

Package ‘FlowSOM’

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AddAnnotation

*AddAnnotation***Description**

Add annotation to a FlowSOM plot

Usage

```
AddAnnotation(
  p,
  fsom,
  toAnnotate = NULL,
  prefix = list(metaclusters = "MCL ", clusters = "CL "),
  ...
)
```

Arguments

<code>p</code>	Plot to add annotation to. When using PlotStars , please use <code>list_insteadof_ggarrange = TRUE</code> .
<code>fsom</code>	FlowSOM object that goes with the plot.
<code>toAnnotate</code>	A named list with "metaclusters" and/or "clusters" as names and a vector with the (meta)clusters that need to be annotated. Names can be abbreviated. Use a named vector with the old names as values and new labels as names for custom labeling.
<code>prefix</code>	Prefix to be added to labels. Default is "MCL " and "CL " for metaclusters and clusters respectively.
<code>...</code>	Arguments passed to <code>geom_text_repel</code> .

Value

The updated plot

Examples

```
# Identify the files
fcs <- flowCore::read.FCS(system.file("extdata", "68983.fcs",
                                     package = "FlowSOM"))

# Build a FlowSOM object
flowSOM.res <- FlowSOM(fcs,
  scale = TRUE,
  compensate = TRUE,
  transform = TRUE,
  toTransform = 8:18,
  colsToUse = c(9, 12, 14:18),
  nClus = 10,
  seed = 1)

p <- PlotStars(flowSOM.res, backgroundValues = flowSOM.res$metaclustering,
  list_insteadof_ggarrange = TRUE)
annotationList <- list("metaclusters" = c("CD8 T cells" = "1", "B cells" = "8"),
  "clusters" = c(97))
AddAnnotation(p, flowSOM.res, toAnnotate = annotationList,
  prefix = list("metaclusters" = "", clusters = "CL "))
```

AddBackground	<i>AddBackground</i>
---------------	----------------------

Description

Function plots the background

Usage

```
AddBackground(
  p,
  backgroundValues,
  backgroundColors = NULL,
  backgroundLim = NULL
)
```

Arguments

p	ggplot object
backgroundValues	Vector of values to be plotted as background for the nodes
backgroundColors	Color palette to be used for the background coloring. Can be either a function or an array specifying colors.
backgroundLim	Background limits (can be used to ensure consistent Color palette between plots). If NULL (default), will be automatically adapted to the data.

Value

Returns nothing, but plots the background

See Also

[PlotFlowSOM](#), [AddLabels](#), [AddNodes](#), [AddPies](#), [AddStars](#)

AddFlowFrame	<i>Add a flowFrame to the data variable of the FlowSOM object</i>
--------------	---

Description

Add a flowFrame to the data variable of the FlowSOM object

Usage

```
AddFlowFrame(fsom, flowFrame)
```

Arguments

fsom	FlowSOM object, as constructed by the ReadInput function
flowFrame	flowFrame to add to the FlowSOM object

Value

FlowSOM object with data added

See Also

[ReadInput](#)

AddLabels	<i>AddLabels</i>
-----------	------------------

Description

AddLabels

Usage

```
AddLabels(  
  p,  
  labels,  
  hjust = 0.5,  
  layout = NULL,  
  textSize = 3.88,  
  textColor = "black",  
  ...  
)
```

Arguments

p	ggplot object
labels	Labels to be added to each node
hjust	Horizontal adjust for labels. Default is centered.
layout	Dataframe with x and y columns. If null, the dataframe from the ggplot object will be reused.
textSize	Size for geom_text. Default (=3.88) is from geom_text.
textColor	Color for geom_text. Default = black.
...	Additional parameters to pass to geom_text

Value

Returns the ggplot object with labels added

See Also

[PlotLabels](#), [PlotNumbers](#)

`AddMST`*AddMST*

Description

Function plots the MST

Usage

```
AddMST(p, fsom)
```

Arguments

<code>p</code>	ggplot object
<code>fsom</code>	FlowSOM object, as generated by FlowSOM

Value

Returns nothing, but plots the MST for FlowSOM MST view

See Also

[PlotFlowSOM](#), [ParseEdges](#), [AddStarsPies](#), [AddLabels](#), [AddNodes](#), [AddBackground](#), [AddPies](#), [AddStars](#)

`AddNodes`*AddNodes*

Description

Function plots the nodes

Usage

```
AddNodes(  
  p,  
  nodeInfo = NULL,  
  values = NULL,  
  lim = NULL,  
  colorPalette = NULL,  
  fillColor = "white",  
  showLegend = TRUE,  
  label = "",  
  ...  
)
```

AddScale	<i>AddScale</i>
----------	-----------------

Description

AddScale

Usage

```
AddScale(  
  p,  
  values = NULL,  
  colors = NULL,  
  limits = NULL,  
  showLegend = TRUE,  
  labelLegend = "",  
  type = "fill"  
)
```

Arguments

p	ggplot object
values	Values used for the fill
colors	Colors to use (can be a vector or a function)
limits	Limits to use in the scale
showLegend	Boolean on whether to show the legend
labelLegend	Label to show as title of the legend
type	fill (default) or color

Value

ggplot object with scale added

AddStars	<i>AddStars</i>
----------	-----------------

Description

Function plots the stars

Usage

```
AddStars(p, fsom, markers = fsom$map$colsUsed, colorPalette = NULL)
```

Arguments

p	ggplot object
fsom	FlowSOM object, as generated by BuildMST
markers	Determines which markers to plot. Default = "fsom\$map\$colsUsed"
colorPalette	Color palette to be used for colors. Can be either a function or an array specifying colors.

Value

ggplot object with the stars added

See Also

[PlotFlowSOM](#), [AddLabels](#), [AddNodes](#), [AddBackground](#), [PlotStars](#), [AddPies](#), [ParseArcs](#)

AddStarsPies	<i>AddStarsPies</i>
--------------	---------------------

Description

Function plots stars or pies

Usage

```
AddStarsPies(p, arcs, colorPalette, showLegend = TRUE)
```

Arguments

p	ggplot object
arcs	Dataframe that contains all the data for the plotting the pies or stars
colorPalette	A vector of colors or a color function
showLegend	Boolean on whether to show the legend

Value

Returns nothing, but plots the stars or pies

See Also

[PlotFlowSOM](#), [AddLabels](#), [AddNodes](#), [AddBackground](#), [AddPies](#), [AddStars](#), [ParseArcs](#), [PlotStars](#), [PlotPies](#)

AggregateFlowFrames *Aggregate multiple FCS files together*

Description

Aggregate multiple FCS files to analyze them simultaneously. A new FCS file is written, which contains about `cTotal` cells, with `ceiling(cTotal/nFiles)` cells from each file. Two new columns are added: a column indicating the original file by index, and a noisy version of this for better plotting opportunities (index plus or minus a value between 0 and 0.1).

Usage

```
AggregateFlowFrames(
  fileNames,
  cTotal,
  channels = NULL,
  writeOutput = FALSE,
  outputFile = "aggregate.fcs",
  keepOrder = FALSE,
  silent = FALSE,
  sampleWithReplacement = FALSE,
  ...
)
```

Arguments

<code>fileNames</code>	Character vector containing full paths to the FCS files or a flowSet to aggregate
<code>cTotal</code>	Total number of cells to write to the output file
<code>channels</code>	Channels/markers to keep in the aggregate. Default NULL takes all channels of the first file.
<code>writeOutput</code>	Whether to write the resulting flowFrame to a file. Default FALSE
<code>outputFile</code>	Full path to output file. Default "aggregate.fcs"
<code>keepOrder</code>	If TRUE, the random subsample will be ordered in the same way as they were originally ordered in the file. Default = FALSE.
<code>silent</code>	If FALSE, prints an update every time it starts processing a new file. Default = FALSE.
<code>sampleWithReplacement</code>	If TRUE and more cells per file are requested than actually present, all cells will be included plus additional resampling. Otherwise, at most all cells will be included once. Default = FALSE.
<code>...</code>	Additional arguments to pass to read.FCS

Value

This function does not return anything, but will write a file with about `cTotal` cells to `outputFile`

See Also

[ceiling](#)

Examples

```
# Define filename
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
# This example will sample 2 times 500 cells.
ff_new <- AggregateFlowFrames(c(fileName, fileName), 1000)
```

AutoMaxNodeSize	<i>AutoMaxNodeSize</i>
-----------------	------------------------

Description

Calculate node size

Usage

```
AutoMaxNodeSize(layout, overlap)
```

Arguments

layout	Coordinates of nodes
overlap	Parameter that determines how much overlap there will be. If negative the nodes will be smaller

Details

Function that calculates the minimum distance between the nodes to use this to adapt the maxNodeSize for better plotting

Value

Returns the maxNodeSize with some overlap

See Also

[PlotFlowSOM](#), [ScaleStarHeights](#), [ParseNodeSize](#)

BuildMST	<i>BuildMST</i>
----------	-----------------

Description

Build Minimal Spanning Tree

Usage

```
BuildMST(fsom, silent = FALSE, tSNE = FALSE)
```

Arguments

fsom	FlowSOM object, as generated by BuildSOM
silent	If TRUE, no progress updates will be printed
tSNE	If TRUE, an alternative t-SNE layout is computed as well

Details

Add minimal spanning tree description to the FlowSOM object

Value

FlowSOM object containing MST description

See Also

[BuildSOM](#), [PlotStars](#)

Examples

```
# Read from file, build self-organizing map
fileName <- system.file("extdata", "68983.fcs", package="FlowSOM")
flowSOM.res <- ReadInput(fileName, compensate=TRUE, transform = TRUE,
                        scale = TRUE)
flowSOM.res <- BuildSOM(flowSOM.res, colsToUse = c(9, 12, 14:18))

# Build the Minimal Spanning Tree
flowSOM.res <- BuildMST(flowSOM.res)
```

BuildSOM

Build a self-organizing map

Description

Build a SOM based on the data contained in the FlowSOM object

Usage

```
BuildSOM(fsom, colsToUse = NULL, silent = FALSE, outlierMAD = 4, ...)
```

Arguments

fsom	FlowSOM object containing the data, as constructed by the ReadInput function
colsToUse	Markers, channels or indices to use for building the SOM
silent	if TRUE, no progress updates will be printed
outlierMAD	Number of MAD when a cell is considered an outlier. See also TestOutliers
...	options to pass on to the SOM function (xdim, ydim, rlen, mst, alpha, radius, init, distf, importance)

Value

FlowSOM object containing the SOM result, which can be used as input for the [BuildMST](#) function

References

This code is strongly based on the kohonen package. R. Wehrens and L.M.C. Buydens, Self- and Super-organising Maps in R: the kohonen package J. Stat. Softw., 21(5), 2007

See Also

[ReadInput](#), [BuildMST](#)

