Package 'RforProteomics'

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Type Package

Title Companion package to the 'Using R and Bioconductor for proteomics data analysis' publication

Version 1.24.0

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Depends MSnbase (>= 2.5.3)

Imports R.utils, shiny, biocViews, BiocManager

Suggests AnnotationDbi, rpx, DT, knitr, rmarkdown, BiocStyle, mzR, xcms, msdata, isobar, MALDIquant (>= 1.12), MALDIquantForeign, readBrukerFlexData, rTANDEM, synapter, synapterdata, IPPD, Rdisop, OrgMassSpecR, SummarizedExperiment, BRAIN, rols, hpar, GO.db, org.Hs.eg.db, e1071, biomaRt, RColorBrewer, ggplot2, reshape2, xtable, lattice, mzID, pRoloc, pRolocdata, MSGFplus, MSnID, msmsTests, msmsEDA, DEP, corrplot, beanplot, Heatplus, gplots, VennDiagram, protViz, genefilter, plotly, gridExtra, dplyr, lubridate

Enhances cleaver

Description This package contains code to illustrate the 'Using R and Bioconductor for proteomics data analysis' and 'Visualisation of proteomics data using R and Bioconductor' manuscripts. The vignettes describe the code and data needed to reproduce the examples and figures described in the paper and functionality for proteomics visualisation. It also contain various function to discover R software for mass spectrometry and proteomics.

URL http://lgatto.github.com/RforProteomics/

biocViews ExperimentData, MassSpectrometryData, ReproducibleResearch

License Artistic-2.0

VignetteBuilder knitr

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R topics documented:

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downloadData   *Download a file*

Description

Unless already present, downloads `src` in the `destdir` directory.

Usage

```
downloadData(src, destdir = ".", unpack = TRUE, ...)
```

Arguments

src	The url of the file to download.
destdir	The destination directory. Default is `"."`.
unpack	Should `src` be uncompressed? Default is `TRUE`.
...	Additional paramters passed to [download.file]().

Value

Invisible returns the full path of the downloaded file.

Author(s)

Laurent Gatto

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getPackagesInBiocView   *Packages in a biocView*

Description

Finds the package names that have a specific biocView.

Usage

```
getPackagesInBiocView(view, rep = c("BioCsoft", "BioCann", "BioCexp",
  "BioCextra"), biocVersion)
```

Arguments

view   The biocView of interest. For example "Proteomics".

rep   Repository of interest. One of "BioCsoft", "BioCann", "BioCexp" or '"Bio-
                Cextra"1.

biocVersion   A character with the Bioconductor version of interest. For example "2.14".

Value

An instance of class BiocView. NULL if the the biocView was not found.

Author(s)

Laurent Gatto

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getPXD000001mzData   *Download the PXD000001 mzTab file*

Description

Unless already present, downloads the PXD000001 mzData file in the destdir directory. The
resulting file is named PRIDE_Exp_Complete_Ac_22134.xml

Usage

```
getPXD000001mzData(destdir = ".")
```

Arguments

destdir   A character with the destination folder.

Value

Invisibly returns the name of the downloaded file.

Author(s)

Laurent Gatto

getPXD000001mzTab   *Download the PXD000001 mzTab file*

Description

Unless already present, downloads the PXD000001 mzTab file in the `destdir` directory. The resulting file is named `F063721.dat-mztab.txt`.

Usage

```
getPXD000001mzTab(destdir = ".")
```

Arguments

destdir   A character with the destination folder.

Value

Invisibly returns the name of the downloaded file.

Author(s)

Laurent Gatto

getPXD000001mzXML   *Download the PXD000001 mzXML file*

Description

Unless already present, downloads the PXD000001 mzXML file in the `destdir` directory. The resulting file is named `TMT_Erwinia_1uLSike_Top10HCD_isol2_45stepped_60min_01.mzXML`.

Usage

```
getPXD000001mzXML(destdir = ".")
```

Arguments

destdir   A character with the destination folder.

Value

Invisibly returns the name of the downloaded file.

