

# Package ‘adSplit’

January 15, 2026

**Title** Annotation-Driven Clustering

**Version** 1.80.0

**Date** 2021-02-01

**Author** Claudio Lottaz, Joern Toedling

**Description** This package implements clustering of microarray gene expression profiles according to functional annotations. For each term genes are annotated to, splits into two subclasses are computed and a significance of the supporting gene set is determined.

**Maintainer** Claudio Lottaz <Claudio.Lottaz@klinik.uni-regensburg.de>

**Depends** R (>= 2.1.0), methods (>= 2.1.0)

**Imports** AnnotationDbi, Biobase (>= 1.5.12), cluster (>= 1.9.1), GO.db (>= 1.8.1), graphics, grDevices, KEGGREST (>= 1.30.1), multtest (>= 1.6.0), stats (>= 2.1.0)

**Suggests** golubEsets (>= 1.0), vsn (>= 1.5.0), hu6800.db (>= 1.8.1)

**LazyLoad** yes

**URL** <http://compdiag.molgen.mpg.de/software/adSplit.shtml>

**License** GPL (>= 2)

**biocViews** Microarray, Clustering

**git\_url** <https://git.bioconductor.org/packages/adSplit>

**git\_branch** RELEASE\_3\_22

**git\_last\_commit** 0efcd35

**git\_last\_commit\_date** 2025-10-29

**Repository** Bioconductor 3.22

**Date/Publication** 2026-01-15

## Contents

adSplit	2
adSplit-internal	4
diana2means	4
drawRandomPS	5
golubKEGGSplits	6

hist.splitSet . . . . .	7
image.splitSet . . . . .	8
makeEID2PROBESenv . . . . .	9
print.split . . . . .	10
print.splitSet . . . . .	10
randomDiana2means . . . . .	11

**Index****12****adSplit***Annotation-Driven Splits***Description**

This function searches for annotation-driven splits of patients in microarray data. A split is a partitioning of patients into two groups. In order to do so it refers to GO terms and KEGG pathways. In addition, a significance measure can be computed by simulating a random distribution of scores. DLD-scores are used to judge the quality of a split.

**Usage**

```
adSplit(mydata, annotation.ids, chip.name,
        min.probes = 20, max.probes = NULL,
        B = NULL, min.group.size = 5, ngenes = 50,
        ignore.genes = 5)
```

**Arguments**

<b>mydata</b>	either an expression set as defined by the package Biobase or a matrix of expression levels (rows=genes, columns=samples).
<b>annotation.ids</b>	a vector of GO or KEGG identifiers in the form "GO:..." or "KEGG:..." respectively. The prefix "KEGG:" is removed from the KEGG-identifiers before accessing the chip's "...PATH2PROBES" hash.
<b>chip.name</b>	the name of the chip by which the expression set is measured. <i>adSplit</i> attempts to load a library of the same name and expects to find a hash called "<chip-name>GO2ALLPROBES" and one called "<chip-name>PATH2PROBES" there.
<b>min.probes</b>	annotation identifiers with fewer than this associated genes are skipped.
<b>max.probes</b>	annotation identifiers with more than this associated genes are skipped. The default is ten percent of the genes on the chip.
<b>B</b>	the number of random gene set samplings to be performed to compute empirical p-values.
<b>min.group.size</b>	filter criteria to avoid splits suggesting tiny groups. Splits where one of the two suggested groups are smaller than this number are removed from the split set.
<b>ngenes</b>	number of genes used to compute DLD scores.
<b>ignore.genes</b>	number of best scoring genes to be ignored when computing DLD scores.

## Details

This function applies the same splitting procedure to all annotation identifiers provided. Firstly, the associated genes for one identifier are determined and extracted from the expression data. Then the `diana2means` function is applied to the restricted data and the different splits generated are collected into a single `splitSet` object.

As annotation identifiers vectors of identifiers of the KEGG:nnnn and GO:nnnnnn are valid. In addition, the keywords "KEGG", "GO" and "all" are allowed, representing all terms in the corresponding ontology.

