

# Package ‘MultiDataSet’

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**Type** Package

**Title** Implementation of the BRGE's (Bioinformatic Research Group in Epidemiology from Center for Research in Environmental Epidemiology) MultiDataSet and MethylationSet

**Version** 1.0.2

**Description** Implementation of the BRGE's (Bioinformatic Research Group in Epidemiology from Center for Research in Environmental Epidemiology) MultiDataSet and MethylationSet. MultiDataSet is designed for integrating multi omics data sets and MethylationSet to contain normalized methylation data. These package contains base classes for MEAL and rexposome packages.

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**LazyData** TRUE

**biocViews** Software, DataRepresentation

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add_eset	<i>Method to add an eSet to MultiDataSet.</i>
----------	---

---

### Description

This method adds or overwrites a slot of a `MultiDataSet` with the content of the given `eSet`.

### Usage

```
add_eset(object, set, dataset.type, dataset.name = NULL, warnings = TRUE,
         overwrite = FALSE, GRanges)
```

### Arguments

<code>object</code>	<code>MultiDataSet</code> that will be filled.
<code>set</code>	Object derived from <code>eSet</code> to be used to fill the slot.
<code>dataset.type</code>	Character with the type of data of the omic set (e.g. expression, methylation...)
<code>dataset.name</code>	Character with the specific name for this set (NULL by default). It is useful when there are several sets of the same type (e.g. multiple expression assays)
<code>warnings</code>	Logical to indicate if warnings will be displayed.
<code>overwrite</code>	Logical to indicate if the set stored in the slot will be overwritten.
<code>GRanges</code>	<code>GenomicRanges</code> to be included in <code>rowRanges</code> slot.

### Value

A new `MultiDataSet` with a slot filled.

### See Also

[add\\_methy](#), [add\\_genexp](#), [add\\_rnaseq](#), [add\\_snps](#)

## Examples

```
multi <- createMultiDataSet()
eset <- new("ExpressionSet", exprs = matrix(runif(10), 5))
multi <- add_eset(multi, eset, "exampledata", GRanges = NA)
```

---

add\_genexp

*Method to add an expression microarray dataset to MultiDataSet.*

---

## Description

This method adds or overwrites the slot "expression" of an MultiDataSet with the content of the given ExpressionSet.

## Usage

```
add_genexp(object, gexpSet, ...)
```

## Arguments

object	MultiDataSet that will be filled.
gexpSet	ExpressionSet to be used to fill the slot.
...	Arguments to be passed to add_eset.

## Value

A new MultiDataSet with the slot "expression" filled.

## Examples

```
multi <- createMultiDataSet()
eset <- new("ExpressionSet", exprs = matrix(runif(4), 2))
fData(eset) <- data.frame(chromosome = c("chr1", "chr2"), start = c(12414, 1234321),
  end = c(121241, 124124114), stringsAsFactors = FALSE)
multi <- add_genexp(multi, eset)
```

---

add\_methy                      *Method to add a slot of methylation to MultiDataSet.*

---

### Description

This method adds or overwrites the slot "methylation" of an MultiDataSet with the content of the given MethylationSet.

### Usage

```
add_methy(object, methySet, ...)
```

### Arguments

object	MultiDataSet that will be filled.
methySet	MethylationSet to be used to fill the slot.
...	Further arguments to be passed to add_eset.

### Value

A new MultiDataSet with the slot "methylation" filled.

### Examples

```
if (require(MEALData)){  
  multi <- createMultiDataSet()  
  betavals <- betavals[1:100, ] ## To speed up the example, the beta values are reduced  
  methy <- prepareMethylationSet(betavals, pheno)  
  multi <- add_methy(multi, methy)  
}
```

---

add\_rnaseq                      *Method to add an expression RNA seq dataset to MultiDataSet.*

---

### Description

This method adds or overwrites the slot "rnaseq" of an MultiDataSet with the content of the given ExpressionSet.

### Usage

```
add_rnaseq(object, rnaSet, ...)
```

**Arguments**

object	MultiDataSet that will be filled.
rnaSet	ExpressionSet to be used to fill the slot.
...	Arguments to be passed to add_eset.

**Value**

A new MultiDataSet with the slot "rnaseq" filled.

