,0,1)), 2)
```

 ExpressionSet helper methods

Convert object to [ExpressionSet](#) or read it from a file

Description

These functions present quick way to create [ExpressionSet](#) objects.

Usage

```
as.ExpressionSet(x, ...)
```

```
## S4 method for signature 'data.frame'
```

```
as.ExpressionSet(x, annotation_cols = !sapply(x, is.double))
```

```
read.ExpressionSet(file, header = TRUE, ...)
```

Arguments

<code>x</code>	data.frame to convert to an ExpressionSet .
<code>...</code>	Additional parameters to read.table
<code>annotation_cols</code>	The data.frame columns used as annotations. All others are used as expressions. (Logical, character or numerical index array)
<code>file</code>	File path to read ASCII data from
<code>header</code>	Specifies if the file has a header row.

Details

They work by using all continuous (double) columns as expression data, and all others as observation annotations.

Value

an [ExpressionSet](#) object

See Also

[read.table](#) on which `read.ExpressionSet` is based, and [ExpressionSet](#).

Examples

```
library(Biobase)
df <- data.frame(Time = seq_len(3), #integer column
                 Actb = c(0.05, 0.3, 0.8),
                 Gapdh = c(0.2, 0.03, 0.1))
set <- as.ExpressionSet(df)
rownames(exprs(set)) == c('Actb', 'Gapdh')
phenoData(set)$Time == 1:3
```

Description

Extract common information from objects. Apart from the input data's branches, you can extract diffusion components via `$DCx`. From [DPT](#) objects, you can also extract the branch label via `$Branch`, or the diffusion pseudo time for a numbered cell via `$DPTx`.

Usage

```
## S4 method for signature 'DiffusionMap'
names(x)

## S4 method for signature 'DPT'
names(x)

## S4 method for signature 'DiffusionMap,character,missing'
x[[i, j, ...]]

## S4 method for signature 'DPT,character,missing'
x[[i, j, ...]]

## S4 method for signature 'DiffusionMap'
x$name

## S4 method for signature 'DPT'
x$name
```

Arguments

x	DiffusionMap or DPT object
i, name	Name of a diffusion component 'DCx', 'DPTx', 'Branch' or column from the data
j	N/A
...	ignored

Value

The names or data row, see respective generics.

See Also

[Extract](#), [names](#) for the generics. [DiffusionMap accession methods](#), [DiffusionMap methods](#), [Coercion methods](#) for more

Examples

```
data(guo)
dm <- DiffusionMap(guo)
dm$DC1      # A diffusion component
dm$Actb     # A gene expression vector
dm$num_cells # Phenotype metadata

dpt <- DPT(dm)
dm$Branch
dm$DPT1
```

find_dm_k	<i>Find a suitable k</i>
-----------	--------------------------

Description

The k parameter for the k nearest neighbors used in [DiffusionMap](#) should be as big as possible while still being computationally feasible. This function approximates it depending on the size of the dataset n.

Usage

```
find_dm_k(n, min_k = 100L, small = 1000L, big = 10000L)
```

Arguments

n	Number of possible neighbors (nrow(dataset) - 1)
min_k	Minimum number of neighbors. Will be chosen for $n \geq big$
small	Number of neighbors considered small. If/where $n \leq small$, n itself will be returned.
big	Number of neighbors considered big. If/where $n \geq big$, min_k will be returned.

Value

A vector of the same length as n that contains suitable k values for the respective n

Examples

```
curve(find_dm_k(n), 0, 13000, xname = 'n')
curve(find_dm_k(n) / n, 0, 13000, xname = 'n')
```

find_knn	<i>kNN search</i>
----------	-------------------

Description

Approximate k nearest neighbor search with flexible distance function.

Usage