Examples

```
# Read from file
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
flowSOM.res <- ReadInput(fileName, compensate = TRUE, transform = TRUE,
                          scale = TRUE)

# Build the Self-Organizing Map
# E.g. with gridsize 5x5, presenting the dataset 20 times,
# no use of MST in neighborhood calculations in between
flowSOM.res <- BuildSOM(flowSOM.res, colsToUse = c(9, 12, 14:18),
                        xdim = 5, ydim = 5, rlen = 20)

# Build the minimal spanning tree and apply metaclustering
flowSOM.res <- BuildMST(flowSOM.res)
metacl <- MetaClustering(flowSOM.res$map$codes,
                         "metaClustering_consensus", max = 10)
```

CountGroups

Calculate differences in cell counts between groups

Description

Calculate differences in cell counts between groups

Usage

```
CountGroups(fsom, groups, plot = TRUE, silent = FALSE)
```

Arguments

fsom	FlowSOM object as generated by BuildSOM
groups	List containing an array with file names for each group
plot	Logical. If TRUE, make a starplot of each individual file
silent	Logical. If TRUE, print progress messages

Value

Distance matrix

See Also

GroupStats

Examples

```

set.seed(1)
fileName <- system.file("extdata", "68983.fcs", package="FlowSOM")
flowSOM.res <- FlowSOM(fileName, compensate = TRUE, transform = TRUE,
                      scale = TRUE, colsToUse = c(9,12,14:18), nClus = 10)

ff <- flowCore::read.FCS(fileName)
# Make an additional file without cluster 7 and double amount of cluster 5
selection <- c(which(GetClusters(flowSOM.res) %in%
                      which(flowSOM.res$metaclustering != 7)),
               which(GetClusters(flowSOM.res) %in%
                      which(flowSOM.res$metaclustering == 5)))
ff_tmp <- ff[selection,]
flowCore::write.FCS(ff_tmp, file="ff_tmp.fcs")

# Compare only the file with the double amount of cluster 10
features <- GetFeatures(flowSOM.res,
                       c(fileName, "ff_tmp.fcs"),
                       level = "clusters",
                       type = "percentages")
stats <- GroupStats(features$cluster_percentages,
                    groups = list("AllCells" = c(fileName),
                                  "Without_ydTcells" = c("ff_tmp.fcs")))

```

Dist.MST

Calculate distance matrix using a minimal spanning tree neighborhood

Description

Calculate distance matrix using a minimal spanning tree neighborhood

Usage

Dist.MST(X)

Arguments

X matrix in which each row represents a point

Value

Distance matrix

FlowSOM

*Run the FlowSOM algorithm***Description**

Method to run general FlowSOM workflow. Will scale the data and uses consensus meta-clustering by default.

Usage

```
FlowSOM(
  input,
  pattern = ".fcs",
  compensate = FALSE,
  spillover = NULL,
  transform = FALSE,
  toTransform = NULL,
  transformFunction = flowCore::logicleTransform(),
  transformList = NULL,
  scale = FALSE,
  scaled.center = TRUE,
  scaled.scale = TRUE,
  silent = TRUE,
  colsToUse = NULL,
  nClus = 10,
  maxMeta = NULL,
  importance = NULL,
  seed = NULL,
  ...
)
```

Arguments

input	a flowFrame, a flowSet, a matrix with column names or an array of paths to files or directories
pattern	if input is an array of file- or directorynames, select only files containing pattern
compensate	logical, does the data need to be compensated
spillover	spillover matrix to compensate with If NULL and compensate = TRUE, we will look for \$SPILL description in FCS file.
transform	logical, does the data need to be transformed with the transformation given in transformFunction.
toTransform	column names or indices that need to be transformed. Will be ignored if transformList is given. If NULL and transform = TRUE, column names of \$SPILL description in FCS file will be used.
transformFunction	Defaults to logicleTransform()
transformList	transformList to apply on the samples.
scale	logical, does the data needs to be rescaled. Default = FALSE

Value

array of n colors

FMeasure	<i>F measure</i>
----------	------------------

Description

Compute the F measure between two clustering results

Usage

```
FMeasure(realClusters, predictedClusters, silent = FALSE)
```

Arguments

- realClusters Array containing real cluster labels for each sample
- predictedClusters Array containing predicted cluster labels for each sample
- silent Logical, if FALSE (default), print some information about precision and recall

Value

F measure score

Examples

```
# Generate some random data as an example
realClusters <- sample(1:5,100,replace = TRUE)
predictedClusters <- sample(1:6, 100, replace = TRUE)

# Calculate the FMeasure
FMeasure(realClusters,predictedClusters)
```

GetChannels	<i>GetChannels</i>
-------------	--------------------

Description

Get channel names for an array of markers, given a flowFrame or a FlowSOM object. As available in "name". [grep](#) is used to look for the markers. Other regex can be added.

Usage

```
GetChannels(object, markers, exact = TRUE)
```

GetClusterMFIs	<i>Get MFI values for all clusters</i>
----------------	--

Description

Get MFI values for all clusters

Usage

```
GetClusterMFIs(fsom, colsUsed = FALSE, prettyColnames = FALSE)
```

Arguments

fsom	FlowSOM object as generated by the FlowSOM function or the BuildSOM function
colsUsed	logical. Should report only the columns used to build the SOM. Default = FALSE.
prettyColnames	logical. Should report pretty column names instead of standard column names. Default = FALSE.

Value

Matrix with median values for each marker

Examples

```
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
flowSOM.res <- FlowSOM(fileName, compensate = TRUE, transform = TRUE,
  scale = TRUE, colsToUse = c(9, 12, 14:18), nClus = 10)
mfis <- GetClusterMFIs(flowSOM.res)
```

GetClusterPercentagesPositive	<i>Get percentage-positive values for all clusters</i>
-------------------------------	--

Description

Get percentage-positive values for all clusters

Usage

```
GetClusterPercentagesPositive(
  fsom,
  cutoffs,
  colsUsed = FALSE,
  prettyColnames = FALSE
)
```

GetCounts

GetCounts

Description

Get counts of number of cells in clusters or metaclusters

Usage

```
GetCounts(fsom, level = "metaclusters")
```

Arguments

fsom	FlowSOM object
level	Character string, should be either "clusters" or "metaclusters" (default) or abbreviations.

Value

A named vector with the counts

Examples

```
# Read from file, build self-organizing map and minimal spanning tree
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
ff <- flowCore::read.FCS(fileName)
ff <- flowCore::compensate(ff, flowCore::keyword(ff)[["SPILL"]])
ff <- flowCore::transform(ff, flowCore::estimateLogicle(ff,
                                                         flowCore::colnames(ff)[8:18]))

flowSOM.res <- FlowSOM(ff,
                      scale = TRUE,
                      colsToUse = c(9, 12, 14:18),
                      nClus = 10,
                      seed = 1)

GetCounts(flowSOM.res)
GetCounts(flowSOM.res, level = "clusters")
```

GetCVs

Get CV values for all clusters

Description

Get CV values for all clusters

Usage

```
GetCVs(fsom)
```

Arguments

fsom	FlowSOM object as generated by the FlowSOM function or the BuildSOM function
------	--

Value

Matrix with coefficient of variation values for each marker

```
fileName <- system.file("extdata", "68983.fcs", package="FlowSOM") flowSOM.res <- FlowSOM(fileName,
compensate=TRUE,transform=TRUE, scale=TRUE,colsToUse=c(9,12,14:18),nClus=10) cvs <- Get-
ClusterCVs(flowSOM.res)
```

GetFeatures	<i>GetFeatures</i>
-------------	--------------------

Description

Map FCS files on an existing FlowSOM object

Usage

```
GetFeatures(
  fsom,
  files,
  level = c("clusters", "metaclusters"),
  type = "counts",
  MFI = NULL,
  positive_cutoffs = NULL,
  filenames = NULL,
  silent = FALSE
)
```

Arguments

fsom	FlowSOM object as generated by the FlowSOM function or the BuildSOM function
files	Either a vector of FCS files or paths to FCS files
level	Level(s) of interest. Default is c("clusters", "metaclusters"), but can also be only one of them. Can be abbreviated.
type	Type of features to extract. Default is "counts", can be a vector of "counts", "percentages", "MFIs" and/or "percentages_positive" or abbreviations.
MFI	Vector with channels / markers for which the MFI values must be returned when "MFIs" is in type
positive_cutoffs	Named vector with fluorescence-intensity values per channel / marker that are the upper bounds for a negative population when "percentages_positive" is in type
filenames	An optional vector with filenames that will be used as rownames in the count matrices. If NULL (default) either the paths will be used or a numerical vector.
silent	Logical. If TRUE, print progress messages. Default = FALSE.

Value

matrix with features per population - type combination

Examples

```
# Build FlowSom result
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
ff <- flowCore::read.FCS(fileName)
ff <- flowCore::compensate(ff, flowCore::keyword(ff)[["SPILL"]])
ff <- flowCore::transform(ff,
  flowCore::transformList(colnames(flowCore::keyword(ff)[["SPILL"]]),
    flowCore::logicTransform()))
flowSOM.res <- FlowSOM(ff[1:1000, ],
  scale = TRUE,
  colsToUse = c(9, 12, 14:18),
  nClus = 10)

# Map new data
counts <- GetFeatures(fsom = flowSOM.res,
  level = "clusters",
  files = c(ff[1001:2000, ], ff[2001:3000, ]))
features <- GetFeatures(fsom = flowSOM.res,
  files = c(ff[1001:2000, ], ff[2001:3000, ]),
  type = c("counts", "percentages", "MFIs"),
  MFI = "APC-A",
  filenames = c("ff_1001-2000", "ff_2001-3000"))

# Get percentages of positive cells
positive_cutoffs <- c('CD8' = 1.5,
  'CD4' = 0.3,
  'CD19' = 1.3,
  'CD3' = -0.3)

perc_pos <- GetFeatures(fsom = flowSOM.res,
  files = c(ff[1001:2000, ], ff[2001:3000, ]),
  type = c("percentages_positive"),
  positive_cutoffs = positive_cutoffs,
  filenames = c("ff_1001-2000", "ff_2001-3000"))
```

GetFlowJoLabels

Process a FlowJo workspace file

Description

Reads a FlowJo workspace file using the flowWorkspace library and returns a list with a matrix containing gating results and a vector with a label for each cell from a set of specified gates

Usage

```
GetFlowJoLabels(
  files,
  wspFile,
  group = "All Samples",
  cellTypes = NULL,
  getData = FALSE,
  ...
)
```

Arguments

<code>files</code>	The FCS files of interest
<code>wspFile</code>	The FlowJo wsp file to read
<code>group</code>	The FlowJo group to parse. Default "All Samples".
<code>cellTypes</code>	Cell types to use for final labeling the cells. Should correspond with a subset of the gate names in FlowJo.
<code>getData</code>	If true, flowFrames are returned as well.
<code>...</code>	Extra arguments to pass to <code>CytoML::flowjo_to_gatingset</code>

Value

This function returns a list, which for every file contains a list in which the first element ("matrix") is a matrix containing filtering results for each specified gate and the second element ("manual") is a vector which assigns one label to each cell. If only one file is given, only one list is returned instead of a list of lists.