If `B` is set to a integer number this number of samplings are used to generate a null-distribution of DLD-scores. This distribution is used to compute empirical p-values for each split. If more than one valid split is found, multiple testing is corrected for by applying Benjamini-Hochbergs correction from the `multtest` package.

## Value

Returns an object of class `splitSet` with the following list elements:

<code>cuts</code>	a matrix of split attributions. One row per annotation identifier (GO term or KEGG pathway for which a split has been generated. One column per object in the dataset.
<code>score</code>	one score per generated split.
<code>pvalue</code>	one empirical p-value per generated split, or <code>NULL</code>
<code>qvalue</code>	one q-value computed according Benjamini-Hochberg's correction for multiple testing per generated split, or <code>NULL</code>

## Author(s)

Claudio Lottaz, Joern Toedling

## See Also

[diana2means](#), [randomDiana2means](#), [image.splitSet](#)

## Examples

```
# prepare data
library(golubEsets)
data(Golub_Merge)

# generate annotation-driven splits for apoptosis and signal transduction
x <- adSplit(Golub_Merge, "GO:0006915", "hu6800")
x <- adSplit(Golub_Merge, c("GO:0007165","GO:0006915"), "hu6800", max.probes=7000)

# generate a split for alanine, aspartate and glutamate metabolism including
# an empirical p-value
x <- adSplit(Golub_Merge, "KEGG:00250", "hu6800", B=100)

# generate splits for all KEGG pathways.
x <- adSplit(Golub_Merge, "KEGG", "hu6800")
image(x)
```

---

adSplit-internal      *adSplit Internal Function*

---

## Description

Function for internal use only.

## Details

No details given. This function is subject to change without further notice.

## Author(s)

Claudio Lottaz

---

diana2means      *2-Means with Hierarchical Initialization*

---

## Description

Split a set of data points into two coherent groups using the k-means algorithm. Instead of random initialization, divisive hierarchical clustering is used to determine initial groups and the corresponding centroids.

## Usage

```
diana2means(mydata, mingroupsize = 5,
            ngenes = 50, ignore.genes = 5,
            return.cut = FALSE)
```

## Arguments

mydata	either an expression set as defined by the package Biobase or a matrix of expression levels (rows=genes, columns=samples).
mingroupsize	report only splits where both groups are larger than this size.
ngenes	number of genes used to compute cluster quality DLD-score.
ignore.genes	number of best scoring genes to be ignored when computing DLD-scores.
return.cut	logical, whether to return the attributions of samples to groups.

## Details

This function uses divisive hierarchical clustering (diana) to generate a first split of the data. Thereby, each column of the data matrix is considered to represent a data element. From the thus generated tentative groups, centroids are deduced and used to initialize the k-means clustering algorithm.

For the split optimized by k-means the DLD-score is determined using the ngenes and ignore.genes arguments.

**Value**

If the logical `return.cut` is set to FALSE (the default), a single number is representing the DLD-score for the generated split is returned. Otherwise an object of class `split` containing the following elements is returned:

<code>cut</code>	one number out of 0 and 1 per column in the original data, specifying the split attribution.
<code>score</code>	the DLD-score achieved by the split.

**Author(s)**

Joern Toedling, Claudio Lottaz

**See Also**

[diana](#)

**Examples**

```
# get golub data
library(vsn)
library(golubEsets)
data(Golub_Merge)

# use 10% most variable genes
e <- exprs(Golub_Merge)
vars <- apply(e, 1, var)
e <- e[vars > quantile(vars,0.9),]

# use diana2means to get splits and scores
diana2means(e)
diana2means(e, return.cut=TRUE)
```

drawRandomPS

*Draw sets of probe-sets*

**Description**

This function draws a given number of probe-sets randomly, such that probe-sets referring to the same are either included or excluded as a whole.

**Usage**

```
drawRandomPS(nps, EID2PSenv, allEIDs)
```

**Arguments**

<code>nps</code>	number of probe-sets to be drawn.
<code>EID2PSenv</code>	a hash mapping EntrezGene to probe-set identifiers.
<code>allEIDs</code>	vector of all EntrezGene identifiers represented on a chip.