```
find_knn(
  data,
  k,
  ...,
  query = NULL,
  distance = c("euclidean", "cosine", "rankcor", "l2"),
  method = c("covertree", "hnsu"),
  sym = TRUE,
  verbose = FALSE
)
```

Arguments

data	Data matrix
k	Number of nearest neighbors
...	Parameters passed to hnsu_knn
query	Query matrix. Leave it out to use data as query
distance	Distance metric to use. Allowed measures: Euclidean distance (default), cosine distance ($1 - \text{corr}(c_1, c_2)$) or rank correlation distance ($1 - \text{corr}(\text{rank}(c_1), \text{rank}(c_2))$)
method	Method to use. 'hnsu' is tunable with ... but generally less exact than 'covertree' (default: 'covertree')
sym	Return a symmetric matrix (as long as query is NULL)?
verbose	Show a progressbar? (default: FALSE)

Value

A [list](#) with the entries:

index A $nrow(data) \times k$ [integer matrix](#) containing the indices of the k nearest neighbors for each cell.

dist A $nrow(data) \times k$ [double matrix](#) containing the distances to the k nearest neighbors for each cell.

dist_mat A [dgCMatrix](#) if sym == TRUE, else a [dsCMatrix](#) ($nrow(query) \times nrow(data)$). Any zero in the matrix (except for the diagonal) indicates that the cells in the corresponding pair are close neighbors.

find_sigmas	<i>Calculate the average dimensionality for m different gaussian kernel widths (σ).</i>
-------------	---

Description

The sigma with the maximum value in average dimensionality is close to the ideal one. Increasing step number gets this nearer to the ideal one.

Usage

```

find_sigmas(
  data,
  step_size = 0.1,
  steps = 10L,
  start = NULL,
  sample_rows = 500L,
  early_exit = FALSE,
  ...,
  censor_val = NULL,
  censor_range = NULL,
  missing_range = NULL,
  vars = NULL,
  verbose = TRUE
)

```

Arguments

data	Data set with n observations. Can be a data.frame , matrix , ExpressionSet or SingleCellExperiment .
step_size	Size of log-sigma steps
steps	Number of steps/calculations
start	Initial value to search from. (Optional. default: $\log_{10}(\min(\text{dist}(\text{data})))$)
sample_rows	Number of random rows to use for sigma estimation or vector of row indices/names to use. In the first case, only used if actually smaller than the number of available rows (Optional. default: 500)
early_exit	logical. If TRUE, return if the first local maximum is found, else keep running
...	Unused. All parameters to the right of the ... have to be specified by name (e.g. <code>find_sigmas(data, verbose = FALSE)</code>)
censor_val	Value regarded as uncertain. Either a single value or one for every dimension
censor_range	Uncertainty range for censoring. A length-2-vector of certainty range start and end. TODO: also allow $2 \times G$ matrix
missing_range	Whole data range for missing value model. Has to be specified if NAs are in the data
vars	Variables (columns) of the data to use. Specifying TRUE will select all columns (default: All floating point value columns)
verbose	logical. If TRUE, show a progress bar and plot the output

Value

Object of class [Sigmas](#)

See Also

[Sigmas](#), the class returned by this; [DiffusionMap](#), the class this is used for

Examples

```
data(guo)
sigs <- find_sigmas(guo, verbose = TRUE)
DiffusionMap(guo, sigs)
```

find_tips

*Find tips in a DiffusionMap object***Description**

Find tips in a DiffusionMap object

Usage

```
find_tips(dm_or_dpt, root = random_root(dm_or_dpt))
```

Arguments

dm_or_dpt	A DiffusionMap or DPT object
root	Root cell index from which to find tips. (default: random)

Value

An integer vector of length 3

Examples

```
data(guo)
dm <- DiffusionMap(guo)
is_tip <- l_which(find_tips(dm), len = ncol(guo))
plot(dm, col = factor(is_tip))
```

Gene Relevance methods

*Gene Relevance methods***Description**

featureNames <- ... can be used to set the gene names used for plotting (e.g. if the data contains hardly readably gene or transcript IDs). dataset gets the expressions used for the gene relevance calculations, and distance the distance measure.

Usage