See Also

[PlotPies](#)

Examples

```
# Identify the files
fcs_file <- system.file("extdata", "68983.fcs", package = "FlowSOM")
wspFile <- system.file("extdata", "gating.wsp", package = "FlowSOM")

# Specify the cell types of interest for assigning one label per cell
cellTypes <- c("B cells",
               "gd T cells", "CD4 T cells", "CD8 T cells",
               "NK cells", "NK T cells")

# Parse the FlowJo workspace
gatingResult <- GetFlowJoLabels(fcs_file, wspFile,
                               cellTypes = cellTypes,
                               getData = TRUE)

# Check the number of cells assigned to each gate
colSums(gatingResult$matrix)

# Build a FlowSOM tree
flowSOM.res <- FlowSOM(gatingResult$flowFrame,
                      colsToUse = c(9, 12, 14:18),
                      nClus = 10,
                      seed = 1)

# Plot pies indicating the percentage of cell types present in the nodes
PlotPies(flowSOM.res,
         gatingResult$manual,
         backgroundValues = flowSOM.res$metaclustering)
```

GetMarkers	<i>GetMarkers</i>
------------	-------------------

Description

Get marker names for an array of channels, given a flowFrame or a FlowSOM object. As available in "desc". If this is NA, defaults to channel name. [grep](#) is used to look for the markers. Other regex can be added.

Usage

GetMarkers(object, channels, exact = TRUE)

Arguments

- | | |
|----------|---|
| object | The flowFrame or the FlowSOM object of interest |
| channels | Vector with markers or channels of interest. Also accepts the index of the channel in the object. |
| exact | If TRUE (default), the grep pattern will be extended to start with ^\\Q and end with \\E\$, so only exact matches are possible. |

Value

Corresponding marker names

See Also

[GetChannels](#)

Examples

```
# Read the flowFrame
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
ff <- flowCore::read.FCS(fileName)
GetChannels(ff, c("FSC-A", "CD3", "FITC-A"))
GetMarkers(ff, c("FSC-A", "CD3", "FITC-A"))
```

GetMetaclusterCVs	<i>GetMetaclusterCVs</i>
-------------------	--------------------------

Description

Compute the coefficient of variation for the metaclusters

Usage

GetMetaclusterCVs(fsom, colsUsed = FALSE, prettyColnames = FALSE)

Examples

```

fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
ff <- flowCore::read.FCS(fileName)
ff <- flowCore::compensate(ff, flowCore::keyword(ff)[["SPILL"]])
ff <- flowCore::transform(ff,
  flowCore::transformList(colnames(flowCore::keyword(ff)[["SPILL"]]),
    flowCore::logicleTransform()))
flowSOM.res <- FlowSOM(ff,
  scale = TRUE,
  colsToUse = c(9, 12, 14:18),
  nClus = 10)
mfis <- GetMetaclusterMFIs(flowSOM.res)

```

GetMetaclusterPercentagesPositive

Get percentage-positive values for all metaclusters

Description

Get percentage-positive values for all metaclusters

Usage

```

GetMetaclusterPercentagesPositive(
  fsom,
  cutoffs,
  colsUsed = FALSE,
  prettyColnames = FALSE
)

```

Arguments

fsom	FlowSOM object as generated by the FlowSOM function or the BuildSOM function
cutoffs	named numeric vector. Upper bounds of negative population fluorescence-intensity values for each marker / channel.
colsUsed	logical. Should report only the columns used to build the SOM. Default = FALSE.
prettyColnames	logical. Should report pretty column names instead of standard column names. Default = FALSE.

Value

Matrix with percentages of cells that are positive in selected markers per each metacluster

Examples

```

fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
flowSOM.res <- FlowSOM(fileName, compensate = TRUE, transform = TRUE,
  scale = TRUE, colsToUse = c(9, 12, 14:18), nClus = 10)
perc_pos <- GetMetaclusterPercentagesPositive(flowSOM.res, cutoffs = c('CD4' = 5000))

```

get_channels	<i>get_channels</i>
--------------	---------------------

Description

Get channel names for an array of markers, given a flowFrame

Usage

```
get_channels(ff, markers)
```

Arguments

ff	The flowFrame of interest
markers	Vector with markers or channels of interest

Value

Corresponding channel names

See Also

[get_markers](#)

Examples

```
# Read the flowFrame
fileName <- system.file("extdata", "68983.fcs", package="FlowSOM")
ff <- flowCore::read.FCS(fileName)
GetChannels(ff, c("FSC-A", "CD3", "FITC-A"))
GetMarkers(ff, c("FSC-A", "CD3", "FITC-A"))
```

get_markers	<i>get_markers</i>
-------------	--------------------

Description

Get marker names, given a flowFrame. As available in "desc". If this is NA, defaults to channel name.

Usage

```
get_markers(ff, markers)
```

Arguments

ff	The flowFrame of interest
markers	Vector with markers or channels of interest

Value

Corresponding marker names

See Also

[get_channels](#)

Examples

```
# Read the flowFrame
fileName <- system.file("extdata", "68983.fcs", package="FlowSOM")
ff <- flowCore::read.FCS(fileName)
GetChannels(ff, c("FSC-A", "CD3", "FITC-A"))
GetMarkers(ff, c("FSC-A", "CD3", "FITC-A"))
```

gg_color_hue	<i>gg_color_hue</i>
--------------	---------------------

Description

Helper function to get the ggplot colors

Usage

```
gg_color_hue(n)
```

Arguments

n Number of colors

Value

array with hexadecimal color values

GroupStats	<i>GroupStats</i>
------------	-------------------

Description

Calculate statistics between 2 groups based on the [GetFeatures](#) output

Usage

```
GroupStats(features, groups)
```

Arguments

features Feature matrix as generated by [GetFeatures](#), e.g. a percentages matrix
groups Named list with file or patient IDs per group (should match with the rownames of the matrix).

Value

array containing the selected selected rows

Examples

```
points <- matrix(1:1000, ncol = 10)
selection <- Initialize_KWSP(points, 3, 3)
```

Initialize_PCA

Create a grid from first 2 PCA components

Description

Create a grid from first 2 PCA components

Usage

```
Initialize_PCA(data, xdim, ydim)
```

Arguments

data	matrix in which each row represents a point
xdim	x dimension of the grid
ydim	y dimension of the grid

Value

array containing the selected selected rows

Examples

```
points <- matrix(1:1000, ncol = 10)
selection <- Initialize_PCA(points, 3, 3)
```

ManualVector

Summarize the gating matrix into one vector, only including the cell types of interest

Description

Extract the compensated and transformed data and all gate labels.

Usage

```
ManualVector(manualMatrix, cellTypes)
```

Arguments

manualMatrix	Matrix containing boolean values, indicating for every gate (column) whether the cell (row) is part of it or not.
cellTypes	Cell types to use in the summary vector. All others will be ignored and cells which do not fall in one of these gates will get the label "Unknown". Order is important!

Value

A factor with one label for every cell

MapDataToCodes	<i>Assign nearest node to each datapoint</i>
----------------	--

Description

Assign nearest node to each datapoint

Usage

```
MapDataToCodes(codes, newdata, distf = 2)
```

Arguments

codes	matrix with nodes of the SOM
newdata	datapoints to assign
distf	Distance function (1 = manhattan, 2 = euclidean, 3 = chebyshev, 4 = cosine)

Value

Array with nearest node id for each datapoint

MetaclusterCVs	<i>MetaclusterCVs</i>
----------------	-----------------------

Description

Compute the coefficient of variation for the metaclusters

Usage

```
MetaclusterCVs(fsom)
```

Arguments

fsom	Result of calling the FlowSOM function
------	--

Value

Metacluster CVs

Examples

```

fileName <- system.file("extdata", "68983.fcs", package="FlowSOM")
ff <- flowCore::read.FCS(fileName)
ff <- flowCore::compensate(ff,ff@description$SPILL)
ff <- flowCore::transform(ff,
  flowCore::transformList(colnames(ff@description$SPILL),
    flowCore::logicTransform()))
flowSOM.res <- FlowSOM(ff,scale=TRUE,colsToUse=c(9,12,14:18), nClus=10)
cvs <- GetMetaclusterCVs(flowSOM.res)

```

MetaClustering

*MetaClustering***Description**

Cluster data with automatic number of cluster determination for several algorithms

Usage

```
MetaClustering(data, method, max = 20, seed = NULL, ...)
```

Arguments

data	Matrix containing the data to cluster
method	Clustering method to use
max	Maximum number of clusters to try out
seed	Seed to pass on to given clustering method
...	Extra parameters to pass along

Value

Numeric array indicating cluster for each datapoint

See Also

[metaClustering_consensus](#)

Examples

```

# Read from file, build self-organizing map and minimal spanning tree
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
flowSOM.res <- ReadInput(fileName, compensate = TRUE, transform = TRUE,
  scale = TRUE)
flowSOM.res <- BuildSOM(flowSOM.res,colsToUse = c(9, 12, 14:18))
flowSOM.res <- BuildMST(flowSOM.res)

# Apply metaclustering
metacl <- MetaClustering(flowSOM.res$map$codes,
  "metaClustering_consensus",
  max = 10)

# Get metaclustering per cell
flowSOM.clustering <- metacl[flowSOM.res$map$mapping[, 1]]

```

```
metaClustering_consensus
```

MetaClustering

Description

Cluster data using hierarchical consensus clustering with k clusters

Usage

```
metaClustering_consensus(data, k = 7, seed = NULL)
```

Arguments

data	Matrix containing the data to cluster
k	Number of clusters
seed	Seed to pass to consensusClusterPlus

Value

Numeric array indicating cluster for each datapoint

See Also

[MetaClustering](#)

Examples

```
# Read from file, build self-organizing map and minimal spanning tree
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
flowSOM.res <- ReadInput(fileName, compensate = TRUE, transform = TRUE,
                        scale = TRUE)
flowSOM.res <- BuildSOM(flowSOM.res, colsToUse = c(9, 12, 14:18))
flowSOM.res <- BuildMST(flowSOM.res)

# Apply consensus metaclustering
metacl <- metaClustering_consensus(flowSOM.res$map$codes, k = 10)
```

```
MetaclusterMFIs
```

