# Package 'DMCFB'

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Type Package

**Title** Differentially Methylated Cytosines via a Bayesian Functional Approach

Version 1.2.0

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Description DMCFB is a pipeline for identifying differentially methylated cytosines using a Bayesian functional regression model in bisulfite sequencing data. By using a functional regression data model, it tries to capture position-specific, group-specific and other covariates-specific methylation patterns as well as spatial correlation patterns and unknown underlying models of methylation data. It is robust and flexible with respect to the true underlying models and inclusion of any covariates, and the missing values are imputed using spatial correlation between positions and samples. A Bayesian approach is adopted for estimation and inference in the proposed method.

**Depends** R (>= 3.6.0), SummarizedExperiment, methods, S4Vectors, BiocParallel, GenomicRanges, IRanges

**Imports** utils, stats, speedglm, MASS, data.table, splines, arm, rtracklayer, benchmarkme, tibble, matrixStats, fastDummies, graphics

Suggests testthat, knitr, rmarkdown

VignetteBuilder knitr

**biocViews** DifferentialMethylation, Sequencing, Coverage, Bayesian, Regression

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BugReports https://github.com/shokoohi/DMCFB/issues

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# Description

DMCFB is a profiling tool for identifying differentially methylated cytosines using Functional Bayesian Model in bisulfite sequencing data.

# $\mathsf{DMCFB}\ methods$

findDMCFB, plotDMCFB, cBSDMC, readBismark.

# ${\tt BSDMC}\ objects$

BSDMC-class

BSDMC-class	BSDMC object

# Description

The BSDMC object is an S4 class that represents differentially methylated CpG sites (DMCs) in BSSeq Data.

# Arguments

The matrix methReads contains the number of methylated reads spanning a CpG-site. The rows represent the CpG sites in rowRanges and the columns represent the samples in colData.
The matrix totalReads contains the number of reads spanning a CpG-site. The rows represent the CpG sites in rowRanges and the columns represent the samples in colData.
The matrix methLevels contains the predicted methylation level spanning a CpG-site using Bayesian functional regression models. The rows represent the CpG sites in rowRanges and the columns represent the samples in colData.

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#### Value

```
A BSDMC-class object
```

#### **Slots**

```
methReads An integer matrix
totalReads An integer matrix
methLevels A numeric matrix
```

#### Author(s)

Farhad Shokoohi <shokoohi@icloud.com>

#### See Also

 ${\tt RangedSummarizedExperiment-class} \ {\tt GRanges-class}$ 

#### **Examples**

```
nr <- 500
nc <- 16
metht <- matrix(as.integer(runif(nr * nc, 0, nr)), nr)
methc <- matrix(rbinom(n = nr * nc, c(metht), prob = runif(nr * nc)), nr, nc)
meths <- matrix(as.integer(runif(nr * nc, 0, 10)), nr)
methl <- methc / metht
methv <- matrix((runif(nr * nc, 0.1, 0.5)), nr)
r1 <- GRanges(rep("chr1", nr), IRanges(1:nr, width = 1), strand = "*")
names(r1) <- 1:nr
cd1 <- DataFrame(Group = rep(c("G1", "G2"), each = nc / 2),
    row.names = LETTERS[1:nc])
OBJ2 <- cBSDMC(
    rowRanges = r1, methReads = methc, totalReads = metht,
    methLevels = methl, methStates = meths, methVars = methv, colData = cd1
)
OBJ2</pre>
```

cBSDMC-method

cBSDMC method

# **Description**

Creates a BSDMC-class object

# Usage

```
cBSDMC(methReads, totalReads, methLevels, rowRanges,
  colData = DataFrame(row.names = colnames(methReads)),
  metadata = list(), ...)

## S4 method for signature 'matrix,matrix,matrix,GRanges'
cBSDMC(methReads, totalReads,
  methLevels, rowRanges, colData = DataFrame(row.names =
  colnames(methReads)), metadata = list(), ...)
```