**Examples**

```
multi <- createMultiDataSet()
eset <- new("ExpressionSet", exprs = matrix(runif(4), 2))
fData(eset) <- data.frame(chromosome = c("chr1", "chr2"), start = c(12414, 1234321),
  end = c(121241, 12122414), stringsAsFactors = FALSE)
multi <- add_genexp(multi, eset)
```

---

 add\_rse

---

*Method to add a RangedSummarizedExperiment to MultiDataSet.*


---

**Description**

This method adds or overwrites a slot of a MultiDataSet with the content of the given RangedSummarizedExperiment.

**Usage**

```
add_rse(object, set, dataset.type, dataset.name = NULL, warnings = TRUE,
  overwrite = FALSE)
```

**Arguments**

object	MultiDataSet that will be filled.
set	Object derived from RangedSummarizedExperiment to be used to fill the slot.
dataset.type	Character with the type of data of the omic set (e.g. expression, methylation...)
dataset.name	Character with the specific name for this set (NULL by default). It is useful when there are several sets of the same type (e.g. multiple expression assays)
warnings	Logical to indicate if warnings will be displayed.
overwrite	Logical to indicate if the set stored in the slot will be overwritten.

**Value**

A new MultiDataSet with a slot filled.

**Examples**

```

if (require(GenomicRanges) & require(SummarizedExperiment)){
multi <- createMultiDataSet()
counts <- matrix(runif(200 * 6, 1, 1e4), 200)
rowRanges <- GRanges(rep(c("chr1", "chr2"), c(50, 150)),
                    IRanges(floor(runif(200, 1e5, 1e6)), width=100),
                    strand=sample(c("+", "-"), 200, TRUE),
                    feature_id=sprintf("ID%03d", 1:200))
colData <- DataFrame(Treatment=rep(c("ChIP", "Input"), 3),
                    row.names=LETTERS[1:6], id = LETTERS[1:6])
names(rowRanges) <- 1:200
rse <- SummarizedExperiment(assays=SimpleList(counts=counts),
                    rowRanges=rowRanges, colData=colData)
multi <- add_rse(multi, rse, "rseEx")
}

```

---

add\_snps

*Method to add a slot of SNPs to MultiDataSet.*


---

**Description**

This method adds or overwrites the slot "snps" of an MultiDataSet with the content of the given SnpSet.

**Usage**

```
add_snps(object, snpSet, ...)
```

**Arguments**

object	MultiDataSet that will be filled.
snpSet	SnpSet to be used to fill the slot.
...	Arguments to be passed to add_eset.

**Value**

A new MultiDataSet with the slot "snps" filled.

**Examples**

```

multi <- createMultiDataSet()
geno <- matrix(c(3,1,2,1), ncol = 2)
colnames(geno) <- c("VAL0156", "VAL0372")
rownames(geno) <- c("rs3115860", "SNP1-1628854")
map <- AnnotatedDataFrame(data.frame(chromosome = c("chr1", "chr2"), position = c(12414, 1234321),
                    stringsAsFactors = FALSE))
rownames(map) <- rownames(geno)
snpSet <- new("SnpSet", call = geno, featureData = map)

```

```
pheno <- data.frame(id = c("VAL0156", "VAL0372"))
rownames(pheno) <- c("VAL0156", "VAL0372")
pData(snpSet) <- pheno
multi <- add_snps(multi, snpSet)
```

---

checkProbes

*Filter MethylationSet probes*

---

### Description

This function selects probes present in the annotation matrix. Probes without annotation and annotation values without beta values are discarded.

### Usage

```
checkProbes(object)
```

### Arguments

object           MethylationSet

### Value

MethylationSet containing the common samples.

### Examples

```
if (require(MEALData)){
  betavals <- betavals[1:100, ] ## To speed up the example, the beta values are reduced
  methy <- prepareMethylationSet(betavals, pheno)
  checkProbes(methy)
}
```

---

checkSamples

*Modify a MethylationSet to only contain common samples*

---

### Description

This function removes samples that have beta values but no phenotypes and vice versa. If snps object is present, only samples present in the three set are retained.

### Usage

```
checkSamples(object)
```

**Arguments**

object           MethylationSet

**Value**

MethylationSet containing the common samples.

**Examples**

```
if (require(MEALData)){
  betavals <- betavals[1:100, ] ## To speed up the example, the beta values are reduced
  methy <- prepareMethylationSet(betavals, pheno)
  checkSamples(methy)
}
```

---

chrNumToChar           *Convert chr numbers to chr strings*

---

**Description**

Given a vector of number representing the chromosomes, convert them to string (e.g 1 to chr1). 23 is consider chrX, 24 is chrY, 25 is chrXY (probes shared between chromosomes X and Y) and 26 is chrMT.