MetaclusterMFIs

Description

Compute the median fluorescence intensities for the metaclusters

Usage

```
MetaclusterMFIs(fsom)
```

Arguments

fsom Result of calling the FlowSOM function

Value

Metacluster MFIs

Examples

```
fileName <- system.file("extdata", "68983.fcs", package="FlowSOM")
ff <- flowCore::read.FCS(fileName)
ff <- flowCore::compensate(ff, ff@description$SPILL)
ff <- flowCore::transform(ff,
  flowCore::transformList(colnames(ff@description$SPILL),
    flowCore::logicleTransform()))
flowSOM.res <- FlowSOM(ff, scale=TRUE, colsToUse=c(9,12,14:18), maxMeta=10)
mfis <- GetMetaclusterMFIs(flowSOM.res)
```

NClusters	<i>NClusters</i>
-----------	------------------

Description

Extracts the number of clusters from a FlowSOM object

Usage

```
NClusters(fsom)
```

Arguments

fsom FlowSOM object

Value

The number of clusters

Examples

```
# Build FlowSom result
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
ff <- flowCore::read.FCS(fileName)
flowSOM.res <- FlowSOM(ff,
  compensate = TRUE, transform = TRUE, scale = TRUE,
  colsToUse = c(9, 12, 14:18),
  maxMeta = 10)
NClusters(flowSOM.res)
```

NewData	<i>NewData</i>
---------	----------------

Description

Map new data to a FlowSOM grid

Usage

```
NewData(
  fsom,
  input,
  madAllowed = 4,
  compensate = NULL,
  spillover = NULL,
  transform = NULL,
  toTransform = NULL,
  transformFunction = NULL,
  transformList = NULL,
  scale = NULL,
  scaled.center = NULL,
  scaled.scale = NULL,
  silent = FALSE
)
```

Arguments

fsom	FlowSOM object
input	A flowFrame, a flowSet or an array of paths to files or directories
madAllowed	A warning is generated if the distance of the new data points to their closest cluster center is too big. This is computed based on the typical distance of the points from the original dataset assigned to that cluster, the threshold being set to median + madAllowed * MAD. Default is 4.
compensate	logical, does the data need to be compensated. If NULL, the same value as in the original FlowSOM call will be used.
spillover	spillover matrix to compensate with. If NULL, the same value as in the original FlowSOM call will be used.
transform	logical, does the data need to be transformed. If NULL, the same value as in the original FlowSOM call will be used.
toTransform	column names or indices that need to be transformed. If NULL, the same value as in the original FlowSOM call will be used.
transformFunction	If NULL, the same value as in the original FlowSOM call will be used.
transformList	If NULL, the same value as in the original FlowSOM call will be used.
scale	Logical, does the data needs to be rescaled. If NULL, the same value as in the original FlowSOM call will be used.
scaled.center	See scale . If NULL, the same value as in the original FlowSOM call will be used.

scaled.scale	See scale . If NULL, the same value as in the original FlowSOM call will be used.
silent	Logical. If TRUE, print progress messages. Default = FALSE.

Details

New data is mapped to an existing FlowSOM object. The input is similar to the [ReadInput](#) function. A new FlowSOM object is created, with the same grid, but a new mapping, node sizes and mean values. The same preprocessing steps (compensation, transformation and scaling) will happen to this file as was specified in the original FlowSOM call. The scaling parameters from the original grid will be used.

Value

A new FlowSOM object

See Also

[FlowSOMSubset](#) if you want to get a subset of the current data instead of a new dataset

Examples

```
# Build FlowSom result
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
ff <- flowCore::read.FCS(fileName)
ff <- flowCore::compensate(ff, flowCore::keyword(ff)[["SPILL"]])
ff <- flowCore::transform(ff,
  flowCore::transformList(colnames(flowCore::keyword(ff)[["SPILL"]]),
    flowCore::logicleTransform()))
flowSOM.res <- FlowSOM(ff[1:1000, ],
  scale = TRUE,
  colsToUse = c(9, 12, 14:18),
  nClus = 10)

# Map new data
fSOM2 <- NewData(flowSOM.res, ff[1001:2000, ])
```

NMetaclusters

NMetaclusters

Description

Extracts the number of metaclusters from a FlowSOM object

Usage

```
NMetaclusters(fsom)
```

Arguments

fsom	FlowSOM object
------	----------------

Value

The number of metaclusters

Examples

```
# Build FlowSom result
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
ff <- flowCore::read.FCS(fileName)
flowSOM.res <- FlowSOM(ff,
  compensate = TRUE, transform = TRUE, scale = TRUE,
  colsToUse = c(9, 12, 14:18),
  maxMeta = 10)
NMetaclusters(flowSOM.res)
```

ParseArcs	<i>ParseArcs</i>
-----------	------------------

Description

Parses stars

Usage

ParseArcs(x, y, arcValues, arcHeights)

Arguments

- x x coordinate of node
- y y coordinate of node
- arcValues A named vector with the frequency of how the node should be divided
- arcHeights The heights of the arcs

Details

Function that parses the FlowSOM object into a dataframe for the star values for ggplot

Value

A dataframe ready to use with ggplot, consisting of the coordinates of centers, the radius and angles of the star values

See Also

[PlotFlowSOM](#), [ParseEdges](#), [ParseNodeSize](#), [ParseQuery](#), [ParseSD](#)

ParseEdges	<i>ParseEdges</i>
------------	-------------------

Description

Parses edges

Usage

ParseEdges(fsom)

Arguments

fsom FlowSOM object, as generated by [FlowSOM](#)

Details

Function that parses the graph edges of the FlowSOM object into a dataframe

Value

A dataframe consisting of start and end coordinates of edges

See Also

[PlotFlowSOM](#), [ParseNodeSize](#), [ParseArcs](#), [ParseQuery](#), [ParseSD](#), [AddMST](#)

ParseLayout	<i>ParseLayout</i>
-------------	--------------------

Description

ParseLayout

Usage

ParseLayout(fsom, layout)

Arguments

fsom FlowSOM object
layout "MST", "grid" or a matrix/dataframe with 2 columns and 1 row per cluster

Value

dataframe with 2 columns and 1 row per cluster

ParseNodeSize	<i>ParseNodeSize</i>
---------------	----------------------

Description

Parses node size

Usage

ParseNodeSize(nodeSizes, maxNodeSize, refNodeSize)

Arguments

- | | |
|-------------|--|
| nodeSizes | A vector with node sizes |
| maxNodeSize | Determines the maximum node size. |
| refNodeSize | Reference for node size against which the nodeSizes will be scaled. Default = max(nodeSizes) |

Details

Function that parses the mapping of the FlowSOM object into node sizes relative to the abundances of cells per cluster
Scales node size relative to the abundances of cells per cluster

Value

A vector is returned consisting of node sizes

See Also

[PlotFlowSOM](#), [ParseEdges](#), [AutoMaxNodeSize](#), [ParseArcs](#), [ParseQuery](#), [ParseSD](#)

ParseQuery	<i>ParseQuery</i>
------------	-------------------

Description

Parses query

Usage

ParseQuery(fsom, query)

Arguments

- | | |
|-------|--|
| fsom | FlowSOM object, as generated by FlowSOM |
| query | Array containing "high" or "low" for the specified column names of the Flow-SOM data |

Details

Identify nodes in the tree which resemble a certain profile of "high" or "low" marker expressions.

Value

A list, containing the ids of the selected nodes, the individual scores for all nodes and the scores for each marker for each node

See Also

[PlotFlowSOM](#), [ParseEdges](#), [ParseNodeSize](#), [ParseArcs](#), [QueryStarPlot](#), [ParseSD](#)

ParseSD	<i>ParseSD Parses SD in FlowSOM object</i>
---------	--

Description

Calculates the standard deviation of a FlowSOM object

Usage

```
ParseSD(fsom, marker = NULL)
```

Arguments

fsom	FlowSOM object, as generated by FlowSOM
marker	If a marker is given, the standard deviation for this marker is shown. Otherwise, the maximum ratio is used.

Value

A vector containing the SDs

See Also

[PlotFlowSOM](#), [ParseEdges](#), [ParseNodeSize](#), [ParseArcs](#), [ParseQuery](#), [PlotSD](#)

Plot2DScatters

Plot2DScatters

Description

Function to draw 2D scatter plots of FlowSOM (meta)clusters

Usage

```
Plot2DScatters(
  fsom,
  channelpairs,
  clusters = NULL,
  metaclusters = NULL,
  maxBgPoints = 3000,
  sizeBgPoints = 0.5,
  maxPoints = 1000,
  sizePoints = 0.5,
  xlim = NULL,
  ylim = NULL,
  xyLabels = c("marker"),
  density = TRUE,
  centers = TRUE,
  colors = NULL,
  plotFile = "2DScatterPlots.png"
)
```

Arguments

fsom	FlowSOM object, as created by FlowSOM
channelpairs	List in which each element is a pair of channel or marker names
clusters	Vector or list (to combine multiple clusters in one plot) with indices of clusters of interest
metaclusters	Vector or list (to combine multiple metaclusters in one plot) with indices of metaclusters of interest
maxBgPoints	Maximum number of background cells to plot
sizeBgPoints	Size of the background cells
maxPoints	Maximum number of (meta)cluster cells to plot
sizePoints	Size of the (meta)cluster cells
xlim	Optional vector of a lower and upper limit of the x-axis
ylim	Optional vector of a lower and upper limit of the y-axis
xyLabels	Determines the label of the x- and y-axis. Can be "marker" and/or "channel" or abbreviations. Default = "marker".
density	Default is TRUE to color the (meta)cluster points according to density. Set to FALSE to use a plain color
centers	Default is TRUE to show the cluster centers

Usage

```
PlotCenters(fsom, marker1, marker2, MST = TRUE)
```

Arguments

fsom	FlowSOM object, as generated by BuildMST
marker1	Marker to show on the x-axis
marker2	Marker to show on the y-axis
MST	Type of visualization, if 1 plot tree, else plot grid

Details

Plot FlowSOM nodes on a 2D scatter plot of the data

Value

Nothing is returned. A 2D scatter plot is drawn on which the nodes of the grid are indicated

See Also

[PlotStars](#), [PlotPies](#), [PlotMarker](#), [BuildMST](#)

Examples

```
# Read from file, build self-organizing map and minimal spanning tree
fileName <- system.file("extdata", "68983.fcs", package="FlowSOM")
flowSOM.res <- ReadInput(fileName, compensate=TRUE, transform=TRUE,
                          scale=TRUE)
flowSOM.res <- BuildSOM(flowSOM.res, colsToUse=c(9,12,14:18))
flowSOM.res <- BuildMST(flowSOM.res)

# Plot centers
plot <- Plot2DScatters(flowSOM.res,
                      channelpairs = list(c("FSC-A", "SSC-A")),
                      clusters = list(seq_len(NClusters(flowSOM.res))),
                      maxPoints = 0,
                      plotFile = NULL)
```