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#### **Arguments**

methReads The matrix methReads contains the number of methylated reads spanning a CpG-site. The rows represent the CpG sites in rowRanges and the columns represent the samples in colData. The matrix totalReads contains the number of reads spanning a CpG-site. The totalReads rows represent the CpG sites in rowRanges and the columns represent the samples in colData. methLevels The matrix methLevels contains the predicted methylation level spanning a CpG-site using Bayesian functional regression models. The rows represent the CpG sites in rowRanges and the columns represent the samples in colData. A GRanges or GRangesList object describing the ranges of interest. Names, rowRanges if present, become the row names of the SummarizedExperiment object. The length of the GRanges or GRangesList must equal the number of rows of the matrices in assays. If rowRanges is missing, a SummarizedExperiment instance is returned. colData Object of class 'DataFrame' containing information on variable values of the samples metadata A list of storing MCMC samples or DMCs other possible parameters . . .

#### **Details**

The rows of a BSDMC object represent ranges (in genomic coordinates) of interest. The ranges of interest are described by a GRanges or a GRangesList object, accessible using the rowRanges function. The GRanges and GRangesList classes contains sequence (e.g., chromosome) name, genomic coordinates, and strand information. Each range can be annotated with additional data; this data might be used to describe the range or to summarize results (e.g., statistics of differential abundance) relevant to the range. Rows may or may not have row names; they often will not.

#### Value

A BSDMC-class

#### Author(s)

Farhad Shokoohi <shokoohi@icloud.com>

```
set.seed(1980)
nr <- 150
nc <- 8
metht <- matrix(as.integer(runif(nr * nc, 0, 100)), nr)
methc <- matrix(rbinom(n = nr * nc, c(metht), prob = runif(nr * nc)), nr, nc)
meths <- matrix(as.integer(runif(nr * nc, 0, 10)), nr)
methl <- methc / metht
methv <- matrix((runif(nr * nc, 0.1, 0.5)), nr)
r1 <- GRanges(rep("chr1", nr), IRanges(1:nr, width = 1), strand = "*")
names(r1) <- 1:nr
cd1 <- DataFrame(
   Group = rep(c("G1", "G2"), each = nc / 2),
   row.names = LETTERS[1:nc]</pre>
```

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```
)
OBJ2 <- cBSDMC(
  rowRanges = r1, methReads = methc, totalReads = metht,
  methLevels = methl, methStates = meths, methVars = methv, colData = cd1
)
OBJ2</pre>
```

combine-method

combine method

# Description

```
combine two BSDMC-class or two BSDMC-class
```

# Usage

```
combine(obj1, obj2)
## S4 method for signature 'BSDMC,BSDMC'
combine(obj1, obj2)
```

# **Arguments**

```
obj1 A BSDMC-class
obj2 A BSDMC-class
```

#### Value

A BSDMC-class or BSDMC-class

#### Author(s)

Farhad Shokoohi <shokoohi@icloud.com>

```
set.seed(1980)
nr <- 150
nc <- 8
metht <- matrix(as.integer(runif(nr * nc * 2, 0, nr)), nr)</pre>
methc <- matrix(</pre>
 rbinom(n = nr * nc, c(metht), prob = runif(nr * nc * 2)),
 nr, nc * 2
methl <- methc / metht</pre>
r1 <- GRanges(rep("chr1", nr), IRanges(1:nr, width = 1), strand = "*")
names(r1) <- 1:nr
cd1 <- DataFrame(Group = rep("G1", each = nc), row.names = LETTERS[1:nc])</pre>
OBJ1 <- cBSDMC(
 rowRanges = r1, methReads = methc[, 1:nc], totalReads = metht[, 1:nc],
 methLevels = methl[, 1:nc], colData = cd1
cd2 <- DataFrame(</pre>
  Group = rep("G2", each = nc),
```

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```
row.names = LETTERS[nc + 1:nc]
)

OBJ2 <- cBSDMC(
  rowRanges = r1, methReads = methc[, nc + 1:nc], totalReads =
    metht[, nc + 1:nc], methLevels = methl[, nc + 1:nc], colData = cd2
)

OBJ3 <- combine(OBJ1, OBJ2)
OBJ3</pre>
```