**Usage**

```
chrNumToChar(vector)
```

**Arguments**

vector           The vector with the chromosome numbers

**Value**

A vector with the chromosomes in string format.

**Examples**

```
chromosomes <- c(1, 3, 4, 23, 15)
stringChrs <- chrNumToChar(chromosomes)
stringChrs
```

---

commonIds	<i>Get the name of the ids common to all datasets</i>
-----------	---

---

### Description

Get the name of the ids common to all datasets

### Usage

```
commonIds(object)
```

### Arguments

object            MultiDataSet that will be filtered.

### Value

Character vector with the common ids.

### Examples

```
multi <- createMultiDataSet()
eset <- new("ExpressionSet", exprs = matrix(runif(9), ncol = 3))
fData(eset) <- data.frame(chromosome = c("chr1", "chr1", "chr1"),
  start = c(1, 5, 10), end = c(4, 6, 14),
  stringsAsFactors = FALSE)
sampleNames(eset) <- c("S1", "S2", "S3")
pData(eset) <- data.frame(id = c("S1", "S2", "S3"))
rownames(pData(eset)) <- c("S1", "S2", "S3")
multi <- add_genexp(multi, eset, dataset.name = "g1")
eset <- new("ExpressionSet", exprs = matrix(runif(8), ncol = 2))
fData(eset) <- data.frame(chromosome = c("chr1", "chr1", "chr1", "chr1"),
  start = c(1, 14, 25, 104), end = c(11, 16, 28, 115),
  stringsAsFactors = FALSE)
sampleNames(eset) <- c("S1", "G2")
pData(eset) <- data.frame(id = c("S1", "G2"))
rownames(pData(eset)) <- c("S1", "G2")

multi <- add_genexp(multi, eset, dataset.name="g2")
commonIds(multi)
```

---

commonSamples	<i>Method to select samples that are present in all datasets.</i>
---------------	---

---

### Description

This method subsets the datasets to only contain the samples that are in all datasets.

### Usage

```
commonSamples(object)
```

### Arguments

object            MultiDataSet that will be filtered.

### Value

A new MultiDataSet with only the common samples.

### Examples

```
multi <- createMultiDataSet()
eset <- new("ExpressionSet", exprs = matrix(runif(9), ncol = 3))
fData(eset) <- data.frame(chromosome = c("chr1", "chr1", "chr1"),
  start = c(1, 5, 10), end = c(4, 6, 14),
  stringsAsFactors = FALSE)
sampleNames(eset) <- c("S1", "S2", "S3")
pData(eset) <- data.frame(id = c("S1", "S2", "S3"))
rownames(pData(eset)) <- c("S1", "S2", "S3")
multi <- add_genexp(multi, eset, dataset.name = "g1")
eset <- new("ExpressionSet", exprs = matrix(runif(8), ncol = 2))
fData(eset) <- data.frame(chromosome = c("chr1", "chr1", "chr1", "chr1"),
  start = c(1, 14, 25, 104), end = c(11, 16, 28, 115),
  stringsAsFactors = FALSE)
sampleNames(eset) <- c("S1", "G2")
pData(eset) <- data.frame(id = c("S1", "G2"))
rownames(pData(eset)) <- c("S1", "G2")

multi <- add_genexp(multi, eset, dataset.name="g2")
commonSamples(multi)
```

---

getMs	<i>Transforms beta values to M-values</i>
-------	---

---

**Description**

Given a MethylationSet or a AnalysisResults returns the matrix of M values using a logit2 transformation. Betas equal to 0 will be transformed to threshold and betas equal to 1, to 1 - threshold.

**Usage**

```
getMs(object, threshold = 1e-04)
```

**Arguments**

object	MethylationSet or AnalysisResults
threshold	Numeric with the threshold to avoid 0s and 1s.

**Value**

Matrix with the M values.

**Examples**

```
if (require(minfiData)){  
  set <- prepareMethylationSet(MsetEx[1:100, ], pData(MsetEx))  
  mvalues <- getMs(set)  
  head(mvalues)  
}
```

---

MethylationSet	<i>MethylationSet instances</i>
----------------	---------------------------------

---

**Description**

Container with the data needed to perform methylation analysis. MethylationSet inherits from eSet and contains meth matrix as assay data member.