```
## S4 method for signature 'GeneRelevance'
print(x)

## S4 method for signature 'GeneRelevance'
show(object)

## S4 method for signature 'GeneRelevance'
featureNames(object)

## S4 replacement method for signature 'GeneRelevance,characterOrFactor'
featureNames(object) <- value

## S4 method for signature 'GeneRelevance'
dataset(object)

## S4 replacement method for signature 'GeneRelevance'
dataset(object) <- value

## S4 method for signature 'GeneRelevance'
distance(object)

## S4 replacement method for signature 'GeneRelevance'
distance(object) <- value
```

Arguments

x, object	GeneRelevance object
value	A text vector (character or factor)

Value

dataset, distance, and featureNames return the stored properties. The other methods return a GeneRelevance object (print, ... <- ...), or NULL (show), invisibly

See Also

[gene_relevance](#), [Gene Relevance plotting](#)

Examples

```
data(guo_norm)
dm <- DiffusionMap(guo_norm)
gr <- gene_relevance(dm)
stopifnot(distance(gr) == distance(dm))
featureNames(gr)[[37]] <- 'Id2 (suppresses differentiation)'
# now plot it with the changed gene name(s)
```

GeneRelevance-class	<i>Gene relevances for entire data set</i>
---------------------	--

Description

The relevance map is cached insided of the [DiffusionMap](#).

Usage

```
gene_relevance(  
  coords,  
  exprs,  
  ...,  
  k = 20L,  
  dims = 1:2,  
  distance = NULL,  
  smooth = TRUE,  
  remove_outliers = FALSE,  
  verbose = FALSE  
)  
  
## S4 method for signature 'DiffusionMap,missing'  
gene_relevance(  
  coords,  
  exprs,  
  ...,  
  k = 20L,  
  dims = 1:2,  
  distance = NULL,  
  smooth = TRUE,  
  remove_outliers = FALSE,  
  verbose = FALSE  
)  
  
## S4 method for signature 'matrix,dMatrixOrMatrix'  
gene_relevance(  
  coords,  
  exprs,  
  ...,  
  pcs = NULL,  
  knn_params = list(),  
  weights = 1,  
  k,  
  dims,  
  distance,  
  smooth,  
  remove_outliers,
```

```
    verbose
  )
```

Arguments

<code>coords</code>	A DiffusionMap object or a cells $\times$ dims matrix .
<code>exprs</code>	An cells $\times$ genes matrix . Only provide if <code>coords</code> is no DiffusionMap .
<code>...</code>	Unused. All parameters to the right of the <code>...</code> have to be specified by name.
<code>k</code>	Number of nearest neighbors to use
<code>dims</code>	Index into columns of coord
<code>distance</code>	Distance measure to use for the nearest neighbor search.
<code>smooth</code>	Smoothing parameters <code>c(window, alpha)</code> (see smth.gaussian). Alternatively TRUE to use the smoother defaults or FALSE to skip smoothing,
<code>remove_outliers</code>	Remove cells that are only within one other cell's nearest neighbor, as they tend to get large norms.
<code>verbose</code>	If TRUE , log additional info to the console
<code>pcs</code>	A cell $\times$ n_pcs matrix of principal components to use for the distances.
<code>knn_params</code>	A list of parameters for find_knn .
<code>weights</code>	Weights for the partial derivatives. A vector of the same length as <code>dims</code> .

Value

A `GeneRelevance` object:

Slots

<code>coords</code>	A cells $\times$ dims matrix or sparseMatrix of coordinates (e.g. diffusion components), reduced to the dimensions passed as <code>dims</code>
<code>exprs</code>	A cells $\times$ genes matrix of expressions
<code>partials</code>	Array of partial derivatives wrt to considered dimensions in reduced space (genes $\times$ cells $\times$ dimensions)
<code>partials_norm</code>	Matrix with norm of aforementioned derivatives. (n_genes $\times$ cells)
<code>nn_index</code>	Matrix of k nearest neighbor indices. (cells $\times$ k)
<code>dims</code>	Column index for plotted dimensions. Can character , numeric or logical
<code>distance</code>	Distance measure used in the nearest neighbor search. See find_knn
<code>smooth_window</code>	Smoothing window used (see smth.gaussian)
<code>smooth_alpha</code>	Smoothing kernel width used (see smth.gaussian)

See Also

[Gene Relevance methods](#), [Gene Relevance plotting](#): `plot_differential_map`/`plot_gene_relevance`

Examples