**Value**

A named vector of probe-set identifiers. The names correspond to the EntrezGene identifiers.

**Author(s)**

Claudio Lottaz

**Examples**

```
# draw ten random probe-sets from hu6800
library(hu6800.db)
EID2PSenv <- makeEID2PROBESenv(hu6800ENTREZID)
drawRandomPS(10, EID2PSenv, ls(EID2PSenv))
```

golubKEGGSplits

*Exemplar splitSet*

**Description**

This is a data object precomputed by `adSplit` for illustration.

**Usage**

```
data(golubKEGGSplits)
```

**Format**

Annotation-driven split set holds 70 splits on 72 elements, scores range is: 3.382672 17.31385, empirical p-values range is: 0.005 0.955, q-value range is: 0.1633333 0.955.

**Details**

This object is generated by the following call:

```
golubKEGGSplits <- adSplit(golubNorm, "KEGG", "hu6800", B=1000)
```

where `golubNorm` is a normalized version of `Golub_Merge` from the `golubEsets` package.

**Examples**

```
data(golubKEGGSplits)
```

---

**hist.splitSet***Overview Histogram for splitSets*

---

**Description**

Draws a histogram of empirical p-values and shows the corresponding q-values corrected for multiple testing.

**Usage**

```
## S3 method for class 'splitSet'  
hist(x, main = "Distribution of p-Values",  
     xlab = "p-values", col = "grey", xlim = c(0, 1), ...)
```

**Arguments**

<code>x</code>	object of type <code>splitSet</code> . Should hold a considerable number of splits.
<code>main</code>	main title of the histogram.
<code>xlab</code>	legend for the x-axis.
<code>col</code>	color for the histogram bars.
<code>xlim</code>	limits for the x-axis (p-values).
<code>...</code>	further parameters passed on to the default <code>hist</code> function.

**Details**

This function draws a regular histogram of empirical p-values observed in the `splitSet` at hand. The corresponding q-values, corrected by the method suggested by Benjamini-Hochberg, are plotted into the same graph. The scale for the q-values is shown at the left hand side of the plot.

**Author(s)**

Claudio Lottaz

**See Also**

[adSplit](#)

**Examples**

```
data(golubKEGGSplits)  
hist(golubKEGGSplits, col="red")
```

---

<code>image.splitSet</code>	<i>Illustrate Split Sets</i>
-----------------------------	------------------------------

---

## Description

Draws an image of all splits, one per row, of a `splitSet` object. Each column corresponds to a patient.

## Usage

```
## S3 method for class 'splitSet'
image(x, filter.fdr = 1, main = "", max.label.length = 50,
      full.names = TRUE, xlab = NULL, sample.labels = FALSE,
      col = c("yellow", "red"), invert = FALSE,
      outfile = NULL, res = 72, pointsize = 7, ...)
```

## Arguments

<code>x</code>	the object of class <code>splitSet</code> to be illustrated.
<code>filter.fdr</code>	worst acceptable false discovery rate for the shown set of splits. All splits with q-values below this level are dropped from the image.
<code>main</code>	a title for the image.
<code>max.label.length</code>	Maximal length of the annotations shown to the right of the image. Longer annotations are truncated.
<code>full.names</code>	Show full names for annotations instead of their identifiers only.
<code>xlab</code>	additional annotation on the x-axis.
<code>sample.labels</code>	whether names of samples are to be shown on the x-axis.
<code>col</code>	two strings encoding the colors to be used to illustrate to which group a sample is attributed.
<code>invert</code>	whether to draw in white on black background.
<code>outfile</code>	the filename on which to draw the image in postscript format. The default is <code>NULL</code> , meaning to produce the image interactively.
<code>res</code>	resolution for bitmap output on postscript.
<code>pointsize</code>	size of font.
<code>...</code>	further arguments passed to <code>image</code> .

## Details

The set of splits given is illustrated as an image. Each row corresponds to an annotation, each column to a patient. In position (x,y), the association of patient x to a group with respect to annotation y is coded as colors (yellow and red by default). The image is ordered by hierarchical clustering such that similar patients and similar splits are brought closer together.