**Usage**

```

methylationSet(betas, phenotypes, annotationDataFrame, annoString = "custom")

## S4 method for signature 'MethylationSet'
betas(object)

## S4 method for signature 'MethylationSet'
getMs(object, threshold = 1e-04)

## S4 method for signature 'MethylationSet'
checkProbes(object)

## S4 method for signature 'MethylationSet'
checkSamples(object)

```

**Arguments**

betas	Matrix of beta values
phenotypes	Data.frame or AnnotatedDataFrame with the phenotypes
annotationDataFrame	Data.frame or AnnotatedDataFrame with the phenotypes with the annotation of the methylation sites. A column with the chromosomes named chr and a column with the positions names pos are required.
annoString	Character with the name of the annotation used.
object	MethylationSet
threshold	Numeric with the threshold to avoid 0s and 1s.

**Details**

FeatureData, which contains annotation data, is required to perform any of the analysis.

**Value**

MethylationSet

**Methods (by generic)**

- betas: Get beta matrix
- getMs: Get Ms values
- checkProbes: Filter probes with annotation
- checkSamples: Modify a MethylationSet to only contain common samples

**Slots**

assayData Contains matrices with equal dimensions, and with column number equal to nrow(phenoData). assayData must contain a matrix meth with rows representing features (e.g., methylation probes sets) and columns representing samples.

phenoData See [eSet](#)

annotation See [eSet](#)

featureData See [eSet](#). fData should contain at least chromosome and positions columns.

### Examples

```
showClass("MethylationSet")
```

---

MultiDataSet

*MultiDataSet: Implementation of the BRGE's basic classes*

---

### Description

Implementation of the BRGE's (Bioinformatic Research Group in Epidemiology from Center for Research in Environmental Epidemiology) MultiDataSet and MethylationSet. MultiDataSet is designed for integrating multi omics data sets and MethylationSet to contain normalized methylation data. MultiDataSet for integrating multi omics data sets

### See Also

[MultiDataSet](#)

---

MultiDataSet-class

*MultiDataSet instances*

---

### Description

The class MultiDataSet is a superior class to store multiple datasets in form of triplets (assayData-phenoData-featureData). The datasets must be eSet or SummarizedExperiment.

### Usage

```
## S4 method for signature 'MultiDataSet,eSet'
add_ eset(object, set, dataset.type,
          dataset.name = NULL, warnings = TRUE, overwrite = FALSE, GRanges)
```

```
## S4 method for signature 'MultiDataSet,ExpressionSet'
add_genexp(object, gexpSet, ...)
```

```
## S4 method for signature 'MultiDataSet,ExpressionSet'
add_rnaseq(object, rnaSet, ...)
```

```
## S4 method for signature 'MultiDataSet,MethylationSet'
add_methy(object, methySet, ...)
```

```
## S4 method for signature 'MultiDataSet,RatioSet'  
add_methy(object, methySet, ...)  
  
## S4 method for signature 'MultiDataSet,RangedSummarizedExperiment'  
add_rse(object, set,  
        dataset.type, dataset.name = NULL, warnings = TRUE, overwrite = FALSE)  
  
## S4 method for signature 'MultiDataSet,SnpSet'  
add_snps(object, snpSet, ...)  
  
## S4 method for signature 'MultiDataSet'  
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## S4 method for signature 'MultiDataSet'  
pData(object)  
  
## S4 method for signature 'MultiDataSet'  
rowRanges(x)  
  
## S4 method for signature 'MultiDataSet,ANY,ANY'  
x[[i]]  
  
## S4 method for signature 'MultiDataSet,ANY,ANY,ANY'  
x[i, j, k, ..., drop = FALSE]
```

**Arguments**

object	MultiDataSet
set	Object derived from eSet to be used to fill the slot.
dataset.type	Character with the type of data of the omic set (e.g. expression, methylation...)
dataset.name	Character with the specific name for this set (NULL by default). It is useful when there
warnings	Logical to indicate if warnings will be displayed.
overwrite	Logical to indicate if the set stored in the slot will be overwritten.
GRanges	GenomicRanges to be included in rowRanges slot.
gexpSet	ExpressionSet to be used to fill the slot.
...	Further arguments passed to add_eset.
rnaSet	ExpressionSet to be used to fill the slot.
methySet	MethylationSet to be used to fill the slot.
snpSet	SnpSet to be used to fill the slot.
x	MultiDataSet
i	Character corresponding to selected sample names. They should match the id column of phenoData.
j	Character with the name of the selected tables.
k	GenomicRange used to filter the features.
drop	...

**Details**

The names of the three lists (assayData, phenoData and featureData) must be the same.