Author(s)

Laurent Gatto

getThermoHelaPRTC *Dowload Thermo Hela PRTC data*

Description

Downloads on of multiple Thermo Hela/PRTC data files.

Usage

```
getThermoHelaPRTC(src, destdir = ".")
```

Arguments

src   The name of the file to be downloaded. If missing, a vector of possible filenames
            is returned. If "all", all files are downloaded. Alternatively, a pattern can be
            used to grep the files from the output getThermoHelaPRTC() the files to be
            downloaded.

destdir   Destination directory. Default is ".".

Value

Invisibly return the path of the downloaded files.

Author(s)

Laurent Gatto

See Also

downloadData

Examples

```
## Not run:
getThermoHelaPRTC()
getThermoHelaPRTC("design")
getThermoHelaPRTC("all")

## End(Not run)
```

id *An* mzIdentML *file*

Description

This file has been generated by searching the raw mzXML file of the ProteomeXchange PXD000001
data set against the erwinia_carotovora.fasta using the MSGF+ search engine: java -jar
~/bin/MSGFPlus.20140630/MSGFPlus.jar -s TMT_Erwinia_1uLSike_Top10HCD_isol2_45stepped_60min_01.mzX
-d erwinia_carotovora.fasta -inst 1 -protocol 2

Examples

```
## source files to repeat the search
library("rpx")
px <- PXDataset("PXD000001")
pxfiles(px)

f <- dir(system.file("extdata", package = "RforProteomics"),
         pattern = "mzid", full.names=TRUE)
library("mzID")
id <- mzID(f)
id
```

---

new_ions   *Create, analyse and detect ions*

Description

This is the constructor function to generate a set of ions that can later be analysed with 'analyse()'
and detected with 'detect()'.

Usage

```
new_ions(npeaks = 10, mzrange = c(100, 1000), nimg = 100)

analyse(x, sleep = 0.1)

analyze(x, sleep = 0.1)

detect(x, new = FALSE)

spectrum(x, ...)
```

Arguments

npeaks	A 'numeric' scalar defining the number of unique peaks (M/Z values). Default is 10.
mzrange	A 'numeric' of length 2 defining the range of possible M/Z values. Default is 'c(100, 1000)'.
nimg	A 'numeric' scalar. When analysing the ions, their separation along their M/Z values will be split along a sequence of length 'nimg'. Default is 100.
x	An object of class 'ions'.
sleep	How much time to wait before producing the next plot.
new	A 'logical' scalar, indicating if the separated ions (last frame of calling 'analyse) should be plotting, or whether the detection should be overlaid. Default is 'FALSE', to add the plot on top of the opened device.
...	Additional arguments passed to [graphics::plot()].

Value

An object of class 'ions'.

'analyse', 'detect' and 'spectrum' are used for their side effect or producing plots. They all invisibly return 'NULL'.

Author(s)

Laurent Gatto

Examples

```
set.seed(1L)
x <- new_ions(nimg = 5)
x
analyse(x)
detect(x)
spectrum(x)
```

---

packageDF   *Package descriptions*

Description

Format a `BiocView` as a `data.frame`.

Usage

```
packageDF(x, nsub = TRUE, version = TRUE)
```

Arguments

x	An instance of class `BiocView`, as produced by `getPackagesInBiocView`.
nsub	A `logical` indicating `"\n"` are to be replaced by a space.
version	A `logical` specifying if the package version should be added.

Value

A `data.frame` with package information.

Author(s)

Laurent Gatto

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proteomicsPackages   *Proteomics and MS biocView packages*

Description

Searches for all the packages with the ″Proteomics″ (software), ″MassSpectrometry″ (software) and ″MassSepctrometryData″ (data) packages and return their names, titles and versions as a data.frame. The (unexported but documented) underlying functions are RforProteomics:::getPackagesInBiocView (to find relevant package) and RforProteomics:::packageDF (data.frame formatting).