## Value

Always returns `NULL`.

**Author(s)**

Claudio Lottaz

**See Also**

[adSplit](#)

**Examples**

```
data(golubKEGGSplits)
image(golubKEGGSplits, filter.fdr=0.5)
```

---

**makeEID2PROBESenv**

*Generate EID2PROBES environment*

---

**Description**

Make hash containing probe-sets per EntrezGene identifier.

**Usage**

```
makeEID2PROBESenv(EIDenv)
```

**Arguments**

EIDenv	an environment containing one entry per probe-set holding all corresponding EntrezGene identifiers.
--------	---

**Value**

An environment containing one entry per EntrezGene identifier holding all corresponding probe-sets.

**Author(s)**

Joern Toedling, Claudio Lottaz

**Examples**

```
library(hu6800.db)
makeEID2PROBESenv(hu6800ENTREZID)
```

---

**print.split** *Print Information on Single Splits*

---

## Description

Print information on a single split.

## Usage

```
## S3 method for class 'split'  
print(x, ...)
```

## Arguments

x the split-object to be described.  
... not used.

## Author(s)

Claudio Lottaz

## See Also

[diana2means](#)

---

**print.splitSet** *Print Summaries for splitSets*

---

## Description

Prints information of splitSets.

## Usage

```
## S3 method for class 'splitSet'  
print(x, ...)
```

## Arguments

x the splitSet-object to be described.  
... not used.

## Author(s)

Claudio Lottaz

## See Also

[adSplit](#)

## Examples

```
data(golubKEGGSplits)
print(golubKEGGSplits)
```

---

randomDiana2means      *Generate null-distributions of DLD-scores*

---

## Description

Draws a number of random sets of probe-sets consisting of the needed size and applies `diana2means` to compute DLD scores.

## Usage

```
randomDiana2means(nprobes, data, chip, ndraws = 10000,
                    ngenes = 50, ignore.genes = 5)
```

## Arguments

<code>nprobes</code>	the size of gene sets.
<code>data</code>	a matrix of expression data, rows correspond to genes, columns to samples.
<code>chip</code>	the name of the used chip.
<code>ndraws</code>	the number of DLD scores computed.
<code>ngenes</code>	the number of genes used to compute DLD scores (passed to <code>diana2means</code> ).
<code>ignore.genes</code>	the number of best scoring genes to be ignored when computing DLD scores (passed to <code>diana2means</code> )

## Details

This function uses `drawRandomPS` to draw `ndraws` gene sets. On these it applies `diana2means` to determine a null-distribution of DLD-scores.

## Value

A vector of DLD-scores.

## Author(s)

Joern Toedling, Claudio Lottaz

## See Also

[drawRandomPS](#), [diana2means](#)

## Examples

```
# prepare data
library(vsn)
library(golubEsets)
data(Golub_Merge)

# generate DLD scores
scores <- randomDiana2means(20, exprs(Golub_Merge), "hu6800", ndraws = 500)
```

# Index

- \* **datagen**
  - adSplit, 2
  - diana2means, 4
  - drawRandomPS, 5
  - makeEID2PROBESenv, 9
  - randomDiana2means, 11
- \* **datasets**
  - golubKEGGSplits, 6
- \* **hplot**
  - hist.splitSet, 7
  - image.splitSet, 8
- \* **internal**
  - adSplit-internal, 4
  - print.split, 10
  - print.splitSet, 10
- adSplit, 2, 7, 9, 10
- adSplit-internal, 4
- diana, 5
- diana2means, 3, 4, 10, 11
- drawRandomPS, 5, 11
- golubKEGGSplits, 6
- hist.splitSet-method (hist.splitSet), 7
- hist.splitSet, 7
- image.splitSet-method (image.splitSet), 8
- image.splitSet, 3, 8
- makeEID2PROBESenv, 9
- print.split-method (print.split), 10
- print.splitSet-method (print.splitSet), 10
- print.split, 10
- print.splitSet, 10
- randomDiana2means, 3, 11
- tscore (adSplit-internal), 4