 $\verb|findDMCFB-method|$ 

findDMCFB method

# Description

DMC identification via Bayesian functional regression models

# Usage

```
findDMCFB(object, bwa, bwb, nBurn, nMC, nThin, alpha, sdv, nCores, pSize,
    sfiles)
## S4 method for signature 'BSDMC'
findDMCFB(object, bwa, bwb, nBurn, nMC, nThin, alpha,
    sdv, nCores, pSize, sfiles)
```

# **Arguments**

object	A BSDMC-class object
bwa	An integer value specifying the band-width size of B-spline basis matrix for a natural cubic spline for the group-specific effects of the Bayesian functional regression model
bwb	An integer value specifying the band-width size of B-spline basis matrix for a natural cubic spline for the individual-specific effects of the Bayesian functional regression model
nBurn	An integer value specifying the number of burn-in samples
nMC	An integer value specifying the number of MCMC samples after burn-in
nThin	An integer value specifying the thining number in MCMC
alpha	A numeric value specifying the level of $\alpha$ in credible interval $(1-\alpha)\%$
sdv	An double value specifying the standard deviation of priors
nCores	An integer value specifying the number of machine cores for parallel computing
pSize	An integer value specifying the number of cytosines in a regrion to be used in a Bayesian functiona regression model for DMC detection
sfiles	A logical value indicating whether files to be saved or not.

# Value

BSDMC-class object

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#### Author(s)

Farhad Shokoohi <shokoohi@icloud.com>

# **Examples**

```
set.seed(1980)
nr <- 1000
nc <- 4
metht <- matrix(as.integer(runif(nr * nc, 0, 100)), nr)</pre>
methc <- matrix(rbinom(n = nr * nc, c(metht), prob = runif(nr * nc)), nr, nc)</pre>
methl <- methc / metht</pre>
r1 <- GRanges(rep("chr1", nr), IRanges(1:nr, width = 1), strand = "*")
names(r1) \leftarrow 1:nr
cd1 <- DataFrame(</pre>
 Group = rep(c("G1", "G2"), each = nc / 2),
 row.names = LETTERS[1:nc]
OBJ1 <- cBSDMC(
  rowRanges = r1, methReads = methc, totalReads = metht,
 methLevels = methl, colData = cd1
OBJ2 <- findDMCFB(OBJ1,
 bwa = 10, bwb = 10, nBurn = 50, nMC = 50, nThin = 1,
 alpha = 0.05, nCores = 2, pSize = 500, sfiles = FALSE
)
OBJ2
```

methLevels-method

methLevels method

# **Description**

Returns methLevels stored in BSDMC-class

Assigns methLevels to BSDMC-class

#### Usage

```
methLevels(object)
methLevels(object) <- value

## S4 method for signature 'BSDMC'
methLevels(object)

## S4 replacement method for signature 'BSDMC,matrix'
methLevels(object) <- value</pre>
```

#### **Arguments**

```
object A BSDMC-class object value An integer matrix
```

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#### Value

A matrix

A BSDMC-class object

# Author(s)

Farhad Shokoohi <shokoohi@icloud.com>

# **Examples**

```
nr <- 150
nc <- 8
metht <- matrix(as.integer(runif(nr * nc, 0, 100)), nr)
methc <- matrix(rbinom(n = nr * nc, c(metht), prob = runif(nr * nc)), nr, nc)
methl <- methc / metht
r1 <- GRanges(rep("chr1", nr), IRanges(1:nr, width = 1), strand = "*")
names(r1) <- 1:nr
cd1 <- DataFrame(
    Group = rep(c("G1", "G2"), each = nc / 2),
    row.names = LETTERS[1:nc]
)
OBJ1 <- cBSDMC(
    rowRanges = r1, methReads = methc, totalReads = metht,
    methLevels = methl, colData = cd1
)
methLevels(OBJ1)
methLevels(OBJ1) <- methl</pre>
```

methReads-method

methReads method

# **Description**

Returns methReads stored in BSDMC-class

Assigns methReads to BSDMC-class

# Usage

```
methReads(object)
methReads(object) <- value

## S4 method for signature 'BSDMC'
methReads(object)

## S4 replacement method for signature 'BSDMC,matrix'
methReads(object) <- value</pre>
```