```
data(guo_norm)
dm <- DiffusionMap(guo_norm)
gr <- gene_relevance(dm)

m <- t(Biobase::exprs(guo_norm))
gr_pca <- gene_relevance(prcomp(m)$x, m)
# now plot them!
```

guo

Guo et al. mouse embryonic stem cell qPCR data

Description

Gene expression data of 48 genes and an annotation column \$num_cells containing the cell stage at which the embryos were harvested.

Usage

```
data(guo)
data(guo_norm)
```

Format

An [ExpressionSet](#) with 48 features, 428 observations and 2 [phenoData](#) annotations.

Details

The data is normalized using the mean of two housekeeping genes. The difference between guo and guo_norm is the LoD being set to 10 in the former, making it usable with the censor_val parameter of [DiffusionMap](#).

Value

an [ExpressionSet](#) with 48 features and 428 observations containing qPCR Ct values and a "num.cells" observation annotation.

Author(s)

Guoji Guo, Mikael Huss, Guo Qing Tong, Chaoyang Wang, Li Li Sun, Neil D. Clarke, Paul Robson
<robsonp@gis.a-star.edu.sg>

References

<http://www.sciencedirect.com/science/article/pii/S1534580710001103>

l_which	<i>Logical which</i>
---------	----------------------

Description

Inverse of [which](#). Converts an array of numeric or character indices to a logical index array. This function is useful if you need to perform logical operation on an index array but are only given numeric indices.

Usage

```
l_which(idx, nms = seq_len(len), len = length(nms), useNames = TRUE)
```

Arguments

idx	Numeric or character indices.
nms	Array of names or a sequence. Required if idx is a character array
len	Length of output array. Alternative to nms if idx is numeric
useNames	Use the names of nms or idx

Details

Either nms or len has to be specified.

Value

Logical vector of length len or the same length as nms

Examples

```
all(l_which(2, len = 3L) == c(FALSE, TRUE, FALSE))
all(l_which(c('a', 'c'), letters[1:3]) == c(TRUE, FALSE, TRUE))
```

plot.DiffusionMap	<i>3D or 2D plot of diffusion map</i>
-------------------	---------------------------------------

Description

If you want to plot the eigenvalues, simply `plot(eigenvalues(dm)[start:end], ...)`

Usage

```

plot.DiffusionMap(
  x,
  dims = 1:3,
  new_dcs = if (!is.null(new_data)) dm_predict(x, new_data),
  new_data = NULL,
  col = NULL,
  col_by = NULL,
  col_limits = NULL,
  col_new = "red",
  pal = NULL,
  pal_new = NULL,
  ...,
  ticks = FALSE,
  axes = TRUE,
  box = FALSE,
  legend_main = col_by,
  legend_opts = list(),
  interactive = FALSE,
  draw_legend = !is.null(col_by) || (length(col) > 1 && !is.character(col)),
  consec_col = TRUE,
  col_na = "grey",
  plot_more = function(p, ..., rescale = NULL) p
)