PlotClusters2D

PlotClusters2D

Description

Plot nodes on scatter plot

Examples

```
## Deprecated - use Plot2DScatters instead ##

# Read from file, build self-organizing map and minimal spanning tree
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
flowSOM.res <- ReadInput(fileName, compensate = TRUE, transform = TRUE,
                        scale = TRUE)
flowSOM.res <- BuildSOM(flowSOM.res, colsToUse = c(9, 12, 14:18))
flowSOM.res <- BuildMST(flowSOM.res)

# Plot cells
## Not run:
Plot2DScatters(flowSOM.res, c(1, 2), clusters = 91)

## End(Not run)
```

PlotDimRed

PlotDimRed

Description

Plot a dimensionality reduction

Usage

```
PlotDimRed(
  fsom,
  colsToUse = fsom$map$colsUsed,
  colorBy = "metaclusters",
  colors = NULL,
  lim = NULL,
  cTotal = NULL,
  dimred = Rtsne::Rtsne,
  extractLayout = function(dimred) {
    dimred$Y
  },
  label = TRUE,
  returnLayout = FALSE,
  seed = NULL,
  title = NULL,
  ...
)
```

Arguments

<code>fsom</code>	FlowSOM object, as generated by BuildMST
<code>colsToUse</code>	The columns used for the dimensionality reduction. Default = <code>fsom\$map\$colsUsed</code> .
<code>colorBy</code>	Defines how the dimensionality reduction will be colored. Can be "metaclusters" (default), "clusters" (or abbreviations) or a marker/channel/index.

colors	A vector of custom colors. Default returns ggplot colors for categorical variables and the FlowSOM colors for continuous variables. When using a categorical variable, the vector must be as long as the levels of the categorical variable.
lim	Limits for the colorscale
cTotal	The total amount of cells to be used in the dimensionality reduction. Default is all the cells.
dimred	A dimensionality reduction function. Default = Rtsne::Rtsne. Alternatively, a data.frame or matrix with either equal number of rows to the fsom or an OriginalID column. Recommended to put cTotal to NULL when providing a matrix (or ensuring that the dimred corresponds to subsampling the flowSOM data for cTotal cells with the same seed).
extractLayout	A function to extract the coordinates from the results of the dimred default = function(dimred)dimred\$Y.
label	If label = TRUE (default), labels are added to plot.
returnLayout	If TRUE, this function returns a dataframe with the layout of dimred and the original IDs and the plot. Default = FALSE.
seed	A seed for reproducibility.
title	A title for the plot.
...	Additional arguments to pass to dimred.

Details

Plot a dimensionality reduction of fsom\$data

Value

A dimensionality reduction plot made in ggplot2

Examples

```
file <- system.file("extdata", "68983.fcs", package="FlowSOM")
flowSOM.res <- FlowSOM(file, compensate = TRUE, transform = TRUE,
  scale = TRUE,
  colsToUse = c(9, 12, 14:18), nClus = 10, silent = FALSE,
  xdim = 7, ydim = 7)
PlotDimRed(flowSOM.res, cTotal = 5000, seed = 1, title = "t-SNE")
PlotDimRed(flowSOM.res, cTotal = 5000, colorBy = "CD3", seed = 1,
  title = "t-SNE")
```

PlotFileScatters

PlotFileScatters

Description

Make a scatter plot per channel for all provided files

Usage

```

PlotFileScatters(
  input,
  fileID = "File",
  channels = NULL,
  yLim = NULL,
  yLabel = "marker",
  quantiles = NULL,
  names = NULL,
  groups = NULL,
  color = NULL,
  legend = FALSE,
  maxPoints = 50000,
  ncol = NULL,
  nrow = NULL,
  width = NULL,
  height = NULL,
  silent = FALSE,
  plotFile = "FileScatters.png"
)

```

Arguments

input	Either a flowSet, a flowFrame with a file ID column (e.g. output from the AggregateFlowFrames includes a "File" column) or a vector of paths pointing to FCS files
fileID	Name of the file ID column when the input is a flowFrame, default to "File" (File ID column in the AggregateFlowFrames flowFrame output).
channels	Vector of channels or markers that need to be plotted, if NULL (default), all channels from the input will be plotted
yLim	Optional vector of a lower and upper limit of the y-axis
yLabel	Determines the label of the y-axis. Can be "marker" and/or "channel" or abbreviations. Default = "marker".
quantiles	If provided (default NULL), a numeric vector with values between 0 and 1. These quantiles are indicated on the plot
names	Optional parameter to provide filenames. If NULL (default), the filenames will be numbers. Duplicated filenames will be made unique.
groups	Optional parameter to specify groups of files, should have the same length as the input. If NULL (default), all files will be plotted in the same color
color	Optional parameter to provide colors. Should have the same lengths as the number of groups (or 1 if groups is NULL)
legend	Logical parameter to specify whether the group levels should be displayed. Default is FALSE
maxPoints	Total number of data points that will be plotted per channel, default is 50000
ncol	Number of columns in the final plot, optional
nrow	Number of rows in the final plot, optional
width	Width of png file. By default NULL the width parameter is estimated based on the input.

Arguments

<code>fsom</code>	FlowSOM object, as created by FlowSOM
<code>view</code>	Preferred view, options: "MST", "grid" or "matrix" with a matrix/dataframe consisting of coordinates. Default = "MST"
<code>nodeSizes</code>	A vector containing node sizes. These will automatically be scaled between 0 and <code>maxNodeSize</code> and transformed with a sqrt. Default = <code>fsom\$MST\$sizes</code>
<code>maxNodeSize</code>	Determines the maximum node size. Default is 1.
<code>refNodeSize</code>	Reference for node size against which the <code>nodeSizes</code> will be scaled. Default = <code>max(nodeSizes)</code>
<code>equalNodeSize</code>	If TRUE, the nodes will be equal to <code>maxNodeSize</code> . If FALSE (default), the nodes will be scaled to the number of cells in each cluster
<code>backgroundValues</code>	Values to be used for background coloring, either numerical values or something that can be made into a factor (e.g. a clustering)
<code>backgroundColors</code>	Color palette to be used for the background coloring. Can be either a function or an array specifying colors.
<code>backgroundLim</code>	Only used when <code>backgroundValues</code> are numerical. Defaults to min and max of the <code>backgroundValues</code> .
<code>title</code>	Title of the plot

Details

Base layer of the FlowSOM plot, where you can choose layout (MST, grid or coordinates of your own choosing), background colors and node size. Can then be extended by e.g. [AddStars](#), [AddLabels](#), [AddPies](#), ...

Value

A ggplot object with the base layer of a FlowSOM plot

See Also

[PlotStars](#), [PlotVariable](#), [PlotMarker](#), [PlotLabels](#), [PlotNumbers](#), [PlotPies](#), [QueryStarPlot](#), [PlotSD](#)

Examples

```
# Locate file on file system
fcs_file <- system.file("extdata", "68983.fcs", package = "FlowSOM")

# Build FlowSOM model
flowSOM.res <- FlowSOM(fcs_file,
  scale = TRUE,
  compensate = TRUE,
  transform = TRUE,
  toTransform = 8:18,
  colsToUse = c(9, 12, 14:18),
  nClus = 10,
  seed = 1)

# Plot with background coloring
```

```
PlotFlowSOM(flowSOM.res,
             backgroundValues = flowSOM.res$metaclustering) %>%
AddLabels(seq(100))
```

PlotGroups	<i>PlotGroups</i>
------------	-------------------

Description

Plot differences between groups

Usage

```
PlotGroups(fsom, groups, threshold = NULL, pThreshold = 0.05, ...)
```

Arguments

fsom	FlowSOM object, as generated by BuildMST
groups	Groups result as generated by CountGroups
threshold	Relative difference in groups before the node is colored
pThreshold	Threshold on p-value from wilcox-test before the node is colored. If this is not NULL, threshold will be ignored.
...	Additional arguments to pass to PlotFlowSOM

Details

Plot FlowSOM trees, where each node is represented by a star chart indicating mean marker values, the size of the node is relative to the mean percentage of cells present in each

Value

A vector containing the labels assigned to the nodes for all groups except the first

See Also

[PlotStars](#), [PlotVariable](#), [PlotFlowSOM](#), [PlotLabels](#), [PlotNumbers](#), [PlotMarker](#), [PlotPies](#), [QueryStarPlot](#), [PlotSD](#)

Examples

```
#Run FlowSOM
fileName <- system.file("extdata", "68983.fcs", package="FlowSOM")
fsom <- FlowSOM(fileName, compensate = TRUE, transform = TRUE,
                scale = TRUE, colsToUse = c(9,12,14:18), nClus = 10)

ff <- flowCore::read.FCS(fileName)
# Make an additional file without cluster 7 and double amount of cluster 5
selection <- c(which(GetClusters(fsom) %in% which(fsom$metaclustering != 7)),
               which(GetClusters(fsom) %in% which(fsom$metaclustering == 5)))
ff_tmp <- ff[selection,]
```

```

flowCore::write.FCS(ff_tmp, file="ff_tmp.fcs")

# Compare only the file with the double amount of cluster 10
features <- GetFeatures(fsom,
  c(fileName, "ff_tmp.fcs"),
  level = "clusters",
  type = "percentages")
stats <- GroupStats(features$cluster_percentages,
  groups = list("AllCells" = c(fileName),
    "Without_ydTcells" = c("ff_tmp.fcs")))

fold_changes <- stats["fold changes", ]
fold_changes_label <- factor(ifelse(fold_changes < -1.5,
  "Underrepresented compared to Group 1",
  ifelse(fold_changes > 1.5,
    "Overrepresented compared to Group 1",
    "--")),
  levels = c("--",
    "Underrepresented compared to Group 1",
    "Overrepresented compared to Group 1"))
fold_changes_label[is.na(fold_changes_label)] <- "--"
gr_1 <- PlotStars(fsom,
  title = "All Cells",
  nodeSizes = stats["medians AllCells", ],
  list_insteadof_ggarrange = TRUE)
gr_2 <- PlotStars(fsom, title = "Group 2",
  nodeSizes = stats["medians Without_ydTcells", ],
  backgroundValues = fold_changes_label,
  backgroundColors = c("white", "red", "blue"),
  list_insteadof_ggarrange = TRUE)
p <- ggpubr::ggarrange(plotlist = c(list(gr_1$tree), gr_2),
  heights = c(3, 1))
p

```

PlotLabels

PlotLabels

Description

Plot labels for each cluster

Usage

```

PlotLabels(
  fsom,
  labels,
  maxNodeSize = 0,
  textSize = 3.88,
  textColor = "black",
  ...
)

```


Usage

```
PlotManualBars(
  fsom,
  fcs = NULL,
  manualVector,
  manualOrder = NULL,
  colors = NULL,
  list_insteadof_plots = FALSE
)
```

Arguments

<code>fsom</code>	FlowSOM object, as generated by FlowSOM or by NewData . The clusters and metaclusters will be plotted in the order of the factor levels.
<code>fcs</code>	FCS file that should be mapped on the FlowSOM object. Default is NULL.
<code>manualVector</code>	Vector with cell labels, e.g. obtained by manual gating
<code>manualOrder</code>	Optional vector with unique cell labels to fix in which order the cell labels should be shown
<code>colors</code>	Optional color vector, should have the same length as the number of unique cell labels
<code>list_insteadof_plots</code>	If FALSE (default), it returns multiple plots. If TRUE, it returns a list of ggplot objects

Value

Either a plot or a ggplot objects list is returned.