Usage

```
proteomicsPackages(biocv, cache = FALSE)

massSpectrometryPackages(biocv, cache = FALSE)

massSpectrometryDataPackages(biocv, cache = FALSE)
```

Arguments

biocv	A character with the Bioconductor version to search for relevant packages. If missing, the running version is used.
cache	A logical indicating whether cached package information should be used. Default is FALSE. All except development versions are up-to-date.

Value

A data.frame with the respective package names, titles and versions.

Author(s)

Laurent Gatto

Examples

```
head(pp <- proteomicsPackages("3.0"))
ppc <- proteomicsPackages("3.0", cache = TRUE)
all.equal(pp, ppc)
```

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qnt   *PXD000001 example MSnSet*

Description

In this TMT 6-plex experiment, four exogenous proteins were spiked into an equimolar *Erwinia carotovora* lysate with varying proportions in each channel of quantitation; yeast enolase (ENO) at 10:5:2.5:1:2.5:10, bovine serum albumin (BSA) at 1:2.5:5:10:5:1, rabbit glycogen phosphorylase (PHO) at 2:2:2:2:1:1 and bovin cytochrome C (CYT) at 1:1:1:1:1:2. Proteins were then digested, differentially labelled with TMT reagents, fractionated by reverse phase nanoflow UPLC (nanoACQUITY, Waters), and analysed on an LTQ Orbitrap Velos mass spectrometer (Thermo Scientific). Files in multiple format will be used to illustrate the input/output capabilities that are available to the proteomics audience. The companion package provides dedicated functions to directly download the data.

The data has been downloaded from the ProteomeXchange repository and imported into R as illustrated in the example. It is of class `MSnSet`. See also the `MSnbase-demo` vignette for more details.

Usage

```
data("qnt")
```

Format

An instance of class `MSnSet`

References

Laurent Gatto (2014). RforProteomcs: Companion package to the 'Using R and Bioconductor for proteomics data analysis' publicationR package version 1.3.1.

Gatto L, Christoforou A. Using R and Bioconductor for proteomics data analysis. Biochim Biophys Acta. 2013 May 18. doi:pii: S1570-9639(13)00186-6. 10.1016/j.bbapap.2013.04.032. [Epub ahead of print] PubMed PMID: 23692960.

Examples

```
## Not run:
    library("rpx")
    px1 <- PXDataset("PXD000001")
    mztab <- pxget(px1, "PXD000001_mztab.txt")
    library("MSnbase")
    qnt <- readMzTabData(mztab, what = "PEP")
    sampleNames(qnt) <- reporterNames(TMT6)
    qnt$conditions <- factor(c("A", "A", "B", "B", "B", "A"))
    qnt <- filterNA(qnt)

    selA <- qnt$conditions == "A"

    fData(qnt)$log2FC <-
        log(rowMeans(exprs(qnt)[, selA]), 2) -
            log(rowMeans(exprs(qnt)[, !selA]), 2)
    fData(qnt)$baseMean <- log(rowMeans(exprs(qnt)), 10)

## End(Not run)

library("RforProteomics")
library("MSnbase")
data(qnt)
```

```
class(qnt)
head(exprs(qnt))
head(fData(qnt))
```

RforProteomics *Opens the package vignettes*

Description

Opens the package and visualisation vignettes.

Usage

```
RforProteomics()

RProtVis()

RProtViz()
```

Value

An instance of class vignette. Used for its side effect, opening a vignette.

Author(s)

Laurent Gatto

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shinyMA *MA and expression plots in* shiny

Description

Starts an interactive shiny application that displays an MA plot and an expression plot side by side. The user can select features of interest on the MAplot and the respective intensities are displayed on the expression plot on the right.

The data has been prepared using the mzTab file from the ProteomeXchange spiked-in data PXD000001 (see qnt for details). Sample 1, 2, 6 and 3, 4, 5 have been arbitrarily chosen to define two groups.

Usage

```
shinyMA()
```

Value

Used for its side effects of starting a shiny application.

Author(s)

Laurent Gatto <lg390@cam.ac.uk>

References

The application is an adaptation of Michael Love's `shinyMA` app available on https://github.com/mikelove/shinyMA.

Examples

```
if (interactive())
    shinyMA()
```

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