## S4 method for signature 'DiffusionMap,numeric'
plot(x, y, ...)

## S4 method for signature 'DiffusionMap,missing'
plot(x, y, ...)

```

Arguments

x	A DiffusionMap
dims, y	Diffusion components (eigenvectors) to plot (default: first three components; 1:3)
new_dcs	An optional matrix also containing the rows specified with y and plotted. (default: no more points)
new_data	A data set in the same format as x that is used to create <code>new_dcs <- dm_predict(dif, new_data)</code>
col	Single color string or vector of discrete or categoric values to be mapped to colors. E.g. a column of the data matrix used for creation of the diffusion map. (default: cluster_louvain if igraph is installed)
col_by	Specify a <code>dataset(x)</code> or <code>phenoData(dataset(x))</code> column to use as color
col_limits	If col is a continuous (=double) vector, this can be overridden to map the color range differently than from min to max (e.g. specify <code>c(0, 1)</code>)

col_new	If new_dcs is given, it will take on this color. A vector is also possible. (default: red)
pal	Palette used to map the col vector to colors. (default: use hcl.colors for continuous and palette() for discrete data)
pal_new	Palette used to map the col_new vector to colors. (default: see pal argument)
...	Parameters passed to plot , scatterplot3d , or plot3d (if interactive == TRUE)
ticks	logical. If TRUE, show axis ticks (default: FALSE)
axes	logical. If TRUE, draw plot axes (default: Only if ticks is TRUE)
box	logical. If TRUE, draw plot frame (default: TRUE or the same as axes if specified)
legend_main	Title of legend. (default: nothing unless col_by is given)
legend_opts	Other colorlegend options (default: empty list)
interactive	Use plot3d to plot instead of scatterplot3d ?
draw_legend	logical. If TRUE, draw color legend (default: TRUE if col_by is given or col is given and a vector to be mapped)
consec_col	If col or col_by refers to an integer column, with gaps (e.g. <code>c(5, 0, 0, 3)</code>) use the palette color consecutively (e.g. <code>c(3, 1, 1, 2)</code>)
col_na	Color for NA in the data. specify NA to hide.
plot_more	Function that will be called while the plot margins are temporarily changed (its p argument is the rgl or scatterplot3d instance or NULL, its rescale argument is NULL, a list(from = c(a, b), to = c(c, d)), or an array of shape $from to \times dims \times min max$, i.e. $2 \times length(dims) \times 2$. In case of 2d plotting, it should take and return a ggplot2 object.

Details

If you specify negative numbers as diffusion components (e.g. `plot(dm, c(-1, 2))`), then the corresponding components will be flipped.

Value

The return value of the underlying call is returned, i.e. a scatterplot3d or rgl object.

Examples

```
data(guo)
plot(DiffusionMap(guo))
```

plot.DPT

*Plot DPT***Description**

Plots diffusion components from a Diffusion Map and the accompanying Diffusion Pseudo Time ([DPT](#))

Usage

```
plot.DPT(
  x,
  root = NULL,
  paths_to = integer(0L),
  dcs = 1:2,
  divide = integer(0L),
  w_width = 0.1,
  col_by = "dpt",
  col_path = rev(palette()),
  col_tip = "red",
  ...,
  col = NULL,
  legend_main = col_by
)

## S4 method for signature 'DPT,numeric'
plot(x, y, ...)

## S4 method for signature 'DPT,missing'
plot(x, y, ...)
```

Arguments

x	A DPT object.
paths_to	Numeric Branch IDs. Are used as target(s) for the path(s) to draw.
dcs	The dimensions to use from the DiffusionMap
divide	If col_by = 'branch', this specifies which branches to divide. (see branch_divide)
w_width	Window width for smoothing the path (see smth.gaussian)
col_by	Color by 'dpt' (DPT starting at branches[[1]]), 'branch', or a variable of the data.
col_path	Colors for the path or a function creating n colors
col_tip	Color for branch tips
...	Graphical parameters supplied to plot.DiffusionMap
col	See plot.DiffusionMap . This overrides col_by

legend_main	See plot.DiffusionMap .
y, root	Root branch ID. Will be used as the start of the DPT. (default: lowest branch ID) (If longer than size 1, will be interpreted as c(root, branches))

Value

The return value of the underlying call is returned, i.e. a scatterplot3d or rgl object for 3D plots.

Examples