Examples

```
# Identify the files
fcs_file <- system.file("extdata", "68983.fcs", package = "FlowSOM")
gating_file <- system.file("extdata", "gatingResult.csv", package = "FlowSOM")

# Specify the cell types of interest for assigning one label per cell
cellTypes <- c("B cells",
               "gd T cells", "CD4 T cells", "CD8 T cells",
               "NK cells", "NK T cells")

# Load manual labels (e.g. GetFlowJoLabels can be used to extract labels from
# an fcs file)

gatingResult <- as.factor(read.csv(gating_file, header = FALSE)[, 1])

# Build a FlowSOM object
flowSOM.res <- FlowSOM(fcs_file,
  scale = TRUE,
  compensate = TRUE,
  transform = TRUE,
  toTransform = 8:18,
  colsToUse = c(9, 12, 14:18),
  nClus = 10,
  seed = 1)
```



```
      "#7FFF7F", "yellow", "#FF7F00", "red", "#7F0000")),
  main = paste0("Cluster ", id)
)
```

Arguments

<code>fsom</code>	FlowSOM object, as generated by BuildMST or the first element of the list returned by FlowSOM
<code>id</code>	Id of the node to plot (check PlotNumbers to get the ids)
<code>markers</code>	Array of markers to use. Default: the markers used to build the tree
<code>colorPalette</code>	Color palette to be used for the markers
<code>main</code>	Title of the plot

Value

Nothing is returned. A plot is drawn in which the node is represented by a star chart indicating the median fluorescence intensities.

See Also

[PlotStars](#), [PlotNumbers](#), [FlowSOM](#)

Examples

```
# Read from file, build self-organizing map and minimal spanning tree
fileName <- system.file("extdata", "68983.fcs", package="FlowSOM")
flowSOM.res <- FlowSOM(fileName, compensate=TRUE, transform=TRUE,
  scale=TRUE, colsToUse=c(9,12,14:18), nClus=10)

# Deprecated, it is currently not possible anymore to plot an individual
# node alone. If necessary, zooming in on a node can be approximated by
# exaggerating the size of the node.
PlotStars(flowSOM.res, nodeSizes = c(100, rep(0,99)), maxNodeSize = 10)
```

PlotNumbers

PlotNumbers

Description

Plot cluster ids for each cluster

Usage

```
PlotNumbers(fsom, level = "clusters", maxNodeSize = 0, ...)
```

PlotOutliers*PlotOutliers*

Description

Visual overview of outliers

Usage

```
PlotOutliers(fsom, outlierReport)
```

Arguments

fsom FlowSOM object.
outlierReport Outlier overview as generated by TestOutliers()

Value

Plot

Examples

```
# Identify the files
fcs <- flowCore::read.FCS(system.file("extdata", "68983.fcs",
                                     package = "FlowSOM"))

# Build a FlowSOM object
flowSOM.res <- FlowSOM(fcs,
                      scale = TRUE,
                      compensate = TRUE,
                      transform = TRUE,
                      toTransform = 8:18,
                      colsToUse = c(9, 12, 14:18),
                      nClus = 10,
                      seed = 1)
outlierReport <- TestOutliers(flowSOM.res)
p <- PlotOutliers(flowSOM.res, outlierReport)
```

PlotOverview2D*PlotOverview2D*

Description

Plot metaclusters on scatter plots

Usage

```
PlotOverview2D(fsom, markerlist, metaclusters, colors = NULL, ff, ...)
```

Arguments

fson	FlowSOM object, as generated by FlowSOM . If using a FlowSOM object as generated by BuildMST , it needs to be wrapped in a list, list(FlowSOM = fson, metaclustering = metaclustering).
markerlist	List in which each element is a pair of marker names
metaclusters	Metaclusters of interest
colors	Named vector with color value for each metacluster. If NULL (default) color-brewer "paired" is interpolated
ff	flowFrame to use as reference for the marker names
...	Other parameters to pass on to PlotClusters2D

Details

Write multiple 2D scatter plots to a png file. All cells of fson\$data are plotted in black, and those of the selected metaclusters are plotted in color.

Value

Nothing is returned, but a plot is drawn for every markerpair and every metacluster. The individual cells are colored, and the center of each FlowSOM cluster is indicated with a blue cross.

See Also

[PlotClusters2D](#)

Examples

```
## Deprecated - use Plot2DScatters instead ##

# Read from file, build self-organizing map and minimal spanning tree
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
flowSOM.res <- FlowSOM(fileName,
                        compensate = TRUE, transform = TRUE, scale = TRUE,
                        colsToUse = c(9, 12, 14:18),
                        nClus = 10,
                        seed = 1)

# Plot cells
markers_of_interest = list(c("FSC-A", "SSC-A"),
                           c("CD3", "CD19"),
                           c("TCRb", "TCRyd"),
                           c("CD4", "CD8"))
metaclusters_of_interest = 1:10

# Recommended to write to png

## Not run:
png("Markeroverview.png",
    width = 500 * length(markers_of_interest),
    height = 500 * length(metaclusters_of_interest))
Plot2DScatters(flowSOM.res,
               channelpairs = markers_of_interest,
               metaclusters = metaclusters_of_interest)

dev.off()
```

```
## End(Not run)
```

PlotPies

PlotPies

Description

Plot comparison with other clustering

Usage

```
PlotPies(
  fsom,
  cellTypes,
  colorPalette = grDevices::colorRampPalette(c("white", "#00007F", "blue", "#007FFF",
    "cyan", "#7FFF7F", "yellow", "#FF7F00", "red", "#7F0000")),
  ...
)
```

Arguments

<code>fsom</code>	FlowSOM object, as generated by FlowSOM
<code>cellTypes</code>	Array of factors indicating the celltypes
<code>colorPalette</code>	Color palette to use.
<code>...</code>	Additional arguments to pass to PlotFlowSOM

Details

Plot FlowSOM grid or tree, with pies indicating another clustering or manual gating result

Value

ggplot plot

See Also

[PlotStars](#), [PlotVariable](#), [PlotFlowSOM](#), [PlotLabels](#), [PlotNumbers](#), [PlotMarker](#), [QueryStarPlot](#), [PlotSD](#)

Examples

```
# Identify the files
fcs_file <- system.file("extdata", "68983.fcs", package = "FlowSOM")
gating_file <- system.file("extdata", "gatingResult.csv", package = "FlowSOM")

# Specify the cell types of interest for assigning one label per cell
cellTypes <- c("B cells",
  "gd T cells", "CD4 T cells", "CD8 T cells",
  "NK cells", "NK T cells")
```

```
# Load manual labels (e.g. GetFlowJoLabels can be used to extract labels from
# an fcs file)

gatingResult <- as.factor(read.csv(gating_file, header = FALSE)[, 1])

# Build a FlowSOM tree
flowSOM.res <- FlowSOM(fcs_file,
                        scale = TRUE,
                        compensate = TRUE,
                        transform = TRUE,
                        toTransform = 8:18,
                        colsToUse = c(9, 12, 14:18),
                        nClus = 10,
                        seed = 1)

# Plot pies indicating the percentage of cell types present in the nodes
PlotPies(flowSOM.res,
          gatingResult,
          backgroundValues = flowSOM.res$metaclustering)
```

PlotSD	<i>PlotSD</i>
--------	---------------

Description

Plot FlowSOM grid or tree, colored by standard deviation.

Usage

```
PlotSD(fsom, marker = NULL, ...)
```

Arguments

fsom	FlowSOM object, as generated by FlowSOM
marker	If a marker/channel is given, the sd for this marker is shown. Otherwise, the maximum ratio is used.
...	Additional arguments to pass to PlotFlowSOM

Value

Nothing is returned. A plot is drawn in which each node is colored depending on its standard deviation

See Also

[PlotStars](#), [PlotVariable](#), [PlotFlowSOM](#), [PlotLabels](#), [PlotNumbers](#), [PlotMarker](#), [PlotPies](#), [QueryStarPlot](#)

Examples

```
# Read from file, build self-organizing map and minimal spanning tree
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
flowSOM.res <- ReadInput(fileName, compensate = TRUE, transform = TRUE,
                        scale = TRUE)
flowSOM.res <- BuildSOM(flowSOM.res, colsToUse = c(9, 12, 14:18))
flowSOM.res <- BuildMST(flowSOM.res)

PlotSD(flowSOM.res)
```

PlotStarLegend

PlotStarLegend

Description

Plots star legend

Usage

```
PlotStarLegend(markers, colors, starHeight = 1)
```

Arguments

markers	Vector of markers used in legend
colors	Color palette for the legend. Can be a vector or a function.
starHeight	Star height. Default = 1.

Details

Function makes the legend of the FlowSOM star plot

Value

Returns nothing, but plots a legend for FlowSOM star plot

See Also

[PlotFlowSOM](#)

Examples

```
PlotStarLegend(c("CD3", "CD4", "CD8"),
               FlowSOM_colors(3))
```

PlotStars

PlotStars

Description

Plot star charts

Usage

```
PlotStars(
  fsom,
  markers = fsom$map$colsUsed,
  colorPalette = FlowSOM_colors,
  list_insteadof_ggarrange = FALSE,
  ...
)
```

Arguments

fsom	FlowSOM object, as generated by BuildMST
markers	Markers to plot (will be parsed by GetChannels)
colorPalette	Color palette to use
list_insteadof_ggarrange	If FALSE (default), the plot and the legend are combined by ggarrange . If TRUE, the separate elements are returned in a list, to allow further customization.
...	Additional arguments to pass to PlotFlowSOM

Details

Plot FlowSOM grid or tree, where each node is represented by a star chart indicating median marker values

Value

Nothing is returned. A plot is drawn in which each node is represented by a star chart indicating the median fluorescence intensities. Resets the layout back to 1 plot at the end.