# **Arguments**

```
object A BSDMC-class object value An integer matrix
```

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#### Value

A matrix

A BSDMC-class object

#### Author(s)

Farhad Shokoohi <shokoohi@icloud.com>

#### **Examples**

```
nr <- 150
nc <- 8
metht <- matrix(as.integer(runif(nr * nc, 0, 100)), nr)</pre>
methc <- matrix(rbinom(n = nr * nc, c(metht), prob = runif(nr * nc)), nr, nc)</pre>
methl <- methc / metht</pre>
r1 <- GRanges(rep("chr1", nr), IRanges(1:nr, width = 1), strand = "*")
names(r1) \leftarrow 1:nr
cd1 <- DataFrame(</pre>
  Group = rep(c("G1", "G2"), each = nc / 2),
  row.names = LETTERS[1:nc]
OBJ1 <- cBSDMC(
  rowRanges = r1, methReads = methc, totalReads = metht,
  methLevels = methl, colData = cd1
methReads(OBJ1)
methReads(OBJ1) <- methc</pre>
```

params

params

#### **Description**

parameters name and their descriptions

#### **Arguments**

methReads The matrix methReads contains the number of methylated reads spanning a

CpG-site. The rows represent the CpG sites in rowRanges and the columns

represent the samples in colData.

totalReads The matrix totalReads contains the number of reads spanning a CpG-site. The

rows represent the CpG sites in rowRanges and the columns represent the sam-

ples in colData.

The matrix methLevels contains the predicted methylation level spanning a methLevels

CpG-site using Bayesian functional regression models. The rows represent the CpG sites in rowRanges and the columns represent the samples in colData.

A GRanges or GRangesList object describing the ranges of interest. Names, rowRanges

if present, become the row names of the SummarizedExperiment object. The length of the GRanges or GRangesList must equal the number of rows of the matrices in assays. If rowRanges is missing, a SummarizedExperiment in-

stance is returned.

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colData Object of class 'DataFrame' containing information on variable values of the

samples

metadata A list of storing MCMC samples or DMCs

object A BSDMC-class object

value An integer matrix
name A character list
obj1 A BSDMC-class
obj2 A BSDMC-class
files A character list

file

nCores An integer value specifying the number of machine cores for parallel computing

pSize An integer value specifying the number of cytosines in a regrion to be used in a

Bayesian functiona regression model for DMC detection

An integer value specifying the band-width size of B-spline basis matrix for

a natural cubic spline for the group-specific effects of the Bayesian functional

regression model

A character

bwb An integer value specifying the band-width size of B-spline basis matrix for a

natural cubic spline for the individual-specific effects of the Bayesian functional

regression model

nBurn An integer value specifying the number of burn-in samples
nThin An integer value specifying the thining number in MCMC

nMC An integer value specifying the number of MCMC samples after burn-in

sdv An double value specifying the standard deviation of priors

alpha A numeric value specifying the level of  $\alpha$  in credible interval  $(1-\alpha)\%$ 

col A character vector indicating which colors to alternate.

sfiles A logical value indicating whether files to be saved or not.

region An integer vector of length two specifying which subset of the object to be

plotted

nSplit A integer value specifying the number of subsets must be done for plotting the

results of DMC identification

parList A list specifying plots parameters, see par

... other possible parameters

#### Author(s)

Farhad Shokoohi <shokoohi@icloud.com>

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plotDMCFB method

#### **Description**

Plotting the results of DMC identifation stored in a BSDMC-class object

# Usage

```
plotDMCFB(object, region, nSplit, parList)
## S4 method for signature 'BSDMC'
plotDMCFB(object, region, nSplit, parList)
```