```
data(guo_norm)
dm <- DiffusionMap(guo_norm)
dpt <- DPT(dm)
plot(dpt)
plot(dpt, 2L, col_by = 'branch')
plot(dpt, 1L, 2:3, col_by = 'num_cells')
plot(dpt, col_by = 'DPT3')
```

plot.Sigmas	<i>Plot Sigmas object</i>
-------------	---

Description

Plot [Sigmas](#) object

Usage

```
## S4 method for signature 'Sigmas,missing'
plot(
  x,
  col = par("fg"),
  col_highlight = "#E41A1C",
  col_line = "#999999",
  type = c("b", "b"),
  pch = c(par("pch"), 4L),
  only_dim = FALSE,
  ...,
  xlab = NULL,
  ylab = NULL,
  main = ""
)
```

Arguments

x	Sigmas object to plot
col	Vector of bar colors or single color for all bars

col_highlight	Color for highest bar. Overrides col
col_line	Color for the line and its axis
type	Plot type of both lines. Can be a vector of length 2 to specify both separately (default: 'b' aka “both lines and points”)
pch	Point identifier for both lines. Can be a vector of length 2 to specify both separately (default: par(pch) and 4 (a '×'))
only_dim	logical. If TRUE, only plot the derivative line
...	Options passed to the call to plot
xlab	X label. NULL to use default
ylab	Either one y label or y labels for both plots. NULL to use both defaults, a NULL in a list of length 2 to use one default.
main	Title of the plot

Value

This method plots a Sigma object to the current device and returns nothing/NULL

Examples

```
data(guo)
sigs <- find_sigmas(guo)
plot(sigs)
```

plot_differential_map *Plot gene relevance or differential map*

Description

plot(gene_relevance, 'Gene') plots the differential map of this/these gene(s), plot(gene_relevance) a relevance map of a selection of genes. Alternatively, you can use plot_differential_map or plot_gene_relevance on a [GeneRelevance](#) or [DiffusionMap](#) object, or with two matrices.

Usage

```
plot_differential_map(
  coords,
  exprs,
  ...,
  genes,
  dims = 1:2,
  pal = hcl.colors,
  faceter = facet_wrap(~Gene)
)
```

```

## S4 method for signature 'matrix,matrix'
plot_differential_map(
  coords,
  exprs,
  ...,
  genes,
  dims = 1:2,
  pal = hcl.colors,
  faceter = facet_wrap(~Gene)
)

## S4 method for signature 'DiffusionMap,missing'
plot_differential_map(
  coords,
  exprs,
  ...,
  genes,
  dims = 1:2,
  pal = hcl.colors,
  faceter = facet_wrap(~Gene)
)

## S4 method for signature 'GeneRelevance,missing'
plot_differential_map(
  coords,
  exprs,
  ...,
  genes,
  dims = 1:2,
  pal = hcl.colors,
  faceter = facet_wrap(~Gene)
)

plot_gene_relevance(
  coords,
  exprs,
  ...,
  iter_smooth = 2L,
  n_top = 10L,
  genes = NULL,
  dims = 1:2,
  pal = palette(),
  col_na = "grey",
  limit = TRUE
)

## S4 method for signature 'matrix,matrix'
plot_gene_relevance(

```

```
    coords,
    exprs,
    ...,
    iter_smooth = 2L,
    n_top = 10L,
    genes = NULL,
    dims = 1:2,
    pal = palette(),
    col_na = "grey",
    limit = TRUE
)