See Also

[PlotMarker](#), [PlotVariable](#), [PlotFlowSOM](#), [PlotLabels](#), [PlotNumbers](#), [PlotPies](#), [QueryStarPlot](#), [PlotSD](#)

Examples

```
# Read from file, build self-organizing map and minimal spanning tree
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
flowSOM.res <- FlowSOM(fileName, compensate = TRUE, transform = TRUE,
  scale = TRUE, colsToUse = c(9, 12, 14:18))

# Plot stars indicating the MFI of the cells present in the nodes
```

```

PlotStars(flowSOM.res, backgroundValues = flowSOM.res$metaclustering)

newLayout <- igraph::layout_with_fr(flowSOM.res[["MST"]][["graph"]])
PlotStars(flowSOM.res, backgroundValues = flowSOM.res$metaclustering,
          view = newLayout)

PlotStars(flowSOM.res, backgroundValues = flowSOM.res$metaclustering,
          view = "grid")

```

PlotVariable	<i>PlotVariable</i>
--------------	---------------------

Description

Plot a variable for all nodes

Usage

```

PlotVariable(
  fsom,
  variable,
  variableName = "",
  colorPalette = FlowSOM_colors,
  lim = NULL,
  ...
)

```

Arguments

fsom	FlowSOM object
variable	A vector containing a value for every cluster
variableName	Label to show on the legend
colorPalette	Color palette to use. Can be a function or a vector.
lim	Limits for the scale
...	Additional arguments to pass to PlotFlowSOM , e.g. view, backgroundValues, equalNodeSize ...

Details

Plot FlowSOM grid or tree, colored by node values given in variable

See Also

[PlotStars](#), [QueryStarPlot](#), [PlotFlowSOM](#), [PlotLabels](#), [PlotNumbers](#), [PlotMarker](#), [PlotPies](#), [PlotSD](#)

Examples

```
# Build FlowSOM model
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
flowSOM.res <- FlowSOM(fileName,
  compensate = TRUE, transform = TRUE, scale = FALSE,
  colsToUse = c(9, 12, 14:18),
  nClus = 10,
  seed = 1)

# Plot some random values
rand <- runif(flowSOM.res$map$nNodes)
PlotVariable(flowSOM.res,
  variable = rand,
  variableName = "Random")

PlotVariable(flowSOM.res,
  variable = flowSOM.res$metaclustering,
  variableName = "Metaclustering") %>%
  AddLabels(labels = flowSOM.res$metaclustering)
```

print.FlowSOM	<i>Print FlowSOM object</i>
---------------	-----------------------------

Description

Print FlowSOM object

Usage

```
## S3 method for class 'FlowSOM'
print(x, ...)
```

Arguments

x	FlowSOM object to print information about
...	Further arguments, not used

Examples

```
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
flowSOM.res <- FlowSOM(fileName, compensate = TRUE, transform = TRUE,
  scale = TRUE, colsToUse = c(9, 12, 14:18), nClus = 10)
print(flowSOM.res)
```

Purity	<i>Calculate mean weighted cluster purity</i>
--------	---

Description

Calculate mean weighted cluster purity

Usage

```
Purity(realClusters, predictedClusters, weighted = TRUE)
```

Arguments

realClusters	array with real cluster values
predictedClusters	array with predicted cluster values
weighted	logical. Should the mean be weighted depending on the number of points in the predicted clusters

Value

Mean purity score, worst score, number of clusters with score < 0.75

Examples

```
# Generate some random data as an example
realClusters <- sample(1:5, 100, replace = TRUE)
predictedClusters <- sample(1:6, 100, replace = TRUE)

# Calculate the FMeasure
Purity(realClusters, predictedClusters)
```

QueryMultiple	<i>QueryMultiple</i>
---------------	----------------------

Description

Function which takes a named list of multiple cell types, where every item is a named vector with values "high"/"low" and the names correspond to the markers or channels (e.g. as generated by `parse_markertable`).

Usage

```
QueryMultiple(fsom, cellTypes, plotFile = "queryMultiple.pdf", ...)
```

Arguments

<code>fsom</code>	FlowSOM object
<code>cellTypes</code>	Description of the cell types. Named list, with one named vector per cell type containing "high"/"low" values
<code>plotFile</code>	Path to a pdf file to save the plots (default is <code>queryMultiple.pdf</code>). If NULL, no plots will be generated
<code>...</code>	Additional arguments to pass to QueryStarPlot

Value

A label for every FlowSOM cluster (Unknown or one of the celltype names of the list, if selected by QueryStarPlot)

Examples

```
file <- system.file("extdata", "68983.fcs", package = "FlowSOM")
ff <- flowCore::read.FCS(file)
# Use the wrapper function to build a flowSOM object (saved in flowSOM.res)
# and a metaclustering (saved in flowSOM.res[["metaclustering"]])
flowSOM.res <- FlowSOM(ff, compensate = TRUE, transform = TRUE, scale = TRUE,
  colsToUse = c(9, 12, 14:18), nClus = 10, silent = FALSE,
  xdim = 7, ydim = 7)
cellTypes <- list("CD8 T cells" = c("PE-Cy7-A" = "high",
  "APC-Cy7-A" = "high",
  "Pacific Blue-A" = "high"),
  "B cells" = c("PE-Cy5-A" = "high"),
  "NK cells" = c("PE-A" = "high",
  "PE-Cy7-A" = "low",
  "APC-Cy7-A" = "low"))
query_res <- QueryMultiple(flowSOM.res, cellTypes, "query_multiple.pdf")
```

QueryStarPlot

QueryStarPlot

Description

Query a certain cell type

Usage

```
QueryStarPlot(
  fsom,
  query,
  plot = TRUE,
  colorPalette = FlowSOM_colors,
  backgroundColors = "#CA0020",
  ...
)
```

Arguments

fsom	FlowSOM object, as generated by BuildMST
query	Array containing "high" or "low" (or abbreviations) for the specified column names of the FlowSOM data.
plot	If true, a plot with a gradient of scores for the nodes is shown.
colorPalette	Color palette to be used for colors for "stars", "pies" or "marker". Can be either a function or an array specifying colors.
backgroundColors	Color to use for nodes with a high score in the plot. Default is red.
...	Additional arguments to pass to PlotFlowSOM

Details

Identify nodes in the tree which resemble a certain profile of "high" or "low" marker expressions.

Value

A list, containing the ids of the selected nodes, the individual scores for all nodes and the scores for each marker for each node

See Also

[PlotStars](#), [PlotVariable](#), [PlotFlowSOM](#), [PlotLabels](#), [PlotNumbers](#), [PlotMarker](#), [PlotPies](#), [PlotSD](#)

Examples

```
file <- system.file("extdata", "68983.fcs", package="FlowSOM")
flowSOM.res <- FlowSOM(file, compensate = TRUE, transform = TRUE,
  scale = TRUE, colsToUse = c(9, 12, 14:18), nClus = 10,
  silent = FALSE, xdim = 7, ydim = 7)
query <- c("CD3" = "high", #CD3
  "CD4" = "low", #TCRb
  "CD8" = "high") #CD8
query_res <- QueryStarPlot(flowSOM.res, query, equalNodeSize = TRUE)

cellTypes <- factor(rep("Unlabeled", 49),
  levels = c("Unlabeled", "CD8 T cells"))
cellTypes[query_res$selected] <- "CD8 T cells"
PlotStars(flowSOM.res,
  backgroundValues = cellTypes,
  backgroundColors = c("#FFFFFF00", "#ca0020aa"))
```

query_multiple

query_multiple

Description

Function which takes a named list of multiple cell types, where every item is a named vector with values "high"/"low" and the names correspond to the markers or channels (e.g. as generated by `parse_markertable`).

Usage

```
query_multiple(fsom, cell_types, pdf_name = "query_multiple.pdf", ...)
```

Arguments

<code>fsom</code>	FlowSOM object
<code>cell_types</code>	Description of the cell types. Named list, with one named vector per cell type containing "high"/"low" values
<code>pdf_name</code>	Path to a pdf file to save figures
<code>...</code>	Additional arguments to pass to QueryStarPlot

Value

A label for every FlowSOM cluster (Unknown or one of the celltype names of the list, if selected by QueryStarPlot)

See Also

[QueryStarPlot](#)

Examples

```
file <- system.file("extdata", "68983.fcs", package="FlowSOM")
ff <- flowCore::read.FCS(file)
# Use the wrapper function to build a flowSOM object (saved in flowSOM.res)
# and a metaclustering (saved in flowSOM.res[["metaclustering"]])
flowSOM.res <- FlowSOM(ff, compensate = TRUE, transform = TRUE, scale = TRUE,
  colsToUse = c(9,12,14:18), nClus = 10, silent = FALSE,
  xdim=7, ydim=7)
cell_types <- list("CD8 T cells" = c("PE-Cy7-A" = "high",
  "APC-Cy7-A" = "high",
  "Pacific Blue-A" = "high"),
  "B cells" = c("PE-Cy5-A" = "high"),
  "NK cells" = c("PE-A" = "high",
  "PE-Cy7-A" = "low",
  "APC-Cy7-A" = "low"))
query_res <- QueryMultiple(flowSOM.res, cell_types, "query_multiple.pdf")
```

ReadInput

Read FCS-files or flowFrames

Description

Take some input and return FlowSOM object containing a matrix with the preprocessed data (compensated, transformed, scaled)

Usage

```
ReadInput(
  input,
  pattern = ".fcs",
  compensate = FALSE,
  spillover = NULL,
  transform = FALSE,
  toTransform = NULL,
  transformFunction = flowCore::logicleTransform(),
  transformList = NULL,
  scale = FALSE,
  scaled.center = TRUE,
  scaled.scale = TRUE,
  silent = FALSE
)
```

Arguments

<code>input</code>	a <code>flowFrame</code> , a <code>flowSet</code> , a matrix with column names or an array of paths to files or directories
<code>pattern</code>	if <code>input</code> is an array of file- or directorynames, select only files containing <code>pattern</code>
<code>compensate</code>	logical, does the data need to be compensated
<code>spillover</code>	spillover matrix to compensate with If <code>NULL</code> and <code>compensate = TRUE</code> , we will look for <code>\$SPILL</code> description in FCS file.
<code>transform</code>	logical, does the data need to be transformed
<code>toTransform</code>	column names or indices that need to be transformed. Will be ignored if <code>transformList</code> is given. If <code>NULL</code> and <code>transform = TRUE</code> , column names of <code>\$SPILL</code> description in FCS file will be used.
<code>transformFunction</code>	Defaults to <code>logicleTransform()</code>
<code>transformList</code>	<code>transformList</code> to apply on the samples.
<code>scale</code>	logical, does the data needs to be rescaled
<code>scaled.center</code>	see scale
<code>scaled.scale</code>	see scale
<code>silent</code>	if <code>TRUE</code> , no progress updates will be printed. Default = <code>FALSE</code>

Value

FlowSOM object containing the data, which can be used as input for the `BuildSOM` function

See Also

[scale](#), [BuildSOM](#)