# **Arguments**

object A BSDMC-class object

region An integer vector of length two specifying which subset of the object to be plotted

nSplit A integer value specifying the number of subsets must be done for plotting the results of DMC identification

parList A list specifying plots parameters, see par

# Value

Plot

# Author(s)

Farhad Shokoohi <shokoohi@icloud.com>

```
set.seed(1980)
nr <- 1000
nc <- 4
metht <- matrix(as.integer(runif(nr * nc, 0, 100)), nr)</pre>
methc <- matrix(rbinom(n = nr * nc, c(metht), prob = runif(nr * nc)), nr, nc)</pre>
methl <- methc / metht</pre>
r1 <- GRanges(rep("chr1", nr), IRanges(1:nr, width = 1), strand = "*")
names(r1) \leftarrow 1:nr
cd1 <- DataFrame(</pre>
 Group = rep(c("G1", "G2"), each = nc / 2),
  row.names = LETTERS[1:nc]
OBJ1 <- cBSDMC(
  rowRanges = r1, methReads = methc, totalReads = metht,
 methLevels = methl, colData = cd1
OBJ2 <- findDMCFB(OBJ1,
  bwa = 10, bwb = 10, nBurn = 50, nMC = 50, nThin = 1,
  alpha = 0.05, nCores = 2, pSize = 500, sfiles = FALSE
```

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```
)
plotDMCFB(OBJ2)
```

readBismark-method

readBismark method

# **Description**

```
reads BS-Seq data
```

# Usage

```
readBismark(files, colData)
## S4 method for signature 'character,DataFrame'
readBismark(files, colData)
## S4 method for signature 'character,data.frame'
readBismark(files, colData)
## S4 method for signature 'character,character'
readBismark(files, colData)
```

# Arguments

files A character list

colData Object of class 'DataFrame' containing information on variable values of the

samples

#### Value

```
A BSDMC-class object
```

# Author(s)

Farhad Shokoohi <shokoohi@icloud.com>

```
fn <- list.files(system.file("extdata", package = "DMCHMM"))
fn.f <- list.files(system.file("extdata", package = "DMCHMM"),
   full.names = TRUE
)
OBJ <- readBismark(fn.f, fn)
cdOBJ <- DataFrame(Cell = factor(c("BC", "TC", "Mono"),
   labels = c("BC", "TC", "Mono")
), row.names = c("BCU1568", "BCU173", "BCU551"))
colData(OBJ) <- cdOBJ
OBJ</pre>
```

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totalReads-method

totalReads method

# **Description**

Returns totalReads stored in BSDMC-class Assigns totalReads to BSDMC-class

# Usage

```
totalReads(object)
totalReads(object) <- value

## S4 method for signature 'BSDMC'
totalReads(object)

## S4 replacement method for signature 'BSDMC,matrix'
totalReads(object) <- value</pre>
```

#### **Arguments**

object A BSDMC-class object value An integer matrix

#### Value

A matrix

A BSDMC-class object

# Author(s)

Farhad Shokoohi <shokoohi@icloud.com>

```
nr <- 150
nc <- 8
metht <- matrix(as.integer(runif(nr * nc, 0, 100)), nr)
methc <- matrix(rbinom(n = nr * nc, c(metht), prob = runif(nr * nc)), nr, nc)
methl <- methc / metht
r1 <- GRanges(rep("chr1", nr), IRanges(1:nr, width = 1), strand = "*")
names(r1) <- 1:nr
cd1 <- DataFrame(
    Group = rep(c("G1", "G2"), each = nc / 2),
    row.names = LETTERS[1:nc]
)
OBJ1 <- cBSDMC(
    rowRanges = r1, methReads = methc, totalReads = metht,
    methLevels = methl, colData = cd1
)
totalReads(OBJ1)
totalReads(OBJ1) <- metht</pre>
```

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