## S4 method for signature 'DiffusionMap,missing'
plot_gene_relevance(
  coords,
  exprs,
  ...,
  iter_smooth = 2L,
  n_top = 10L,
  genes = NULL,
  dims = 1:2,
  pal = palette(),
  col_na = "grey",
  limit = TRUE
)

## S4 method for signature 'GeneRelevance,missing'
plot_gene_relevance(
  coords,
  exprs,
  ...,
  iter_smooth = 2L,
  n_top = 10L,
  genes = NULL,
  dims = 1:2,
  pal = palette(),
  col_na = "grey",
  limit = TRUE
)

plot_gene_relevance_rank(
  coords,
  exprs,
  ...,
  genes,
  dims = 1:2,
  n_top = 10L,
  pal = c("#3B99B1", "#F5191C"),
```

```

    bins = 10L,
    faceter = facet_wrap(~Gene)
)

## S4 method for signature 'matrix,matrix'
plot_gene_relevance_rank(
  coords,
  exprs,
  ...,
  genes,
  dims = 1:2,
  n_top = 10L,
  pal = c("#3B99B1", "#F5191C"),
  bins = 10L,
  faceter = facet_wrap(~Gene)
)

## S4 method for signature 'DiffusionMap,missing'
plot_gene_relevance_rank(
  coords,
  exprs,
  ...,
  genes,
  dims = 1:2,
  n_top = 10L,
  pal = c("#3B99B1", "#F5191C"),
  bins = 10L,
  faceter = facet_wrap(~Gene)
)

## S4 method for signature 'GeneRelevance,missing'
plot_gene_relevance_rank(
  coords,
  exprs,
  ...,
  genes,
  dims = 1:2,
  n_top = 10L,
  pal = c("#3B99B1", "#F5191C"),
  bins = 10L,
  faceter = facet_wrap(~Gene)
)

## S4 method for signature 'GeneRelevance,character'
plot(x, y, ...)

## S4 method for signature 'GeneRelevance,numeric'
plot(x, y, ...)

```

```
## S4 method for signature 'GeneRelevance,missing'
plot(x, y, ...)
```

Arguments

coords	A DiffusionMap / GeneRelevance object or a cells $\times$ dims matrix .
exprs	An cells $\times$ genes matrix . Only provide if coords is a matrix.
...	Passed to plot_differential_map / plot_gene_relevance .
genes	Genes to base relevance map on (vector of strings). You can also pass an index into the gene names (vector of numbers or logicals with length > 1). The default NULL means all genes.
dims	Names or indices of dimensions to plot. When not plotting a GeneRelevance object, the relevance for the dimensions 1:max(dims) will be calculated.
pal	Palette. Either A colormap function or a list of colors.
faceter	A ggplot faceter like facet_wrap (~ Gene).
iter_smooth	Number of label smoothing iterations to perform on relevance map. The higher the more homogenous and the less local structure.
n_top	Number the top n genes per cell count towards the score defining which genes to return and plot in the relevance map.
col_na	Color for cells that end up with no most relevant gene.
limit	Limit the amount of displayed gene labels to the amount of available colors in pal?
bins	Number of hexagonal bins for plot_gene_relevance_rank .
x	GeneRelevance object.
y	Gene name(s) or index/indices to create differential map for. (integer or character)

Value

ggplot2 plot, when plotting a relevance map with a list member \$ids containing the gene IDs used.

See Also

[gene_relevance](#), [Gene Relevance methods](#)

Examples

```
data(guo_norm)
dm <- DiffusionMap(guo_norm)
gr <- gene_relevance(dm)
plot(gr) # or plot_gene_relevance(dm)
plot(gr, 'Fgf4') # or plot_differential_map(dm, 'Fgf4')

guo_norm_mat <- t(Biobase::exprs(guo_norm))
pca <- prcomp(guo_norm_mat)$x
```

```
plot_gene_relevance(pca, guo_norm_mat, dims = 2:3)
plot_differential_map(pca, guo_norm_mat, genes = c('Fgf4', 'Nanog'))
```

projection_dist	<i>Projection distance</i>
-----------------	----------------------------

Description

Projection distance

Usage