Examples

```
# Read from file
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
flowSOM.res <- ReadInput(fileName, compensate = TRUE, transform = TRUE,
  scale = TRUE)
```

```

# Or read from flowFrame object
ff <- flowCore::read.FCS(fileName)
ff <- flowCore::compensate(ff, flowCore::keyword(ff)[["SPILL"]])
ff <- flowCore::transform(ff,
  flowCore::transformList(colnames(flowCore::keyword(ff)[["SPILL"]]),
    flowCore::logicTransform()))
flowSOM.res <- ReadInput(ff, scale = TRUE)

# Build the self-organizing map and the minimal spanning tree
flowSOM.res <- BuildSOM(flowSOM.res, colsToUse = c(9, 12, 14:18))
flowSOM.res <- BuildMST(flowSOM.res)

# Apply metaclustering
metacl <- MetaClustering(flowSOM.res$map$codes,
  "metaClustering_consensus", max = 10)

# Get metaclustering per cell
flowSOM.clustering <- metacl[flowSOM.res$map$mapping[, 1]]

```

SaveClustersToFCS

Write FlowSOM clustering results to the original FCS files

Description

Write FlowSOM clustering results to the original FCS files

Usage

```

SaveClustersToFCS(
  fsom,
  originalFiles,
  preprocessedFiles = NULL,
  selectionColumn = NULL,
  silent = FALSE,
  outputDir = ".",
  suffix = "_FlowSOM.fcs",
  ...
)

```

Arguments

<code>fsom</code>	FlowSOM object as generated by BuildSOM
<code>originalFiles</code>	FCS files that should be extended
<code>preprocessedFiles</code>	FCS files that correspond to the input of FlowSOM, If NULL (default), the originalFiles are used.
<code>selectionColumn</code>	Column of the FCS file indicating the original cell ids. If NULL (default), no selection is made.
<code>silent</code>	If FALSE (default), print some extra output

outputDir	Directory to save the FCS files. Default to the current working directory (".")
suffix	Suffix added to the filename. Default _FlowSOM.fcs
...	Options to pass on to the read.FCS function (e.g. truncate_max_range)

Value

Saves the extended FCS file as [originalName]_FlowSOM.fcs

Examples

```
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
flowSOM.res <- FlowSOM(fileName, compensate = TRUE, transform = TRUE,
                      scale = TRUE, colsToUse = c(9, 12, 14:18), nClus = 10)
SaveClustersToFCS(flowSOM.res, fileName)
```

ScaleStarHeights	<i>ScaleStarHeights</i>
------------------	-------------------------

Description

Scales starheights

Usage

```
ScaleStarHeights(data, nodeSizes)
```

Arguments

data	Median values of relevant markers extracted from FlowSOM object
nodeSizes	A vector that is returned from ParseNodeSize

Details

Function that scales the star values between 0 and the node size

Value

A dataframe consisting of the scaled values of the stars. The stars are scaled between 0 and the maximum of all stars

See Also

[PlotFlowSOM](#), [ParseNodeSize](#), [AutoMaxNodeSize](#)

SOM

*Build a self-organizing map***Description**

Build a self-organizing map

Usage

```
SOM(
  data,
  xdim = 10,
  ydim = 10,
  rlen = 10,
  mst = 1,
  alpha = c(0.05, 0.01),
  radius = stats::quantile(nhbrdist, 0.67) * c(1, 0),
  init = FALSE,
  initf = Initialize_KWSP,
  distf = 2,
  silent = FALSE,
  map = TRUE,
  codes = NULL,
  importance = NULL
)
```

Arguments

data	Matrix containing the training data
xdim	Width of the grid
ydim	Hight of the grid
rlen	Number of times to loop over the training data for each MST
mst	Number of times to build an MST
alpha	Start and end learning rate
radius	Start and end radius
init	Initialize cluster centers in a non-random way
initf	Use the given initialization function if init == T (default: Initialize_KWSP)
distf	Distance function (1 = manhattan, 2 = euclidean, 3 = chebyshev, 4 = cosine)
silent	If FALSE, print status updates
map	If FALSE, data is not mapped to the SOM. Default TRUE.
codes	Cluster centers to start with
importance	array with numeric values. Parameters will be scaled according to importance

Value

A list containing all parameter settings and results

References

This code is strongly based on the kohonen package. R. Wehrens and L.M.C. Buydens, Self- and Super-organising Maps in R: the kohonen package J. Stat. Softw., 21(5), 2007

See Also

[BuildSOM](#)

TestOutliers	<i>TestOutliers</i>
--------------	---------------------

Description

Test if any cells are too far from their cluster centers

Usage

```
TestOutliers(
  fsom,
  madAllowed = 4,
  fsomReference = NULL,
  plotFile = NULL,
  channels = NULL
)
```

Arguments

fsom	FlowSOM object
madAllowed	Number of median absolute deviations allowed. Default = 4.
fsomReference	FlowSOM object to use as reference. If NULL (default), the original fsom object is used.
plotFile	If NULL (default), no plot will be created. If a filepath is given for a pdf, the plot will be written in the corresponding file
channels	If channels are given, the number of outliers in the original space for those channels will be calculated and added to the final results table.

Details

For every cluster, the distance from the cells to the cluster centers is used to label cells which deviate too far as outliers. The threshold is chosen as the median distance + madAllowed times the median absolute deviation of the distances.

Value

An outlier report

See Also

[FlowSOMSubset](#) if you want to get a subset of the current data instead of a new dataset

Examples

```
# Build FlowSom result
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
ff <- flowCore::read.FCS(fileName)
flowSOM.res <- FlowSOM(ff,
                        compensate = TRUE, transform = TRUE, scale = TRUE,
                        colsToUse = c(9, 12, 14:18),
                        nClus = 10)

# Map new data
outlier_report <- TestOutliers(flowSOM.res,
                              madAllowed = 5,
                              channels = flowSOM.res$map$colsUsed)

# Number of cells which is an outlier for x channels
outlier_on_multiple_markers <- table(rowSums(outlier_report$channel_specific != 0))
outlier_type <- paste(GetClusters(flowSOM.res),
                     apply(outlier_report$channel_specific, 1, paste0, collapse = ""))
outlier_counts <- table(grep(".*1.*", outlier_type, value = TRUE))
outliers_of_interest <- names(which(outlier_counts > 10))
outlier_boolean <- outlier_type %in% outliers_of_interest
```

UpdateFlowSOM

*UpdateFlowSOM***Description**

Update old FlowSOM object to a new one and checks if it is a flowSOM object

Usage

```
UpdateFlowSOM(fsom)
```

Arguments

fsom FlowSOM object, as generated by [BuildMST](#) or [FlowSOM](#)

Details

Determines whether or not the fsom input is of class "FlowSOM" and returns the FlowSOM object and metaclustering object inside fsom

Value

A FlowSOM object

See Also

[PlotFlowSOM](#)

UpdateMetaclusters	<i>UpdateMetaclusters</i>
--------------------	---------------------------

Description

Adapt the metacluster levels. Can be used to rename the metaclusters, split or merge existing metaclusters, add a metaclustering and/or reorder the levels of the metaclustering.

Usage

```
UpdateMetaclusters(
  fsom,
  newLabels = NULL,
  clusterAssignment = NULL,
  levelOrder = NULL
)
```

Arguments

<code>fsom</code>	Result of calling the FlowSOM function.
<code>newLabels</code>	Optional. Named vector, with the names the original metacluster names and the values the replacement. Can be used to rename or merge metaclusters.
<code>clusterAssignment</code>	Optional. Either a named vector, with the names the cluster numbers (characters) or a vector of length <code>NClusters(fsom)</code> . Can be used to assign clusters to existing or new metaclusters.
<code>levelOrder</code>	Optional. Vector showing the preferred order of the <code>fsom</code> metacluster levels.

Value

Updated FlowSOM object

Examples

```
fileName <- system.file("extdata", "68983.fcs", package = "FlowSOM")
ff <- flowCore::read.FCS(fileName)
ff <- flowCore::compensate(ff, flowCore::keyword(ff)[["SPILL"]])
ff <- flowCore::transform(ff,
  flowCore::transformList(colnames(flowCore::keyword(ff)[["SPILL"]]),
    flowCore::logicleTransform()))
flowSOM.res <- FlowSOM(ff,
  scale = TRUE,
  colsToUse = c(9, 12, 14:18),
  nClus = 10,
  seed = 1)

PlotStars(flowSOM.res, backgroundValues = flowSOM.res$metaclustering)
GetCounts(flowSOM.res)

# Merge MC8 and MC9
flowSOM.res <- UpdateMetaclusters(flowSOM.res, newLabels = c("8" = "8+9",
  "9" = "8+9"))
```

```

PlotStars(flowSOM.res, backgroundValues = flowSOM.res$metaclustering)
GetCounts(flowSOM.res)

# Split cluster 24 from metacluster 2 and order the metacluster levels
flowSOM.res <- UpdateMetaclusters(flowSOM.res,
                                clusterAssignment = c("24" = "debris?"),
                                levelOrder = c("debris?", as.character(c(1:7)),
                                                "8+9", "10"))
PlotStars(flowSOM.res, backgroundValues = flowSOM.res$metaclustering)
PlotNumbers(flowSOM.res, level = "metaclusters")

GetCounts(flowSOM.res)

```

UpdateNodeSize

*UpdateNodeSize***Description**

Update nodesize of FlowSOM object

Usage

```

UpdateNodeSize(
  fsom,
  count = NULL,
  reset = FALSE,
  transform = sqrt,
  maxNodeSize = 15,
  shift = 0,
  scale = NULL
)

```

Arguments

fsom	FlowSOM object, as generated by BuildMST
count	Absolute cell count of the sample
reset	Logical. If TRUE, all nodes get the same size
transform	Transformation function. Use e.g. square root to let counts correspond with area of node instead of radius
maxNodeSize	Maximum node size after rescaling. Default: 15
shift	Shift of the counts, defaults to 0
scale	Scaling of the counts, defaults to the maximum of the value minus the shift. With shift and scale set as default, the largest node will be maxNodeSize and an empty node will have size 0

Details

Add size property to the graph based on cellcount for each node

Value

Updated FlowSOM object

See Also

[BuildMST](#)

Examples

```
# Read from file, build self-organizing map and minimal spanning tree
fileName <- system.file("extdata", "68983.fcs", package="FlowSOM")
flowSOM.res <- ReadInput(fileName, compensate=TRUE, transform=TRUE,
                        scale=TRUE)
flowSOM.res <- BuildSOM(flowSOM.res, colsToUse=c(9,12,14:18))
flowSOM.res <- BuildMST(flowSOM.res)

# Give all nodes same size
PlotStars(flowSOM.res, equalNodeSize = TRUE)

# Node sizes relative to amount of cells assigned to the node
PlotStars(flowSOM.res)
```

%>%

Pipe operator

Description

See `magrittr::%>%` for details.

Usage

lhs %>% rhs

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