**Value**

MultiDataSet

**Methods (by generic)**

- `add_eset`: Method to add an eSet to MultiDataSet.
- `add_genexp`: Method to add a slot of expression to MultiDataSet.
- `add_rnaseq`: Method to add a slot of (RNASeq) expression to MultiDataSet.
- `add_methy`: Method to add a slot of methylation to MultiDataSet.
- `add_methy`: Method to add a slot of methylation to MultiDataSet.
- `add_rse`: Method to add a RangedSummarizedExperiment to MultiDataSet.
- `add_snps`: Method to add a slot of SNPs to MultiDataSet.
- `as.list`: Returns a list with the first matrix of each dataset.
- `assayData`: Retrieve all assay data blocks.
- `commonIds`: Get the name of the ids common to all datasets

- `commonSamples`: Get a MultiDataSet only with the samples present in all the tables
- `fData`: Retrieve information on features.
- `length`: Returns the number of sets into the object.
- `names`: Get the names of the slots.
- `sampleNames`: Get sample names
- `pData`: Retrieve information on experimental phenotypes.
- `rowRanges`: Retrieve information on feature ranges.
- `[[]`: Get a set from a slot
- `[`: Subset a MultiDataSet

### Slots

`assayData` List of assayData elements.

`phenoData` List of AnnotatedDataFrame containing the phenoData of each assayData.

`featureData` List of AnnotatedDataFrame containing the featureData of each assayData.

`rowRanges` List of GenomicRanges containing the rowRanges of each assayData.

`return_method` List of functions used to create the original object.

### See Also

[add\\_eset](#), [add\\_rse](#)

### Examples

```
createMultiDataSet()
```

---

`prepareMethylationSet` *Generating a MethylationSet*

---

### Description

This function creates a MethylationSet using from a matrix of beta values and a data.frame of phenotypes.

### Usage

```
prepareMethylationSet(matrix, phenotypes,
  annotation = "IlluminaHumanMethylation450kanno.ilmn12.hg19",
  chromosome = "chr", position = "pos", genes = "UCSC_RefGene_Name",
  group = "UCSC_RefGene_Group", filterNA_threshold = 0.05,
  verbose = FALSE)
```

**Arguments**

matrix	Data.frame or a matrix with samples on the columns and cpgs on the rows. A <code>minfi</code> object can be used to.
phenotypes	Data.frame or vector with the phenotypic features of the samples. Samples will be in the rows and variables in the columns. If matrix is a <code>minfi</code> object, phenotypes can be taken from it.
annotation	Character with the name of the annotation package or data.frame or Annotation-DataFrame with the annotation.
chromosome	Character with the column containing chromosome name in the annotation data.
position	chromosome Character with the column containing position coordinate in the annotation data.
genes	Character with the column containing gene names related to the methylation site in the annotation data. (Optional)
group	Character with the column containing the position of the probe related to the gene named in gene column. (Optional)
filterNA_threshold	Numeric with the maximum percentage of NA allowed for each of the probes. If 1, there will be no filtering, if 0 all probes containing at least a NA will be filtered.
verbose	Logical value. If TRUE, it writes out some messages indicating progress. If FALSE nothing should be printed.

**Details**

`prepareMethylationSet` is a useful wrapper to create `MethylationSet`. Right now, `prepareMethylationSet` supports two entry points: a `minfi` object and a matrix of betas.

Phenotypes are compulsory and can be supplied as data.frame or AnnotatedDataFrame.

By default, annotation is taken from `minfi` package and `IlluminaHumanMethylation450kanno.ilmn12.hg19` package is used, being the default arguments adapted to use this annotation. To use this annotation, `IlluminaHumanMethylation450kanno.ilmn12.hg19` must be installed and methylation sites must be named like in Illumina 450k chip. Use of this annotation ensures correct results in all the analysis.

If custom annotation is desired, there are two compulsory features: chromosomes and positions. Chromosomes should be supplied in the character form (e.g. chr1). Two additional features will be used during the presentation of results but not during the analyses: genes and group. Genes are the gene names of the genes around the cpg site and group defines the groups of the genes. Both columns will appear in the results but they are not used through the workflow. It should be noticed that `BlockFinder` only supports `minfi` annotation, so it is not advised to be used with custom annotation.

**Value**

`MethylationSet` with phenotypes and annotation.

**Examples**

```
if (require(minfiData)){  
  betas <- getBeta(MsetEx)[1:1000, ]  
  pheno <- pData(MsetEx)  
  set <- prepareMethylationSet(betas, pheno)  
}
```

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