```
projection_dist(dm, new_dcs = NULL, ..., new_data, verbose = FALSE)
```

Arguments

dm	A DiffusionMap object.
new_dcs	Diffusion component matrix of which to calculate the distance to the data.
...	Passed to proxy::dist if new_data was passed.
new_data	New data points to project into the diffusion map. Can be a matrix , data.frame , ExpressionSet , or SingleCellExperiment .
verbose	If TRUE , log additional info to the console.

Value

A vector of distances each new data point has to the existing data.

Examples

```
data(guo_norm)
g2_32 <- guo_norm[, guo_norm$num_cells < 64]
g64 <- guo_norm[, guo_norm$num_cells == 64]
dm <- DiffusionMap(g2_32)
d <- projection_dist(dm, new_data = g64)
```

random_root	<i>Find a random root cell index</i>
-------------	--------------------------------------

Description

Finds a cell that has the maximum DPT distance from a randomly selected one.

Usage

```
random_root(dm_or_dpt)
```

Arguments

dm_or_dpt A [DiffusionMap](#) or [DPT](#) object

Value

A cell index

Examples

```
data(guo)
dm <- DiffusionMap(guo)
random_root(dm)
```

Sigmas-class	<i>Sigmas Object</i>
--------------	----------------------

Description

Holds the information about how the sigma parameter for a [DiffusionMap](#) was obtained, and in this way provides a plotting function for the [find_sigmas](#) heuristic. You should not need to create a Sigmas object yourself. Provide sigma to [DiffusionMap](#) instead or use [find_sigmas](#).

Usage

```
Sigmas(...)

## S4 method for signature 'Sigmas'
optimal_sigma(object)

## S4 method for signature 'Sigmas'
print(x)

## S4 method for signature 'Sigmas'
show(object)
```

Arguments

object, x [Sigmas](#) object
 ... See “**Slots**” below

Details

A [Sigmas](#) object is either created by [find_sigmas](#) or by specifying the sigma parameter to [DiffusionMap](#).

In the second case, if the sigma parameter is just a number, the resulting [Sigmas](#) object has all slots except of `optimal_sigma` set to `NULL`.

Value

[Sigmas](#) creates an object of the same class

`optimal_sigma` retrieves the numeric value of the optimal sigma or local sigmas

Slots

`log_sigmas` Vector of length m containing the $\log_{10}$ of the σ s

`dim_norms` Vector of length $m - 1$ containing the average dimensionality $\langle p \rangle$ for the respective kernel widths

`optimal_sigma` Multiple local sigmas or the mean of the two global σ s around the highest $\langle p \rangle$ (`c(optimal_idx, optimal_idx+1L)`)

`optimal_idx` The index of the highest $\langle p \rangle$.

`avrd_norms` Vector of length m containing the average dimensionality for the corresponding sigma.

See Also

[find_sigmas](#), the function to determine a locally optimal sigma and returning this class

Examples

```
data(guo)
sigs <- find_sigmas(guo, verbose = FALSE)
optimal_sigma(sigs)
print(sigs)
```

updateObject methods *Update old destiny objects to a newer version.*

Description

Handles [DiffusionMap](#), [Sigmas](#), and [GeneRelevance](#).

Usage

```
## S4 method for signature 'DiffusionMap'  
updateObject(object, ..., verbose = FALSE)
```

```
## S4 method for signature 'Sigmas'  
updateObject(object, ..., verbose = FALSE)
```

```
## S4 method for signature 'GeneRelevance'  
updateObject(object, ..., verbose = FALSE)
```

Arguments

object	An object created with an older destiny release
...	ignored
verbose	tells what is being updated

Value

A [DiffusionMap](#) or [Sigmas](#) object that is valid when used with the current destiny release

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